SUBMITTED VERSION

Georgina M. Sylvia, Erik P. Schartner, Hanna J. McLennan, Avishkar Saini, Kylie R. Dunning, Malcolm S. Purdey, Andrew D. Abell, Jeremy G. Thompson **High precision pH measurements in biological environments using a portable optical fibre pH sensor** Biophotopics Australasia 2019, 2019 / Goldys, E. Gibson, B. (ed /s), vol 11202

Biophotonics Australasia 2019, 2019 / Goldys, E., Gibson, B. (ed./s), vol.11202, pp.112020F-1-112020F-2

© 2019 SPIE

Originally published at: http://dx.doi.org/10.1117/12.2541300

PERMISSIONS

https://spie.org/conferences-and-exhibitions/authors-and-presenters/copyright-formrequired-for-publication?SSO=1

SPIE Web Posting Policy for papers, posters, and presentation recordings published in SPIE Proceedings and SPIE Journals

SPIE grants to authors (and their employers) of papers, posters, and presentation recordings published in SPIE Proceedings or SPIE Journals on the SPIE Digital Library the right to post an author-prepared version or an official version (preferred version) of the published paper, poster, or presentation recording on an internal or external repository controlled exclusively by the author/employer, or the entity funding the research, provided that (a) such posting is noncommercial in nature and the paper, poster, or presentation recording is made available to users without charge; (b) an appropriate copyright notice and citation appear with the paper, poster, or presentation recording; and (c) a link to SPIE's official online version of the paper, poster, or presentation recording is provided using the DOI (Document Object Identifier) link.

This authorization does not extend to third-party web sites not owned and maintained by the author/employer such as ResearchGate, Academia.edu, YouTube, etc.

SPIE content published under a Creative Commons CC-BY license is exempt from the above requirements.

14 December 2020

High precision pH measurements in biological environments using a portable optical fibre pH sensor

Georgina M. Sylvia^{*a,b}, Erik P. Schartner^{a,b}, Hanna J. McLennan^{a,c}, Avishkar Saini^{a,c}, Kylie R. Dunning^{a,c}, Malcolm S. Purdey^a, Andrew D. Abell^{a,b}, Jeremy G. Thompson^{a,c}

^aARC Centre of Excellence for Nanoscale Biophotonics (CNBP); ^bInstitute for Photonics and Advanced

Sensing (IPAS); ^cRobinson Research Institute, Adelaide Medical School;

University of Adelaide, Adelaide SA 5005, Australia

ABSTRACT

We have demonstrated that an optical fibre-based pH sensor can be utilised to accurately assess pH in a biological environment. Initial measurements were performed on 5 μ L drops of culture medium containing individual female mouse reproductive cells (cumulus-oocyte-complexes, COCs), with the goal of obtaining a biomarker of individual cell health during assisted reproductive processes. Improvements to the measurement procedure were found to reduce fluorescence signal variability, enabling improved measurement precision compared to previous studies. Results show the application of treatments which serve to increase lactic acid production by the COC, and thus induce an acidification of the local microenvironment, are detectable by the pH sensor. This optical technology presents a promising platform for the measurement of pH and the detection of other extracellular biomarkers to assess cell health during assisted reproduction.

Keywords: pH, fluorescence, sensing, optical fibre, cumulus-oocyte-complex (COC), microenvironment, lactic acid

INTRODUCTION

Optical fibre-based fluorescent probes have found use in a range of applications, where the fibre geometry (~100-200 μ m in diameter) provides access to microvolumes of analyte for non-invasive chemical detection in spatially-hindered biological environments¹. We recently reported a pH-sensitive optical fibre probe, referred to here as optical fibre probe 1 (**OFP1**), and its measurement of extracellular surface pH in excised human breast cancer tissue samples². The functionalised surface of this probe contains the pH sensitive fluorophore 5(-6)-carboxynapthofluorescein (CNF) embedded within an acrylamide polymer coating on the tip of a 200 μ m diameter multimodal optical fibre³. **OFP1** successfully detected differences in extracellular acidity, which enabled the margins between healthy and cancerous breast tissue to be discerned.

One promising application for fibre probes is in biosensing, where the local environment makes measurement with conventional pH probes extremely challenging, such as in assisted reproductive technologies, where the limited size of the culture medium drop presents an ideal opportunity to employ **OFP1**. Here we present a new application for **OFP1** measuring the pH in 5 μ L drops of culture medium containing unfertilised eggs (oocytes) with their surrounding support cells (the cumulus), which is collectively known as the cumulus-oocyte-complex (COC). Treatments applied to the COC and known to result in the production of lactic acid should decrease the pH of the local COC microenvironment⁴⁻⁶.

RESULTS AND DISCUSSION

2.1 OFP1 Pre-bleaching

Initial repeated scans 5 s apart of freshly prepared **OFP1** probes revealed that the CNF response signal exponentially decays over time (Figure 1a), suggesting that a potential method to reduce the impact of photobleaching is to "pre bleach" the probes with 800-100 scans prior to performing biological measurements. After this pre-bleaching process, the decay during experimental measurements is of a smaller magnitude (see Figure 1b); therefore, any observed changes in fluorescence signal after this point may be attributed to meaningful pH changes in the medium being measured.

2.2 In vitro Measurements

OFP1 was used to assess the local pH immediately adjacent to the COC, with the probe positioned as shown in Fig. 1c. Trials measured culture medium pH following chemical (CoCl₂) or hormonal (FSH) treatment of the cells. COCs treated with 100 μ M CoCl₂ (chemical treatment) showed a statistically lower calculated pH than untreated COCs, with **OFP1** measuring a clearly distinguishable 0.06 unit drop in pH after treatment.

*georgina.sylvia@adelaide.edu.au; phone (+61) 8 8313 6327; https://researchers.adelaide.edu.au/profile/georgina.sylvia

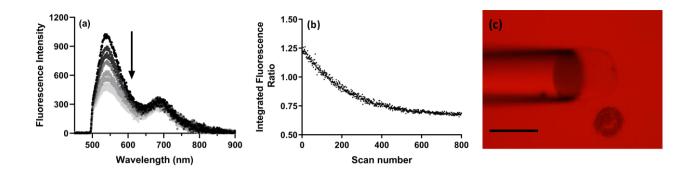


Figure 1. a) Reduction in overall fluorescence signal and b) fluorescence signal ratio change during the CNF pre-bleaching process. c) Position of **OFP1** relative to a COC during measurement. Scale bar represents 200 µm.

Hormonal stimulation of COCs was next explored in order to investigate whether a biological event can trigger a measurable change in pH like the one caused by CoCl₂ exposure. In all replicates, FSH stimulation resulted in 0.02-0.03 mean pH unit decrease compared to untreated COCs. As FSH increases lactic acid production by increasing the metabolic activity of cumulus cells⁴, the difference detected here by **OFP1** is a direct result of a biological event.

This work presents the first instance of local pH being used as an indirect measure of individual COC metabolism *in vitro*, and confirms that **OFP1** is a suitable technology to detect biological signals in the microenvironment surrounding the COC.

ACKNOWLEDGEMENTS

The authors acknowledge funding support from the ARC (CE140100003, LP110200736, LP150100657). H.J.M. and A.S. are supported through the Australian Government Research Training Program (RTP). J.G.T. is supported by a National Health & Medical Research Council (NHMRC) Research Fellowship (ID 1077694). The University of Adelaide, Adelaide Microscopy (AM) and Microscopy Australia. The OptoFab node of the Australian National Fabrication Facility (ANFF) utilising Commonwealth and South Australian State Government funding. The authors acknowledge Suliman Yagoub for assistance with the micromanipulators.

REFERENCES

- Wang, X. -d., Wolfbeis, O. S., "Fiber-Optic Chemical Sensors and Biosensors (2013-2015)," Anal. Chem. 88(1), 203-227 (2016).
- [2] Schartner, E. P., Henderson, M. R., Purdey, M. S., Dhatrak, D., Monro, T. M., Gill, G. P., Callen, D. F., "Cancer Detection in Human Tissue Samples Using a Fiber-Tip pH Probe," Cancer Research 76(23), 6795–801 (2016).
- [3] Song, A., Parus, S., Kopelman, R., "High-performance fiber-optic pH microsensors for practical physiological measurements using a dual-emission sensitive dye," Anal. Chem. 69, 863-867 (1997).
- [4] Sutton-McDowall, M. L., Gilchrist, R. B., Thompson, J. G., "Cumulus expansion and glucose utilisation by bovine cumulus-oocyte complexes during in vitro maturation: the influence of glucosamine and follicle-stimulating hormone," Reproduction 128, 313-319 (2004).
- [5] Roberts, R., Stark, J., Iatropoulou, A., Becker, D. L., Frank, S., Hardy, K., "Energy substrate metabolism of mouse cumulus-oocyte complexes: response to follicle-stimulating hormone is mediated by the phosphatidylinositol 3kinase pathway and is associated with oocyte maturation," Biol. Reprod. 71(1), 199-209 (2004).