

# FUNCTION OF LATENT TRANSFORMING GROWTH FACTOR-β BINDING PROTEIN-2 (LTBP-2) IN ELASTINOGENESIS AND MODULATION OF GROWTH FACTOR STORAGE, EXPRESSION AND ACTIVITY IN NORMAL AND FIBROTIC TISSUES

## **Mohamed Arshad Mohamed Sideek**

## Discipline of Anatomy and Pathology School of Medicine The University of Adelaide

### **April 2016**

A thesis submitted in fulfilment of the requirement for the Degree of

Doctor of Philosophy in Medicine and Surgery

## Table of Contents

Summary	8
Declaration	10
Acknowledgements	11
Publication Arising During PhD Candidature	12
Scientific Communications	13
Awards and Achievements During Candidature	15
Abbreviations	16
CHAPTER 1: The Role of LTBP-2 in Elastinogenesis and Fibrotic Diseases	19
1.1 The extracellular matrix (ECM) and its components	19
1.2 Collagens	21
1.3 Proteoglycans	23
1.3.1 Heparin/Heparan sulphate proteoglycans	24
1.4 Glycoprotein	26
1.5 Elastic fibres	28
1.5.1 Elastin	28
1.5.2 Fibrillin-microfibrils	30
1.6 Fibulins	32
1.6.1 Fibulin-5	34
1.7 Fibrotic diseases	37
1.8 Growth factors	39
1.8.1 Transforming growth factor – beta (TGF-β)	39
1.8.2 Fibroblast growth factor -2 (FGF-2)	42
1.9 Latent TGF-β binding proteins (LTBPs)	44
1.9.1 LTBP-2	45
1.10 Aims of the present study	50
1.10.1 Specific aims	50
1.10.2 Hypotheses	50
1.11 References	51
CHAPTER 2: LTBP-2 competes with tropoelastin for binding to fibulin-5 and hep-	arin, and is
a negative modulator of elastinogenesis	
2.1 Abstract	80
2.2 Introduction	81

2.3 Experimental procedures	83
2.3.1 Materials	83
2.3.2 Molecular binding assays	83
2.3.3 Confocal microscopy	84
2.4 Results and discussion	86
2.4.1 LTBP-2 and tropoelastin have similar binding affinities for fibulin-5.	86
2.4.2 LTBP-2 can completely block the interaction of tropoelastin with fibu	ılin-586
2.4.3 Heparin shows only minor inhibition of fibulin-5 interaction with trop LTBP-2.	
2.4.4 LTBP-2 substantially inhibits the interaction of tropoelastin with hepa	arin87
2.4.5 Co-localization of LTBP-2 with fibrillin-1, fibulin-5 and heparan sulf proteoglycans in fibroblast matrix.	
2.4.6 Exogenous LTBP-2 blocks elastinogenesis by cultured chondrocytes.	88
2.5 Acknowledgments	91
2.6 References	91
2.7 Figures	96
Figure 2.1 LTBP-2 and tropoelastin bind fibulin-5 with similar affinities	96
Figure 2.2 Inhibition of the fibulin-5- tropoelastin interaction by LTBP-2 at terminal fragment (LTBP-2 NT(H))	
Figure 2.3 Heparin shows minor but specific inhibition of LTBP-2 interactifibulin-5	
Figure 2.4 LTBP-2 inhibits the interaction of tropoelastin with heparin in a dependent manner.	
Figure 2.5 Confocal co-localisation of LTBP-2 with fibulin-5, fibrillin-1, fi heparan sulfate proteoglycans in fibroblast matrix	
Figure 2.6 Exogenous LTBP-2 inhibits elastinogenesis by ear cartilage cho culture.	•
Figure 2.7 LTBP-2 and elastinogenesis	108
2.8 Mini-research paper: LTBP-2 inhibits elastin and fibrillin assembly in mabovine ear cartilage chondrocytes	
2.8.1 Introduction	111
2.8.2 Method	111
2.8.3 Results and Discussion	112
2.8.4 References	116

CHAPTER 3: LTBP-2 has a single high-affinity binding site for FGF-2 and blocks FG induced cell proliferation.	
3.1 Abstract	
3.2 Introduction	
3.3 Materials and Methods.	
3.3.1 Recombinant protein production and purification	
3.3.2 Solid Phase Binding Assays	
3.3.3 Cell proliferation assay	
3.3.4 Detection and quantitation of FGFR1 activation	
3.3.5 Immunohistochemistry	
3.4 Results and Discussion	
3.4.1 FGF-2 has a strong affinity for LTBP-2	
3.4.2 FGF-2 binding is confined to a small central region of the LTBP-2 molecul	
3.4.3 The FGF-2 binding site is close to a heparin-binding region of LTBP-2	
3.4.4 LTBP-2 blocks FGF-2-induced cell proliferation	
3.4.5 LTBP-2 and FGF-2 show similar distributions in fibrotic skin	
3.5 References	137
3.6 Figures	142
Figure 3.1 Recombinant LTBP-2 fragments	142
Figure 3.2 LTBP-2 specifically binds FGF-2 but not VEGF, BMP-4, BMP-7 or beta	
Figure 3.3 LTBP-2 interacts strongly with FGF-2	144
Figure 3.4 FGF-2 has a single binding domain in the central region of LTBP-2	145
Figure 3.5 The FGF-2 binding site is close to the central heparin binding site on	
Figure 3.6 LTBP-2 blocks FGF-2-induced cell proliferation	
Figure 3.7 LTBP-2 and fibrillin-1 colocalize in fibrotic skin	
Figure 3.8 LTBP-2 and FGF-2 co-localize in keloid tissue	
Figure 3.9 Quantitation of LTBP-2 and FGF-2 in normal skin and keloid	
CHAPTER 4: Co-localization of LTBP-2 with FGF-2 in fibrotic human keloid and	132
hypertrophic scarhypertrophic scar	153
4.1 Abstract	
4.2 Introduction	159
4.3 Materials and Methods	162

4.3.1 Human tissue samples	162
4.3.2 Antibodies	162
4.3.3 Immunohistochemistry	162
4.3.4 Quantitative Polymerase Chain Reaction	164
4.3.5 Statistical analysis	165
4.4 Results	166
4.4.1 LTBP-2 and FGF-2 do not co-localize in fibroblast cell culture	166
4.4.2 LTBP-2 and FGF-2 show extensive co-localizations in fibrotic skin lesions	166
4.4.3 LTBP-2 shows similar distribution with elastin and fibrillin-1 in fibrotic skin	167
4.4.4 Both LTBP-2 and FGF-2 mRNA expression levels are greatly elevated in hypertrophic scar	168
4.5 Discussion	
4.6 References	
4.7 Figures	
Figure 4.1 Immunofluorescence staining for LTBP-2 and FGF-2 in human foreskin fibroblast (HFF) cultures	
Figure 4.2 LTBP-2 and FGF-2 co-localize in fibrotic skin lesions	180
Figure 4.3 Quantitative immunofluorescence analysis of LTBP-2 and FGF-2 in norm skin, mature scar, hypertrophic scar and keloid tissue	
Figure 4.4 LTBP-2 co-localizes with fibrillin-1 and elastin in fibrotic skin	183
Figure 4.5 The expression levels of mRNA for LTBP-2, FGF-2 and TGF-β in fibrot skin tissues	
4.8 Supplementary figures	185
Figure 4.6 Specificity of LTBP-2 and FGF-2 antibodies.	185
Figure 4.7 Specificity of secondary antibodies and laser channels.	186
CHAPTER 5: LTBP-2 stimulates the expression of TGF-β in human fibroblasts via Akt α p38 MAPK signalling pathways	
5.1 Abstract	192
5.2 Introduction	193
5.3 Materials and Methods	196
5.3.1 Reagents	196
5.3.2 Expression and purification of recombinant LTBP-2 and fragments	196
5.3.3 Cell culture and treatment	196
5.3.4 Detection and quantitation of TGF-β	197

5.3.5 Real-time PCR	198
5.3.6 Measurement of signal phosphorylation	198
5.3.7 Statistical analysis	199
5.4 Results	200
5.4.1 Exogenous LTBP-2 stimulates expression and secretion of TGF-β1 in MSU fibroblasts	
5.4.2 LTBP-2 stimulation of TGF-β synthesis and secretion is dose-dependent	201
5.4.3 Time course for LTBP-2 stimulation of TGF-β upregulation	202
5.4.4 A short exposure to exogenous LTBP-2 is sufficient to stimulate TGF-β upregulation	202
5.4.5 The TGF-β stimulating activity maps to a central region of LTBP-2 consists an eight -cys motif flanked by pairs of EGF-like repeats.	_
5.4.6 Induction of Akt and p38 phosphorylation by LTBP-2	204
5.4.7 LTBP-2 stimulates the expression of TGF-β via Akt and p38 MAPK signal pathways	_
5.4.8 Blocking of integrin $\alpha V\beta 3$ receptors partially attenuates TGF- $\beta$ production LTBP-2	•
5.5 Discussion	207
5.6 References	213
Figure 5.1 Exogenous LTBP-2 increases TGF-β in conditioned medium which is independent of extracellular matrix.	
Figure 5.2 LTBP-2 upregulates TGF-β expression in MSU 1.1 cells	221
Figure 5.3 LTBP-2 upregulates TGF-β in a dose-dependent manner	222
Figure 5.4 Time course for LTBP-2 stimulation of TGF-β production	223
Figure 5.5 A short incubation of MSU 1.1 cells with LTBP-2 is sufficient to upre TGF-β expression and secretion.	_
Figure 5.6 A central region of LTBP-2 consisting of an 8-cys motif flanked by pa EGF-like repeats (fragment LTBP-2C F3) contains the stimulatory activity	
Figure 5.7 Exogenous LTBP-2 stimulates phosphorylation of AKT and p38 MAI human fibroblasts	
Figure 5.8 LTBP-2 stimulation of TGF-β upregulation involves Akt and p38 MA signalling pathways.	
Figure 5.9 Blocking of integrin αVβ3 receptors partially attenuates TGF-β produ induced by LTBP-2	
Figure 5.10. Schematic representation of possible signalling pathways involved in production of TGF-β1 by LTBP-2	

CHAPTER 6: Discussion	230
6.1 LTBP-2 plays an important role in elastinogenesis and elastic fibre assembly	230
6.2 Role of LTBP-2 in the regulation of growth factor activities	233
6.3 Co-localization of LTBP-2 and FGF-2 in fibrotic human tissues	234
6.4 LTBP-2 stimulates the expression and secretion of TGF-β	236
6.5 The domain organization of the human LTBP-2 protein	243
6.6 References	245
CHAPTER 7: Conclusions and Future Directions	258
7.1 Conclusion	258
7.2 Future directions	258

## **Summary**

LTBP-2 is tightly associated with fibrillin microfibrils and elastic fibres in a range of tissues mainly in the lung, heart, skeletal muscle, placenta, liver and the aorta. LTBP-2 belongs to the fibrillin-LTBP superfamily of extracellular matrix proteins. Unlike other LTBPs, LTBP-2 does not covalently bind TGF-beta and its molecular function remains unclear. LTBP-2 complexes with fibulin-5, an elastin-chaperone protein critical for normal elastic fibre assembly, and it has been suggested that LTBP-2 may preferentially direct fibulin-5-elastin globules onto fibrillin-1 (rather than fibrillin-2) microfibrils during elastinogenesis. However, we have now shown that LTBP-2 inhibits rather than enhances the interaction of tropoelastin with fibulin-5 in vitro. In addition LTBP-2 inhibited elastic fibre assembly in ear cartilage chondrocyte cultures largely at the stage of elastin deposition onto the fibrillin microfibril scaffold. In parallel experiments, LTBP-2 was shown to significantly inhibit the binding of heparin to tropoelastin suggesting LTBP-2 may compete with tropoelastin for binding to certain cell surface HSPGs and contribute to controlling the release of elastin microassemblies from the cell surface. Confocal microscopy showed strong co-distribution of LTBP-2 with fibulin-5 and fibrillin-1 and partial co-distribution with HSPGs, perlecan and syndecan-4 in fibroblast matrix Thus it is evident that LTBP-2 is a negative modulator of elastinogenesis and that LTBP-2 levels may regulate the rate and extent of elastinogenesis in some tissues.

A recent study has linked LTBP-2 gene mutations to recessive form of Weill-Marchesani syndrome which is characterised by short stature, thick fibrotic skin and ectopia lentis. Since fibrillin-1 mutations can also cause this syndrome it is now clear that LTBP-2 is linked to fibrillin biology, growth factor regulation and fibrosis. To investigate growth factor binding to LTBP-2, our laboratory screened a number of cytokines involved in the pathogenesis of fibrotic disorders and identified a very strong specific interaction of FGF-2. The activity was confined to a central region of the LTBP-2 consisting of 6 EGF-like repeats, suggesting a single binding sequence. The finding presented in this thesis found that 5-fold molar excess LTBP-2 can completely block FGF-2 stimulation of fibroblast proliferation via its receptor. In addition increased levels and extensive co-localisation of LTBP-2 and FGF-2 were observed and quantitated in human hypertrophic scars and keloids. Furthermore, qPCR confirmed consistent elevation of LTBP-2 and FGF-2 expression in samples of these fibrotic tissues. The results

support the concept that increased LTBP-2 expression in fibrotic disorders may increase FGF-2 binding and reduce FGF-2 activity, inhibiting normal repair processes.

Previously we have shown that LTBP-2 competes with LTBP-1 for binding to fibrillin *in vitro*, suggesting that LTBP-2 may modulate TGF- $\beta$  storage and activation. In experiments designed to measure displacement of TGF- $\beta$  complexes from fibrillin microfibrils, our laboratory discovered addition of LTBP-2, or a small bioactive fragment LTBP-2C F3 to MSU 1.1 skin fibroblasts resulted in a large increase in TGF- $\beta$  levels in culture medium. However the increase in TGF- $\beta$  the medium was cycloheximide sensitive indicating elevated cellular expression and secretion of TGF- $\beta$  rather than release of matrix-stored TGF- $\beta$ . Exogenous LTBP-2 or fragment F3 significantly increased levels of latent TGF- $\beta$  in the medium after 9h peaking at 15h. The signalling mechanism appears to involve the PI3K/Akt and p38 MAPK pathways, as incubation of cells with LTBP-2 (10µg/ml) elevated Akt 1/2/3 Ser473 and P38 D-8 phosphorylation and inhibition of each pathway completely blocked the synthesis of TGF- $\beta$ . Investigation of the cell surface receptor for the bioactive fragment of LTBP-2 was less informative. Inhibitory antibody to  $\beta$ 1 integrins did not affect the TGF- $\beta$  upregulation but it was partially inhibited by an antibody to the integrin  $\alpha$ V $\beta$ 3 receptor, suggesting it may be involved in LTBP-2-cell interaction(s) resulting in elevated TGF- $\beta$  expression.

In conclusion, these findings are consistent with LTBP-2 having novel regulatory functions in elastinogenesis, growth factor modulation and fibrosis which may lead to novel therapy development for fibrotic diseases and tissue repair.

## **Declaration**

I certify that this work contains no material which has been accepted for the award of any other degree or diploma in my name in any university or other tertiary institution and, to the best of my knowledge and belief, contains no material previously published or written by another person, except where due references has been made in the text. In addition, I certify that no part of this work will, in the future, be used in a submission in my name for any other degree or diploma in any university or other tertiary institution without the prior approval of the University of Adelaide and where applicable, any partner institution responsible for the joint award of this degree.

I give consent to this copy of my thesis when deposited in the University Library, being made available for loan and photocopying, subject to the provisions of the Copyright Act 1968.

The author acknowledges that copyright of published works contained within this thesis resides with the copyright holder(s) of those works.

I also give permission for the digital version of my thesis to be made available on the web, via the University's digital research repository, the Library Search and also through web search engines, unless permission has been granted by the University to restrict access for a period of time.

Beyona	tnis, i	ao not	wish to	o prace	any	restriction	n on	access	to	tnis	tnesis.

(Mohamed Arshad Mohamed Sideek)

1154273 The University of Adelaide

Date: 15/04/2016

## Acknowledgements

I am very grateful to a lot of people for their support throughout my Ph.D journey and the production of this thesis. The highest gratitude to my primary supervisor, **Dr. Mark Gibson** for his constant guidance and encouragement. Thank you for being patience and understanding, and for standing by my side when times get hard. Thank you for teaching me that every mistake is just a learning experience. I am forever indebted to the best supervision over the last four years. I really appreciate it from the bottom of my heart. I would like to thank my co-supervisor, **Prof. Allison Cowin**, and his research team especially **Dr. Zlatko Kopecki**, for the opportunity to work in their reputable laboratory. I thank the fantastic members of the Gibson's lab, **Mahroo Parsi**, **Clementine Menz**, and **Josh Smith** for their help and advice throughout my Ph.D work.

I wholeheartedly thank the greatest gift I have ever had, my mother, Fazila Begum (Amma) for her warm hugs, pleasing smiles and encouraging words, and for all the countless times she has been there for me. To my wonderful dad, Mohamed Sideek (Atta), I could not begin to list all the ways his love has made all the difference in my life. Amma and Atta, I am blessed to have both of you in my life, I love you so much! To my one and only brother and sister, Jamal Mohideen and Nur Saminah, I cannot thank you enough for their amazing support, unconditional love and care. I could not imagine my life without you.

My deepest appreciation belongs to my wife and my soul mate, **Noor Shafiqa**, who has always been my strength. I do not know how I can ever thank you for being such a loving and caring person. Thank you to other family members, **Abdul Hadi** (grandfather), **Mumtaj Begum** (grandmother), **Haji Mohamed** (father-in-law), **Fathimunnisa** (mother-in-law), **Mohamed Zahirudin** (brother-in-law), **Thilsath Yasmine** (sister-in-law), **Imthiyaz** (brother), **Rosmah akka** (sister), **Mohamed Hussain** (uncle), **Jalifah Nachiya** (aunty), **Ruknudin mama** (uncle), **Sajeetha mami** (aunty) and **Dato' Haji Mohamed Mustafa** (uncle). Without your persistent love and dedication over the past years, none of this would have been possible.

Special thanks to **Iqbal Jamaludin** and **Azuwan Musa**, who have been an awesome friends and colleagues. Thank you for always being a good listener and making me laugh when I had almost forgotten how to do so.

## **Publication Arising During PhD Candidature**

#### **CHAPTER 2**

LTBP-2 competes with tropoelastin for binding to fibulin-5 and heparin, and is a negative modulator of elastinogenesis

**Mohamed A. Sideek**, Clementine Menz, Mahroo K. Parsi, Mark A. Gibson *Matrix Biology 34 (2014) 114-123(Impact factor: 5.074)* 

LTBP-2 inhibits elastin and fibrillin assembly in matrix of fetal bovine ear cartilage chondrocytes

Mohamed A. Sideek and Mark A. Gibson

(manuscript in preparation)

#### **CHAPTER 3**

LTBP-2 has a single high-affinity binding site for FGF-2 and blocks FGF-2-induced cell proliferation

Clementine Menz, Mahroo K. Parsi, Julian R.J. Adams, **Mohamed A. Sideek**, Zlatko Kopecki, Allison J. Cowin, Mark A. Gibson

PLOS ONE 10(8): e013557 (Impact factor: 3.234)

#### **CHAPTER 4**

Co-localization of LTBP-2 with FGF-2 in fibrotic human keloid and hypertrophic scar **Mohamed A. Sideek**, Abdulrahman Teia, Zlatko Kopecki, Allison J. Cowin, Mark A. Gibson

Journal of Molecular Histology (Impact factor: 1.815)

#### **CHAPTER 5**

LTBP-2 stimulates the expression of TGF- $\beta$  via Akt & p38 MAPK signalling pathway in human fibroblast

Mohamed A. Sideek, Josh Smith, Clementine Menz, Julian R.J. Adams, Allison J. Cowin, Mark A. Gibson

*Matrix Biology (Impact factor: 5.074)* (manuscript in preparation)

## **Scientific Communications**

#### **INTERNATIONAL:**

#### <u>2011</u>

The Elastin and Elastic Fibers Gordon Research Conference, University of New England, Biddeford, ME, United States (oral and poster)

#### **NATIONAL AND LOCAL:**

#### <u>2015</u>

Australian Society of Medical Research (ASMR) South Australia Annual Scientific Meeting, National Wine Centre, Adelaide, Australia (poster)

#### **2014**

Matrix Biology Society of Australia and New Zealand (MBSANZ) 38<sup>th</sup> Annual Scientific Meeting, Queenscliff, Victoria, Australia (oral and poster)

Florey International Postgraduate Research Conference, Faculty of Health Sciences (FHS) Postgraduate Research Conference 2014, The University of Adelaide, Adelaide, Australia (poster)

Australian Society of Medical Research (ASMR) South Australia Annual Scientific Meeting, Adelaide Convention Center, Adelaide, Australia (poster)

#### <u>2013</u>

Matrix Biology Society of Australia and New Zealand (MBSANZ) 37<sup>th</sup> Annual Scientific Meeting, McCracken Country Club, South Australia, Australia (poster)

Faculty of Health Sciences (FHS) Postgraduate Research Conference 2013, Adelaide, Australia (poster)

Australian Society of Medical Research (ASMR) South Australia Annual Scientific Meeting, Adelaide Convention Center, Adelaide, Australia (poster)

1<sup>st</sup> Malaysian Postgraduate Student Symposium of South Australia 2013, Adelaide, Australia (oral and poster)

#### <u>2012</u>

Matrix Biology Society of Australia and New Zealand (MBSANZ) 36<sup>th</sup> Annual Scientific Meeting, Mantra Legends, Gold Coast, Queensland, Australia (poster)

Postgraduate Research Expo, Faculty of Health Science, University of Adelaide, The National Wine Centre, Adelaide, South Australia, Australia (poster)

Australian Society of Medical Research (ASMR) South Australia Annual Scientific Meeting, Adelaide Convention Center, Adelaide, Australia (poster)

School of Medical Sciences, The University of Adelaide. PhD Introductory Seminar (oral)

## **Awards and Achievements During Candidature**

#### <u>2014</u>

Shortlisted for the Dennis Lowther Award at Matrix Biology Society of Australia and New Zealand 38<sup>th</sup> Annual Scientific Meeting, Queenscliff, Victoria, Australia.

School of Medical Sciences Postgraduate Travel Award (National Conference Attendance), Faculty of Health Sciences (FHS), The University of Adelaide, Australia.

#### <u>2013</u>

Shortlisted for the Dennis Lowther Award at Matrix Biology Society of Australia and New Zealand 37<sup>th</sup> Annual Scientific Meeting, Victor Harbour, South Australia, Australia.

Academic and Character Excellence Award 2013 (Anugerah Kecemerlangan Akademik dan Sahsiah (AKASIA)). Australian-Malaysian Muslim Solidarity of South Australia (ISMA SA), South Australia, Australia.

#### 2012

Shortlisted for the Dennis Lowther Award at Matrix Biology Society of Australia and New Zealand 36<sup>th</sup> Annual Scientific Meeting, Mantra Legends, Gold Coast, Queensland, Australia.

IIUM Scholarship (Full), Kulliyyah of Allied Health Sciences, International Islamic University Malaysia (IIUM), Pahang, Malaysia.

SLAB/SLAI (PhD) Scholarship (Full), Ministry of Education (MoE), Malaysia.

## **Abbreviations**

BCIP-	5-bromo-4-chloro-3-indolylphosphate toluidine salt
BMP-	bone morphogenetic protein
BSA-	bovine serum albumin
C-	carboxy-terminus
C-6-S-	chondroitin-6-sulphate
Ca <sup>2+</sup> -	calcium ions
CaCl <sub>2</sub> -	calcium chloride
cbEGF-	calcium binding epidermal growth factor
cDNA-	complementary deoxyribonucleic acid
CHX-	cycloheximide
CS-	chondroitin sulphate
DAPI-	4',6-Diamidino-2-phenylindole dihydrochloride
ddH <sub>2</sub> 0-	double distilled water
DMEM-	Dulbecco's Modification of Eagles Medium
DMSO-	dimethyl sulphoxide
DNA-	deoxyribonucleic acid
DTT-	dithiothreitol
ECM-	extracellular matrix
EDTA-	ethylene diamine tetraacetic acid
EGF-	epidermal growth factor
ELISA-	enzyme-linked immuno-sorbent assay
EMILIN-	elastinmicrofibril interface located protein
FCS-	foetal calf serum
FGF-2	fibroblast growth factor -2
FGFRs	FGF receptors
FITC-	fluorescein isothiocyanate
GAG-	glycosaminoglycan
GuHCL-	guanidine hydrochloride
HAC-	heparin-albumin conjugates
HEK-	human embryo kidney
HFF-	human foreskin fibroblasts
his <sub>6</sub> -	6-histidine
HS-	heparan sulphate
HSPGs-	heparan sulphate proteoglycans
HTS-	hypertrophic scar
IgG-	Immunoglobulin G
IL-	interleukin
$K_d$ -	dissociation constant

kDa-	kiloDalton
LAP-	latency-associated protein
LLC-	large latent complex
LTBP-	latent transforming growth factor-β binding protein
LTBP-2C (H)-	LTBP-2 C-terminal
LTBP-2NT (H)-	LTBP-2 N-terminal
M-	molar
MAGP-	microfibrillar-associated glycoprotein
MAPK-	mitogen-activated protein kinases
MFS-	Marfan syndrome
min-	minutes
mM-	millimolar
MMP-	matrix metalloprotease
mRNA-	messenger RNA
N-	amino-terminus
NaCl-	sodium chloride
NBCS-	new born calf serum
NBT-	nitro-blue tetrazolium chloride
NEAA-	non-essential amino acids
ng-	nanogram
Ni-	nickel
NRS-	normal rabbit serum
PBS-	phosphate-buffered saline
PCR-	polymerase chain reactions
PG-	proteoglycan
PVDF-	polyvinylidene difluoride
r-	recombinant
RGD-	arginine-glycine-aspartic acid motif
RNA-	ribonucleic acid
RT-	room temperature
SD-	standard deviation
SDS-PAGE-	sodium sulphate polyacrylamide gel electrophoresis
SLC-	small latent comlex
SMA-	smooth muscle actin
SMC-	smooth muscle cell
TBS-	tris buffered saline
TGF-β-	transforming growth factor-β
TIMP-	tissue inhibitor of metalloproteinases

TMB-	tetramethylbenzidine
v or vol-	volume
W-	weight
WMS-	Weill-Marchesani syndrome
α-	alpha
αVβ3-	alpha V beta 3
αVβ5-	alpha V beta 5
β-	beta
μl-	microliter
Δ	delta
8-cys-	8-cysteine containing motif