

**Physiological studies of whey
protein in older people, with a
focus on feeding behaviour,
nutrition, obesity, type 2 diabetes
mellitus and post-prandial
blood pressure**

A thesis by
Avneet Kaur Oberoi BSc, MSc

For the Degree of
Doctor of Philosophy

**Faculty of Health and Medical Sciences
Adelaide Medical School
The University of Adelaide
October 2022**

Jis ke sir upar tu swami so dukh kaisa pave

O Lord, one who is under your protection, one who considers you to be above himself

He will not experience any suffering in life

Guru Granth Sahib, 473

TABLE OF CONTENTS

TABLE OF CONTENTS.....	I
THESIS ABSTRACT.....	VIII
DECLARATION.....	XIII
ACKNOWLEDGEMENTS.....	XIV
PUBLICATIONS ARISING FROM THIS THESIS.....	XIX
ABSTRACTS AND PRESENTATIONS.....	XXI
CHAPTER 1: INTRODUCTION.....	23
1.1 The rising burden of ageing in the population of Australia and other developed countries.....	25
1.2 Nutritional problems and adverse body composition changes associated with human ageing.....	25
1.2.1 Undernutrition and sarcopaenia.....	25
1.2.1.1 Nutritional management of under-nutrition and sarcopaenia.....	28
1.2.2 Whey protein.....	30
1.2.3 Obesity in older people.....	34
1.2.4 Questions about effect of whey on nutritional factors addressed in this thesis.....	36
1.3 Effect of whey protein ingestion on glucose metabolism.....	37
1.3.1 Type 2 diabetes mellitus (T2D).....	37
1.3.1.1 Type 2 diabetes mellitus: Demographics and impact.....	37
1.3.1.2 Management of T2D: Focus on dietary measures.....	37
1.3.2 Questions about effect of whey on glucose metabolism addressed in this thesis.....	41
1.4 Effects of whey on blood pressure and pulse rate in older people.....	41
1.4.1 Physiological cardiovascular responses to eating.....	41
1.4.2 Postprandial hypotension.....	42
1.4.3 Questions about effect of whey on blood pressure and pulse rate addressed in this thesis.....	46
1.5 Gastric emptying.....	46
1.5.1 Control of gastric emptying.....	46
1.5.2 Measurement of gastric emptying rate.....	48
CHAPTER 2: RATIONAL USE OF PROTEIN SUPPLEMENTS IN THE ELDERLY- RELEVANCE OF GASTROINTESTINAL MECHANISMS.....	51
STATEMENT OF AUTHORSHIP.....	52
2.1 Abstract.....	54
2.2 Introduction.....	55

2.2.1 Ageing, weight loss and undernutrition	55
2.2.2 Ageing and sarcopenia	56
2.3 Prevention and management of under-nutrition and sarcopenia in older people	56
2.3.1 Nutritional measures	57
2.3.2 Nutritional supplements	57
2.3.3 Protein nutritional supplements	58
2.4 Dietary protein requirements in older people	58
2.4.1 Do men and women have different dietary protein needs?	60
2.4.2 Summary recommendations for dietary protein intake in older people	60
2.4.2.1 Total Daily Protein Intake	60
2.4.3 How many older people need supplements to reach the recommended protein intake?	61
2.4.4 Possible adverse effects of protein supplements	61
2.4.4.1 Renal effects	62
2.4.4.2 Bone effects	62
2.4.4.3 Post-prandial hypotension	63
2.4.5 Evidence for benefits of nutritional supplements in older people	64
2.4.6 Does the type of protein or amino acid in the supplement matter?	65
2.4.7 How much protein should there be in the supplement?	66
2.4.8 Timing of protein intake relative to exercise	67
2.5 Gastrointestinal responses to protein ingestion: Effects of ageing	67
2.5.1 Effect of ageing on appetite and feeding responses to whey protein	68
2.5.1.1 Intra-duodenal whey	68
2.5.1.2 Oral whey	69
2.5.2 What is the best timing of the protein supplement use by older people? ..	71
2.5.3 Effect of ageing on gastric function and emptying	71
2.5.3.1 Intra-gastric food distribution and antral area	72
2.5.3.2 Gastric emptying	72
2.5.3.3 Small intestinal satiety mechanisms	74
2.5.4 Selected hormones	75
2.5.4.1 Cholecystokinin (CCK)	75
2.5.4.2 Glucagon-like peptide 1 (GLP-1)	76
2.5.4.3 Gastric inhibitory peptide (GIP)	77
2.5.4.4 Insulin and glucose	77
2.5.4.5 Glucagon	78
2.5.4.6 Ghrelin	78

2.6 Future directions	78
2.7 Conclusions.....	79
2.8 Overall aims and hypothesis	81
CHAPTER 3: WHEY PROTEIN DRINK INGESTION BEFORE BREAKFAST SUPPRESSED ENERGY INTAKE AT BREAKFAST AND LUNCH, BUT NOT DURING DINNER, AND WAS LESS SUPPRESSED IN HEALTHY OLDER THAN YOUNGER MEN	83
STATEMENT OF AUTHORSHIP.....	84
3.1 Abstract.....	87
3.2 Introduction.....	88
3.3 Materials and methods.....	89
3.3.1 Participants	89
3.3.2 Protocol	90
3.3.3 Measurements.....	95
3.3.3.1 Energy intake.....	95
3.3.3.2 Antral area	95
3.3.3.3 Perceptions of appetite and palatability	96
3.4 Data and statistical analysis	97
3.5 Results	97
3.5.1 Energy intake.....	97
3.5.2 Protein intake.....	102
3.5.3 Gastric emptying.....	103
3.5.4 Appetite.....	104
3.5.5 Palatability of drinks and meals	106
3.6 Discussion.....	107
3.7 Conclusions.....	110
CHAPTER 4: ACUTE EFFECTS OF WHEY PROTEIN ON ENERGY INTAKE, APPETITE AND GASTRIC EMPTYING IN YOUNGER AND OLDER, OBESE MEN	111
STATEMENT OF AUTHORSHIP	112
4.1 Abstract.....	115
4.2 Introduction.....	117
4.3 Materials and methods	119
4.3.1 Protocol	120
4.3.2 Measurements.....	121
4.3.2.1 Energy intake.....	121
4.3.2.2 Gastric emptying.....	122
4.3.2.3 Perceptions of appetite and gastrointestinal symptoms	123

4.3.2.4 Blood glucose	123
4.4 Data and statistical analysis	123
4.5 Results	124
4.5.1 Energy intake	124
4.5.2 Gastric emptying.....	125
4.5.3 Blood glucose concentrations.....	126
4.5.4 Perceptions of appetite and gastrointestinal symptoms	127
4.5.4.1 Baseline	127
4.5.4.2 After study drink.....	128
4.6 Discussion.....	132
CHAPTER 5: COMPARATIVE EFFECTS OF CO-INGESTING WHEY PROTEIN AND GLUCOSE ALONE AND COMBINED ON BLOOD GLUCOSE, PLASMA INSULIN AND GLUCAGON CONCENTRATIONS IN YOUNGER AND OLDER MEN	137
STATEMENT OF AUTHORSHIP	138
5.1 Abstract.....	141
5.2 Introduction.....	142
5.3 Materials and methods.....	143
5.3.1 Participants	143
5.3.2 Protocol	145
5.3.3 Measurements.....	146
5.3.3.1 Blood glucose and plasma insulin and glucagon concentrations	146
5.3.3.2 Gastric emptying.....	147
5.3.3.3 Energy intake.....	147
5.3.3.4 Perceptions of appetite	148
5.4 Data and statistical analysis	148
5.5 Results	150
5.5.1 Blood glucose	150
5.5.1.1 Interaction effects	150
5.5.1.2 Drink-condition effects	150
5.5.1.3 Age effects.....	151
5.5.2 Plasma insulin.....	155
5.5.2.1 Interaction effects	155
5.5.2.2 Drink-condition effects	155
5.5.2.3 Age effects.....	155
5.5.3 Plasma glucagon.....	156
5.5.3.1 Interaction effects	156

5.5.3.2 Drink-condition effects	156
5.5.3.3 Age effects.....	157
5.5.4 Gastric emptying.....	159
5.5.5 Energy intake.....	160
5.5.6 Perceptions of appetite.....	161
5.6 Discussion.....	162
5.7 Conclusions.....	169
CHAPTER 6: EFFECTS OF CO-INGESTING GLUCOSE AND WHEY PROTEIN ON BLOOD GLUCOSE, PLASMA INSULIN AND GLUCAGON CONCENTRATIONS, AND GASTRIC EMPTYING, IN OLDER MEN WITH AND WITHOUT TYPE 2 DIABETES.....	170
STATEMENT OF AUTHORSHIP	171
6.1 Abstract.....	174
6.2 Introduction.....	175
6.3 Materials and methods.....	176
6.3.1 Participants	176
6.3.2 Protocol	178
6.3.3 Measurements.....	179
6.3.3.1 Blood glucose, plasma insulin and plasma glucagon concentrations ...	179
6.3.3.2 Gastric emptying.....	180
6.3.3.3 Energy intake.....	180
6.3.3.4 Perception of hunger and fullness.....	181
6.4 Data and statistical analysis	181
6.5 Results	183
6.5.1 Blood glucose concentrations.....	183
6.5.1.1 Interaction effects	183
6.5.1.2 Drink-condition effects	184
6.5.1.3 Group effects	184
6.5.2 Plasma insulin.....	189
6.5.2.1 Interaction effects	189
6.5.2.2 Drink-condition effects	189
6.5.2.3 Group effects	189
6.5.3 Plasma glucagon	190
6.5.3.1 Interaction effects	190
6.5.3.2 Drink-condition effects	190
6.5.3.3 Group effects	190
6.5.4 Gastric emptying.....	192

6.5.4.1 Interaction effects	192
6.5.4.2 Drink-condition effects	192
6.5.4.3 Group effects	192
6.5.5 Energy intake	193
6.5.6 Perception of hunger and fullness	194
6.6 Discussion.....	196
6.7 Conclusions.....	199
CHAPTER 7: EFFECTS OF AGE ON BLOOD PRESSURE AND HEART RATE RESPONSES TO WHEY PROTEIN IN YOUNGER AND OLDER MEN.....	201
STATEMENT OF AUTHORSHIP.....	202
7.1 Abstract.....	204
7.2 Introduction.....	206
7.3 Methods	207
7.3.1 Participants	207
7.3.2 Protocol	208
7.3.3 Measurements.....	209
7.3.3.1 Blood pressure and heart rate	209
7.3.3.2 Perceptions of light-headedness and drowsiness.....	209
7.3.3.3 Gastric emptying.....	210
7.4 Statistical analysis	210
7.5 Results	211
7.5.1 Systolic blood pressure	211
7.5.2 Diastolic blood pressure.....	214
7.5.3 Heart rate.....	214
7.5.4 Postprandial hypotension	215
7.5.5 Perceptions of light-headedness and drowsiness.....	216
7.5.6 Gastric emptying.....	217
7.6 Discussion.....	218
CHAPTER 8: BLOOD PRESSURE AND HEART RATE RESPONSES FOLLOWING DIETRAY PROTEIN INTAKE IN OLDER MEN	223
STATEMENT OF AUTHORSHIP	224
8.1 Abstract.....	226
8.2 Introduction.....	227
8.3 Materials and methods.....	229
8.3.1 Participants	229
8.3.2 Protocol	230

8.3.3 Measurements.....	231
8.4 Data and statistical analysis	231
8.5 Results	232
8.5.1 Systolic blood pressure (SBP).....	232
8.5.2 Diastolic blood pressure (DBP).....	236
8.5.3 Heart rate (HR)	237
8.6 Discussion.....	237
8.7 Conclusions.....	240
CHAPTER 9: ACUTE EFFECTS OF WHEY PROTEIN, ALONE AND MIXED WITH OTHER MACRONUTRIENTS, ON BLOOD PRESSURE AND HEART RATE IN OLDER MEN.....	241
STATEMENT OF AUTHORSHIP	242
9.1 Abstract.....	245
9.2 Introduction.....	247
9.3 Materials and methods.....	248
9.3.1 Participants	248
9.3.2 Protocol	249
9.3.3 Measurements.....	250
9.4 Data and statistical analysis	250
9.5 Results	251
9.5.1 Systolic blood pressure (SBP).....	251
9.5.2 Diastolic blood pressure (DBP).....	253
9.5.3 Heart rate (HR)	254
9.6 Discussion.....	255
9.7 Conclusions.....	258
CHAPTER 10: CONCLUSIONS OF THE THESIS	259
GLOSSARY.....	264
REFERENCES.....	266

THESIS ABSTRACT

Australia's population is ageing. Health problems are more prevalent in elderly than younger people, and lead to much of the health expenditure and health resource allocation in this and other countries. Nutritional problems are common in the elderly and often contribute to the development and worsening of health problems in older people. A particular problem is the interacting effects of under-nutrition, reduced dietary protein intake, anabolic resistance to dietary protein (the need for greater intakes to produce the same benefits, particular on muscle mass and function) and consequent muscle loss, reduced function, and in some cases the development of sarcopaenia. Both type 2 diabetes (T2D) and obesity are prevalent in older people – and often coexist. Dietary measures are key to the management of both conditions, in older as in younger adults. Protein supplements either alone or in combination with other macronutrients are increasingly recommended to, and used by, older people, to prevent and/or manage these problems. Whey protein is often used alone or as part of these supplements, as it has particularly beneficial effects on muscle anabolism. There remain gaps in our knowledge of the effects of whey protein ingestion in older people. This thesis comprises clinical studies designed to fill some of those gaps.

The studies in this thesis from five clinical trials involved assessing responses to oral whey protein ingestion particularly whey protein concentrate on feeding behaviour, nutrition, obesity, T2D and blood pressure (BP) in older and younger people with a focus on older people. Dietary protein supplementation may play a role in the management of these conditions. The principal outcomes assessed were appetite; food intake; gut hormone release; gastric emptying rate; circulating glucose,

glucagon and insulin concentrations; BP and heart rate (HR).

Chapter 1: Introduction focusses on rising burden of ageing in Australia and other developed countries; nutritional problems and adverse body composition changes associated with ageing i.e. undernutrition and sarcopaenia; nutritional management of under-nutrition and sarcopaenia with a focus on the effects of whey protein; rising rates of obesity, T2D and BP effects in older people and whey protein as a management strategy; gastric emptying and its effects in older people.

Chapter 2 is a review of the current literature relating to the use of protein supplements in the elderly, dietary protein requirements in older people, and the effects of ageing on the gastrointestinal responses to protein ingestion.

Chapter 3 describes the effect of differing whey protein loads (control, 30 g whey protein and 70 g whey protein) before breakfast on appetite and food intake at subsequent breakfast, lunch and dinner in healthy younger and older men. Energy intake was suppressed by whey protein drinks in a protein load-responsive fashion at breakfast and particularly, at lunch, but not at dinner, and suppression of energy intake by protein was less in healthy older than younger men. Cumulative protein intake was increased in a protein load responsive fashion. These findings support the use of whey-protein drink supplements in healthy older patients who aim to increase their protein intake without decreasing their overall energy intake (ACTRN12618000881235).

Chapter 4 describes the effect of whey protein load on energy intake, appetite and gastric emptying in young and older, obese men. The 30 g whey protein drink did not suppress appetite or energy intake in obese younger or older men suggesting that

obesity may blunt/abolish the age-related effect of whey protein on suppression of energy intake (ANZCTR12616001216404).

Chapter 5 describes the effects of co-ingesting whey protein and glucose alone and combined (a drink containing either 30 g glucose, 30 g whey protein, 30 g whey-protein plus 30 g glucose or control) on blood glucose, plasma insulin and glucagon concentrations in healthy younger and older men. The addition of 30 g of whey protein to 30 g of glucose in drink form substantially attenuated the increase in blood glucose concentrations induced by glucose alone; the magnitude of the whey-induced reduction in blood glucose was not affected by age, with comparable reductions in older to those in younger adult men; the stimulation of plasma insulin concentrations by whey protein was not reduced by ageing, unlike the insulin response to glucose; whey protein suppressed hunger less in older than younger men. Glucagon concentrations were unaffected by age. These results demonstrate that the ability of whey-protein to reduce carbohydrate-induced postprandial hyperglycaemia is retained in older men and that protein supplementation may be a useful strategy in the prevention and management of T2D in older people (ACTRN12619000420145).

Chapter 6 describes the effects of co-ingesting whey protein and glucose alone and combined (a drink containing either 30 g glucose, 30 g whey protein, 30 g whey-protein plus 30 g glucose or control) on blood glucose, plasma insulin and glucagon concentrations in older men with (not on injectable treatment) and without T2D. The addition of 30 g of whey protein to 30 g of glucose in drink form substantially attenuated the increase in blood glucose concentrations induced by glucose alone; the magnitude of the whey-induced reduction in blood glucose was not affected by the presence of T2D and the stimulation of plasma insulin concentrations by whey

protein. The ability of whey-protein to reduce carbohydrate-induced postprandial hyperglycaemia is retained in men with T2D (ACTRN12619000420145).

Chapter 7 describes the effects of age on BP and heart rate responses to whey protein in healthy younger and older men after oral ingestion of 0 g and 70 g whey protein. The older men exhibited a greater fall in SBP after whey-protein versus control than the younger men, with no BP change after the two drinks in younger men. The nadir in SBP occurred later in the older than younger men with SBP still apparently declining 180 min after whey-protein ingestion in the older men. The magnitude of the rise in HR was greater in the younger men indicating that following ingestion of 70 g whey protein, healthy older men exhibited a sustained fall in BP, despite an increase in HR, whereas in younger men there was no change in BP. BP may need to be monitored after high protein meals in older people at risk of postprandial hypotension (ACTRN12612000941864 and ACTRN12614000846628).

Chapter 8 describes the BP and heart rate responses to oral protein intake in healthy older men after ingestion of 30 g and 70 g of whey protein. The older men exhibited a decrease in systolic blood pressure (SBP) after ingestion of 30 g and 70 g of whey protein to a similar degree after both the drinks and greatest between 120 and 180 mins after ingestion. HR was increased maximally after 70 g particularly in the third hour (hr) and diastolic blood pressure (DBP) decreased non significantly after protein drinks. Even modest protein loads in older men can result in postprandial hypotension (PPH) and care must be taken (ACTRN12612000941864).

Chapter 9 describes the acute effects of whey protein, alone and mixed with other macronutrients in varying amounts such as 70 g whey-protein (P₂₈₀); (ii) 14 g whey-

protein, 28 g carbohydrate, 12.4 g fat (M₂₈₀); (iii) 70 g whey-protein, 28 g carbohydrate, 12.4 g fat (M₅₀₄); or (iv) a non-caloric control drink (C) on BP and heart rate in healthy older men. SBP decreased after all three nutrient drinks with the greatest reduction after the M₅₀₄. Maximal decreases in SBP occurred about 2 hr after drink ingestion and were sustained thereafter. Maximum DBP decreases and HR increases occurred after M₅₀₄, with no differences between the effects of the P₂₈₀ and M₂₈₀ drinks, thereby demonstrating that the effects of whey-protein containing drinks to lower BP and increase HR appear to be primarily dependent on their energy content rather than macronutrient composition and may persist for at least 3 hr after ingestion. Pure whey-protein drinks may represent the best approach to maximize protein intake without increasing the potential for deleterious BP falls in older people (ACTRN12614000846628).

DECLARATION

I certify that this work contains no material which has been accepted for the award of any other degree or diploma in my name, in any university or other tertiary institution and, to the best of my knowledge and belief, contains no material previously published or written by another person, except where due reference has been made in the text. In addition, I certify that no part of this work will, in the future, be used in a submission in my name, for any other degree or diploma in any university or other tertiary institution without the prior approval of the University of Adelaide and where applicable, any partner institution responsible for the joint award of this degree.

The author acknowledges that copyright of published works contained within the thesis resides with the copyright holder(s) of those works.

I give permission for the digital version of my thesis to be made available on the web, via the University's digital research repository, the Library Search and also through web search engines, unless permission has been granted by the University to restrict access for a period of time.

I acknowledge the support I have received for my research through the provision of an Australian Government Research Training Program Scholarship.

Avneet Kaur Oberoi October, 2022 BSc, MSc

ACKNOWLEDGEMENTS

This thesis represents the scientific work I have conducted in the last 4 years of my PhD journey at the Discipline of Medicine, The University of Adelaide - a unique journey with loads of experience, challenges and great learning that I will carry and cherish all my life. This would not have been possible without the enormous and incredible support of people who made it possible and I would like to thank them for their contribution and support for this achievement.

Firstly, I would like to express my gratitude to my supervisors, without whose support and guidance this accomplishment could never have been achieved. Thank you for your immense knowledge, continuous support, professional guidance, appreciation, encouragement, insightful comments, patience, and motivation. I consider myself very blessed to have such a wonderful and supportive supervisory panel each with their own unique research and academic expertise, and I think I could not have gotten through it without their constant support and guidance.

I am deeply indebted to, Professor Ian Chapman, my main supervisor, for believing in me and providing me a platform to grow. Ian has been a great mentor and advisor in my life by giving me invaluable professional and personal advice, helping to shape and polish my ideas, pushing me beyond my comfort zones, and motivating me. Your excellent academic and writing skills have guided me to improve and build upon my presentation skills and present my work with confidence. Your knowledge and wisdom has been invaluable and I will always be grateful for your experience. Your perseverance, time and feedback that was put into my work will always be valued by me. Thank you for your immense patience. Words are not enough to express what I have

Acknowledgements

learnt from you as a great leader. It has been an absolute pleasure working under your supervision. Thank you for being my generous financial sponsor. I hope I have never let you down. Thanks, Ian.

I am extremely grateful to Professor Karen Jones, my co-supervisor, for being my family and guardian any one would wish for, in a foreign land and a brand-new challenging role. I appreciate I could knock on your door anytime and discuss even the silliest things with you anytime. Your eye for detail, constant feedback and guidance has been incredible in shaping me who I am. I cannot thank you more for the support and motivation you have provided me. Your easy-going approach has contributed to a comfortable working environment. I am going to cherish all the laughs and conversations we had over coffee. Your appreciation encourages me to overcome my limitations. Thank you for helping me getting the scholarship. Thank you, Karen.

I would like to express my deep gratitude to Dr Stijn Soenen, my co-supervisor. I have learnt a lot from your attention to detail. I still remember our first meeting at AHMS where you spent a significant amount of time motivating me to join PhD when I was a bit confused. Your enthusiasm for research, experience and subject in-depth knowledge has taught me to persist, learn and move forward even when things did not go as planned. Thank you for the valuable input, detailed and timely feedback and guidance given throughout the duration of the project. Your valuable criticism at times has motivated me to be on top of things. Thank you for helping me out and making me understand the statistical analysis and results for the studies. I truly appreciate your patience. Your positive attitude has always boosted my confidence. I look forward to meeting you in Adelaide. Thank you, Stijn.

Acknowledgements

To Professor Michael Horowitz, I appreciate your thorough revisions and detailed input in my manuscripts, and your expertise has greatly improved my interest in the world of clinical research. Thank you for helping me getting a radiation licence and guiding me through always. Thank you, Professor Michael.

A very special thanks to Rachael Tippet, who has always been there for me- a friend, a mentor. Someone I can always rely upon, listening to my stories and sharing ins and outs. I am going to miss every moment spent with her. From helping me out with my experiments to going out for walks and having coffee together, the bond we developed cannot be shared in words. Thank you, Rachael.

I express my gratitude to my postgraduate coordinator (PGC), Dr. Christina Bursill for her support and guidance in this journey.

A very special thanks to the senior researchers in the Discipline of Medicine, Seva Hatzinikolas, Penelope Fitzgerald, Michelle Bound, for always standing by my side, from teaching me venepuncture and cannulation and guiding me throughout.

Thank you to all the staff of the CRE for a supportive working atmosphere, especially the reliable and experienced staff I was fortunate enough to work with. To Bree Hodgson, I am grateful for your organisational skills from which I have benefited many times along the way. Thanks a lot to the interns who assisted me with my studies. Thank you to Scott Standfield and Judith Wishart for the hormone analyses and Kylie Lange for your support with the statistical analyses, which has greatly improved the quality of my manuscripts. Your experience in research and interest in my work have been a great motivation. I would not have been able to complete the

studies in this thesis without all of your help.

To my fellow PhD students, Vida, Cong, Peyman, Maryam, although we did not work in the same group, I felt like we still worked as a great team. Your thoughtfulness made my life easier on many occasions.

The time and generosity gifted by the research participants has really moved me, and without them there would not have been any results to report on. Not only have I benefited from the data I have collected from you, also your life stories and advice have inspired me. I would like to thank you for your time and trust to make this possible.

I would like to acknowledge that my PhD candidature was funded by the University of Adelaide through Research Training Program Scholarship by the Commonwealth Government and Nutritional physiology PhD supplementary scholarship. Thank you, University of Adelaide.

I would also like to thank Joe Miller from The Edit Bureau for timely editing some parts of my thesis such as references, table of contents, margins and spacing. Thanks, Joe.

Above all, I would like to thank the almighty Waheguru and my family for always standing by my side and giving me strength to face all the challenges in life. I have been blessed with two sets of the most supportive and loving parents, my dad, mum, father-in law and mother-in law. This journey would not have been possible without you. No amount of words can ever explain how grateful I am to all of you. A special call out for my mum, my dad, my sister, my little nieces, my father-in law and

Acknowledgements

mother-in law for always been caring, understanding and motivating me to successfully complete this journey. I am what I am because of my mum and dad, who have forever trusted me and gave me their company for many early mornings and late nights while I was working and when I needed their support. They have given up on many things to support me in all possible ways especially providing a listening ear. I am grateful to my parents for putting up with me all through this long journey even when I was irritable. I am sure all of you are proud of me.

A very special thanks to the most important person in my life, my husband Karan for all his love, support, encouragement and motivation through this incredible journey which I could not have imagined to achieve myself. Thank you for listening to my stories and spending sleepless nights with me in making me what I am today. You are a great friend and mentor. You have been by my side for the last 7 years and I do not think I could have achieved anything without the love and strength I get from you. I am forever grateful and thankful for the day you walked into my life and have always pushed me towards excellence, for you are the person who believed in me when nobody else did. Thank you, Karan.

A big thank you to everyone who has been a part of my PhD journey in one way or the other and whose names I may have missed out. This journey has indeed been life changing.

Avneet Oberoi

October, 2022

PUBLICATIONS ARISING FROM THIS THESIS

1. Chapman I, **Oberoi AO**, Giezenaar C, Soenen S. *Rational use of protein supplements in the elderly- relevance of gastrointestinal mechanisms*. *Nutrients* 2021; 13: 1227. doi: doi.org/10.3390/nu13041227 (**Chapter 2**).
2. **Oberoi AO**, Giezenaar C, Clames A, Böhler K, Lange K, Horowitz M, Jones KL, Chapman I, Soenen S. *Whey protein drink ingestion before breakfast suppressed energy intake at breakfast and lunch, but not during dinner, and was less suppressed in healthy older than younger men*. *Nutrients* 2020; 12: 3318. doi: 10.3390/nu12113318 (**Chapter 3**).
3. **Oberoi AO**, Giezenaar C, Jensen C, Lange K, Hausken T, Jones KL, Horowitz M, Chapman I, Soenen S. *Acute effects of whey protein on energy intake, appetite and gastric emptying in younger and older, obese men*. *Nutrition & Diabetes* 2020; 10: 37. doi.org/10.1038/s41387-020-00139-8 (**Chapter 4**).
4. **Oberoi AO**, Giezenaar C, Rigda RS, Lange K, Horowitz M, Jones KL, Chapman I, Soenen S. *Comparative effects of co-ingesting whey protein and glucose alone and combined on blood glucose, plasma insulin and glucagon concentrations in younger and older men*. *Nutrients* 2022; 14: 3111. doi: 10.3390/nu14153111 (**Chapter 5**).
5. **Oberoi AO**, Giezenaar C, Rigda RS, Horowitz M, Jones KL, Chapman I, Soenen S. *Effects of co-ingesting glucose and whey protein on blood glucose, plasma insulin and glucagon concentration, and gastric emptying, in older men with and without type 2 diabetes men*. Submitted to *Diabetes, Obesity and Metabolism* 2022 (**Chapter 6**).

6. Giezenaar C, **Oberoi AO**, Jones KL, Horowitz M, Chapman I, Soenen S. *Effects of age on blood pressure and heart rate responses to whey protein in younger and older men*. Journal of the American Geriatrics Society 2021; 69:1291–1299. doi: 10.1111/jgs.17083 (**Chapter 7**).

7. **Oberoi AO**, Giezenaar C, Lange K, Jones KL, Horowitz M, Chapman I, Soenen S. *Blood pressure and heart rate responses following dietary protein intake in older men*. Nutrients 2022; 14: 1913. doi:10.3390/nu14091913 (**Chapter 8**).

8. **Oberoi AO**, Giezenaar C, Lange K, Jones KL, Horowitz M, Chapman I, Soenen S. *Acute effects of whey protein, alone and mixed with other macronutrients, on blood pressure and heart rate in older men*. BMC Geriatrics 2022; 22: 535. doi:10.1186/s12877-022-03213-1 (**Chapter 9**) (Work published on Central Adelaide Local Health Network and Royal Adelaide Hospital website-<https://centraladelaide.health.sa.gov.au/protein-and-blood-pressure/>).

ABSTRACTS AND PRESENTATIONS

1. **Oberoi AO**, Giezenaar C, Rigda R.S, Lange K, Horowitz M, Jones KL, Chapman I, Soenen S. *Effects of whey protein on blood glucose concentrations with oral glucose intake in younger and older men.* Australasian Diabetes Congress (ADC), Brisbane, August, 2021 (**Poster**).
2. **Oberoi AO**, Giezenaar C, Rigda R.S, Lange K, Horowitz M, Jones KL, Chapman I, Soenen S. *Effects of whey protein on blood glucose concentrations with oral glucose intake in younger and older men.* 15th Florey Postgraduate Conference, Adelaide, 2021 (**Poster**).
3. **Oberoi AO**, Giezenaar C, Jensen C, Lange K, Hausken T, Jones KL, Horowitz M, Chapman I, Soenen S. *Effects of whey protein on energy intake, gastric emptying and blood glucose in obese, older and younger men.* ESNM NeuroGastro, September, 2021 (**Poster**).
4. **Oberoi AO**, Giezenaar C, Rigda R.S, Lange K, Horowitz M, Jones KL, Chapman I, Soenen S. *Effects of whey protein on blood glucose concentrations with oral glucose intake in younger and older men.* 49th American Aging Association, Virtual 2021 (**Poster**).
5. **Oberoi AO**, Giezenaar C, Jensen C, Lange K, Hausken T, Jones KL, Horowitz M, Chapman I, Soenen S. *Effects of whey protein on energy intake, gastric emptying and blood glucose in obese, older and younger men.* 4th Meeting of the Federation of Neurogastroenterology and Motility (FNM), Adelaide, 2021 (**Poster**). **Recipient of AMS HDR Travel Research Support Award, Round 1 2021.**

6. **Oberoi AO**, Giezenaar C, Rigda R.S, Lange K, Horowitz M, Jones KL, Chapman I, Soenen S. *Effects of whey protein and glucose intake on glycaemia and energy intake in older men*. Precision in Diabetes Medicine (PDM), 2021 **(Poster)**.
7. **Oberoi AO**, Giezenaar C, Rigda R.S, Lange K, Horowitz M, Jones KL, Chapman I, Soenen S. *Effects of whey protein and glucose intake on glycaemia and energy intake in older men*. 56th European Association for the Study of Diabetes (EASD), Vienna, 2020 **(Poster)**. **Recipient of AMS HDR Travel Research Support Award.**
8. **Oberoi AO**, Giezenaar C, Jensen C, Lange K, Hausken T, Jones KL, Horowitz M, Chapman I, Soenen S. *The effect of a whey protein drink on energy intake, appetite and gastric emptying in younger and older, obese men*. The Medical Staff Society Research Prize, Royal Adelaide Hospital, Adelaide, 2020 **(Oral)**.
9. **Oberoi AO**, Giezenaar C, Rigda R.S, Lange K, Horowitz M, Jones KL, Chapman I, Soenen S. *Effects of whey protein and glucose intake on glycaemia and energy intake in older men*. Australian Diabetes Congress (ADC), Brisbane, 11-13 November, 2020 **(Poster)**.
10. **Oberoi AO**, Giezenaar C, Jensen C, Lange K, Hausken T, Jones KL, Horowitz M, Chapman I, Soenen S. *The effect of a whey protein drink on energy intake, appetite and gastric emptying in younger and older, obese men*. 58th Australian Society of Medical Research National Scientific Conference (ASMR), Adelaide, 2019 **(Oral)**.

CHAPTER 1: INTRODUCTION

INTRODUCTION

Our research group has a longstanding interest in undernutrition in older people, its possible causes, adverse consequences and management strategies. One of the more widely used management strategies is protein or protein-enriched nutritional supplements. These have been shown to help preserve muscle mass and function in older people (1-4). Whey protein has particular anabolic properties which make it a favourable choice as a protein supplement in older people and such supplements are being used increasingly by older people (5-8). Our group has conducted studies assessing appetite and feeding responses to whey protein in older people and the effect of healthy ageing on these responses (9-18). These studies have largely involved assessment of appetite ratings from the time of whey intake to a single later test meal and of food intake at that meal. More recently, we have expanded the scope of these studies to examine the effects of whey on feeding behaviour not just at the next meal but throughout the study day (chapter 3) and in obese as well as non-obese participants (chapter 4). We have also now studied the effects of whey in older people on outcomes that are possibly both beneficial (blood glucose lowering: chapters 5 and 6) and harmful (excessive BP lowering: chapters 7-9). These studies on additional effects of whey protein form the basis of this thesis.

The following sections of the introduction provides a background to and rationale for the studies described in this thesis.

1.1 The rising burden of ageing in the population of Australia and other developed countries

The populations of developed countries, including Australia, are ageing. Worldwide, the percentage of populations over 60 years increased from 8% in 1950 to 12% in 2014 and is expected to reach 21% by 2050 (19). Global life expectancy increased from 47 years to about 70 years from 1950 to 2014 and will increase to 75 years by 2050 (20, 21). The proportion of Australia's population over 65 years of age increased from 5% in 1937 to 9% in 1976 and 15% in 2016, and is expected to rise to more than 22% by 2056 (22). These increases impose a significant personal and national economic burden, as many disease conditions and disabilities become more common with increasing age, adversely impacting quality of life (22). The percentage of Australian health spending expended on people over 65 years of age is expected to increase from 71% in 2012 to 84% by 2035 (23). Therefore, optimising the health of older people is crucial for individual and societal benefits.

1.2 Nutritional problems and adverse body composition changes associated with human ageing

1.2.1 Undernutrition and sarcopaenia

Ageing is associated with changes in appetite, feeding behaviour, gut function and body composition (2). With healthy ageing, there is reduction in appetite and food intake, which has been termed the 'anorexia of ageing' (1, 24). Between 20 and 80 years, the average daily energy intake decreases by up to 30%. Older people have reduced hunger ratings, and increased fullness and satiety compared to younger adults (25-30). In the American National Health and Nutrition Examination Survey,

NHANES III, 1989, for example, there was a reduction in the average daily energy intake of about 1321 calories/day in men and 629 calories/day in women between the ages of 20-80 years (31, 32). The mechanisms responsible for reduced appetite and food intake in older people are multiple and complex, and include both central and peripheral (gastrointestinal) factors.

Often not appreciated is that weight loss is more common than weight gain in older people (33, 34). Prospective and cross-sectional studies demonstrate that body weight and body mass index (BMI) tends to increase up until 50-60 years of life and decline thereafter (1, 35). In one study older American men > 65 years of age lost 0.5% of their body weight per year on an average over 2 years and 13.1% had involuntary weight loss of 4% per annum or more (36). The prevalence of nutritional risk in older people as per the Mini Nutritional Assessment varies from 45% to 100% (community, domiciliary settings, hospitals and aged care facilities) (37-41), and when marked this can result in nutritional problems, particularly undernutrition which can have deleterious effects on the function and quality of life of older people (33).

Weight loss, especially if involuntary, is a predictor of poor outcomes in older people (42) with an approximately two-fold increased risk of mortality with a weight loss of 4-5% in about 1-3 years (34). In a longitudinal observation study known as the cardiovascular health study conducted in 1989 on 4714 community dwelling people aged 65 years and above showed that a weight loss of 5% in 17% participants resulted in a significant increase in the mortality rate (crude mortality rate- 6 in 100 person; age and gender adjust hazard ratio was 2.09; 95% confidence interval (CI), 1.67–2.62 and risk-adjusted was 1.67; 95% CI, 1.29–2.15) (34). Furthermore, in the

systolic hypertension in the elderly program from 1984-1990, an average weight loss of 1.6 kg/year was associated with all-cause mortality (odds ratio- 4.9, 95% CI, 3.5–6.8) as compared to a weight stable group used as a reference (43).

The age-associated weight loss is disproportionately due to the loss of lean rather than fat tissue. On average, body fat increases and lean muscle mass (fat free mass) decreases throughout adult life. With ageing, there are morphological and (44) physiological changes in skeletal muscle such as reduced number and size of skeletal muscle fibres along with infiltration of adipose cells into skeletal muscle. Skeletal muscle atrophy begins in the third or fourth decade of life, with a rate of muscle loss of about 3-8% per decade after 30 years of age (45, 46). This muscle loss is chiefly due to an imbalance between muscle protein synthesis and muscle protein breakdown, due to increased oxidative stress, inflammation and reduced mitochondrial capacity (47).

Loss of muscle mass contributes to reduced physical strength. The rate of decline in muscle strength is about 2-4% per year after 50-60 years of age (48). This further results not only in the functional impairments such as reduced grip strength and gait but speed but also disability (49). Age-related reductions in muscle mass, strength and function are associated with increased rates of falls, fractures and frailty (2, 50-52). These adverse outcomes are even more likely when muscle mass is excessive, a condition that has been termed sarcopaenia (53). Age related changes in body composition are chiefly responsible for sarcopaenia when it develops, but environmental factors such as reduced physical activity due to chronic diseases such as obesity, and medical conditions leading to fatigue, pain, inflammation and inadequate calorie and protein intake can also contribute (54). These changes are

associated with adverse health outcomes such as poor cognitive function, immunodeficiency, increased rates of hospitalisation and admission to nursing homes and ultimately increased mortality (32).

1.2.1.1 Nutritional management of under-nutrition and sarcopaenia

A recognition of the adverse outcomes associated with age-related weight loss and sarcopaenia has led to attempts to prevent and treat it. Nutritional measures and exercise are commonly used strategies and usually first line (55, 56). For example, the Food First approach (an evidence based strategy enabling individuals to consume a variety of food for good health and performance and also eat intuitively, promoting social and emotional wellbeing) is one such first line of treatment to maintain weight, prevent frailty and sarcopaenia (57). This approach combines strategies aimed at increasing food intake (consuming large amounts of usual foods to maintain body weight i.e. increasing the nutrient content of the food particularly energy and protein rich or cooking nutritious meals for the elderly or providing home meal services like meals on wheels especially in aged care setting; fortification with additional nutrients) and strategies aimed at improving knowledge and behaviour (provide one on one counselling, tailored diet plan, a resource kit by a professional and increased encouragement to eat) and often helps to counteract weight loss (58-62). However, the successful implementation of such strategies is not always achievable due to a various reasons such as affordability, convenience and staffing issues and variable adherence (58).

The next step in improving the nutrition of older people is often the use of nutritional supplements, particularly protein supplements, which are increasingly recommended

to and used by older people to achieve increased energy and protein intake (63). Supplements provide the opportunity to tailor the timing and composition of added protein for maximum benefit (63).

Protein is considered to be the most satiating of all macronutrients, at least in non-elderly people. High protein diets are associated with increased satiety, thermogenesis and improved insulin sensitivity (64-66). They have shown to assist in weight loss in young adults with obesity and improve oral glucose tolerance, and cardiovascular outcomes in young adults with T2D (67-69). In non-elderly adults, dietary protein facilitates weight loss by suppression of energy intake and appetite (9, 15, 25-28, 70-72), preservation of lean mass (73-76) and has greater satiating effect than carbohydrates or fats (77, 78). A classic study by Weigle et al. showed that a diet containing 30% of protein versus an iso-caloric diet with 15% of protein was more satiating and associated with greater weight loss over time and decreased energy intake (79). McAuley et al. showed greater insulin sensitivity in obese women with 30% protein diet at 8 weeks as compared to high fat or carbohydrate diets (80). A diet containing 30% protein showed ~ 0.5 % reductions in HbA1c in patients with T2D (66).

The ability of ingested protein to stimulate muscle protein synthesis decreases with ageing, a change that has been termed anabolic resistance (6, 81). As a result, older people need to consume larger amounts of protein to preserve and enhance muscle mass than young adults and this is reflected in age-related recommendations for dietary protein intake (82, 83). Whereas recommendations for dietary protein intake in non-elderly adults usually recommend approximately 0.8 g/kg/day, the European PROT-AGE group, for example, recommends 1.0–1.2 g/kg dietary protein for

healthy older adults, 1.2-1.5 g/kg for older adults with chronic illness. This equates to about 30–45 g protein per serving, whereas lower doses < 20 g are sufficient for younger adults (84). In order to meet these guidelines, older people are often recommended to consume high protein foods, plus or minus protein or protein-enriched supplements. Further detail about the type of protein, amount and timing along and possible adverse effects of protein supplements are detailed in **chapter 2, page 58-67**.

Because dairy products are a good source of protein, increasing the intake of milk and dairy products is often recommended as a way to increase protein intake (85, 86).

1.2.2 Whey protein

Our group has been interested in the nutritional management of under-nutrition and sarcopaenia, particularly the role of whey protein.

Whey protein is obtained from milk, as a by-product of cheese making process in which it is separated from casein. It accounts for 20% of the total milk protein. As compared to casein it is considered a “fast protein” as it is more soluble in stomach resulting in rapid digestion. It is also considered a “complete” protein with a high biological value of 104 (proportion of protein retained in the body for growth and/or maintenance and expressed in percent of nitrogen absorbed) and net protein utilisation of 92 (ratio of amino acid mass converted to proteins to the mass of amino acids supplied) (87, 88). It contains bioactive peptides with β -lactoglobulin (45–57%) present in high amounts, whereas α -lactalbumin (15–25%), immunoglobulin (10–15%), glycomacropeptide (10–15%), bovine serum albumin (10%), lactoferrin

(~1%) and lactoperoxidase (< 1%) accounts for smaller amounts (4, 15, 16) (Table 1).

Whey protein drinks in doses of 30–70 g slow gastric emptying in a dose-responsive manner (42, 44, 50), and probably slow it more in older than in young adults. Whey ingestion has effects on a number of gastrointestinal factors controlling appetite and food intake, particularly mediated by the release of satiety-inducing hormone e.g. concentrations of cholecystokinin (CCK), insulin, glucagon, gastric inhibitory peptide (GIP), glucagon-like peptide-1 (GLP-1), peptide tyrosine-tyrosine (PYY), and amino acids increase, while glucose does not change and ghrelin concentrations decrease (89). Food intake is reduced via the release of gut hormones such as CCK and GLP-1 in response to amino acid and peptide release by consumption of whey protein (89). Ingestion of nutrients results in CCK and GLP-1 release from small bowel that slows gastric emptying and reduces food intake, typically seen in older people causing satiety (29, 90). Bioactive peptides present in whey or formed during digestion are the main stimulators of GIP secretion which increase more after oral whey ingestion in older than in young adults (11, 12, 14, 91). Whey suppresses circulating concentrations of the appetite-stimulating hormone ghrelin, and increases glucagon concentrations in older and younger adults (90).

An important factor influencing the satiating effect of different proteins is their amino-acid composition and effects on circulating amino acid concentrations after ingestion (89).

Table 1: Essential amino acid content of whey protein

Amino acid	Amount
Leucine	8.6
Threonine	5.4
Methionine	1.8
Phenylalanine	2.5
Histidine	1.4
Lysine	7.1
Valine	3.5
Isoleucine	3.8
Sum of essential amino acid	34.1

Values presented are in g per 100 g raw material

Whey has a relatively high content of the branched chain amino acids leucine, isoleucine and valine and these amino acids, particularly leucine are more potent than other amino acids in stimulating muscle protein synthesis (92, 93). In older people, muscle protein synthesis was increased with 42% content of leucine and not with a mixture having leucine content of 26% (94). Consequently whey protein is considered to be an effective anabolic protein. It is commercially available in varying forms, including as a whey protein concentrate, whey protein isolate, and whey protein hydrolysate (86, 95, 96). These are increasingly used as supplements in an attempt to preserve or increase muscle function.

Previous studies by our group and others have indicated that oral ingestion of whey is capable of suppressing appetite and food intake in older, as in young adults, but

that this effect is less potent in older adults, with substantially higher doses of whey necessary to suppress food intake in older people. In one study when intraduodenal infusions of 8 g, 24 g and 48 g of whey protein were administered to older and young men, the older men had suppression of *ad libitum* energy intake only with the highest dose, whereas the younger men suppressed with lower doses (9). This finding has been supported by the results of other studies (27). Our group has conducted a number of studies using 30 g and 70 g of whey protein, 30 g approximating the minimum amount required in older people to stimulate muscle protein synthesis (see 1.2.1.1 above) (63, 83, 97). Compared to ingestion of an energy-free control drink oral administration of 30 g of whey protein suppressed energy intake less in older ($1 \pm 5\%$) than compared to younger men ($15 \pm 2\%$; $P = 0.008$) (14, 15, 98) Appetite decreased in young, while it increased in older men (desire to eat, young vs control, a decrease of 647 ± 910 mm vs an increase of 1027 ± 458 mm, $P = 0.024$; prospective food consumption, young vs control, a decrease of 1616 ± 706 mm vs an increase of 119 ± 370 mm, $P = 0.021$) (15). This reduced suppressive effect of whey on feeding behaviour in older people is likely to be a good thing in most situations, as whey ingestion is therefore less likely to lead to appetite suppression and weight loss when whey supplements are used in an attempt to enhance muscle function in older people.

The optimum timing of ingestion of whey supplements in older people is uncertain. If maximisation of protein intake without suppression of appetite and food intake overall, and whey suppresses appetite, it might be best to give the supplements between meals, so as not to reduce food intake at main meals. If, on the other hand, as suggested by the above studies, appropriate doses of whey have little or no appetite suppressant effect in older people, the timing of ingestion may be less important. Also, by ingesting whey with a main meal, which contains other protein in “usual”

foods, it may be possible to achieve optimum single dose protein intake for muscle benefit (30 g or higher) with a lower dose of whey that would be needed if ingested between meals. The duration of effect of whey on appetite and feeding behaviour in older people have also not been well defined.

1.2.3 Obesity in older people

Our group's previous studies have focused on the effects of whey on non-obese older people. Given the adverse effects of obesity and its rising prevalence, including in older people, we have now examined the effect of whey on feeding behaviour in obese, older people.

Increasing rates of obesity are a global health problem (99-101). Although weight loss is more common than weight gain in older people (see 1.2.1), obesity rates are rising in Australia, including in older people, mainly because of the increasing number of people achieving old age already overweight (102, 103). This is a key health issue for older Australians as it is associated with an increased risk of developing long term, chronic degenerative disorders including T2D, osteoarthritis, cardiovascular disorders, hypertension, certain types of cancers and increased rates of overall poor health and disability (104, 105). For example, accumulation of adipose tissue, particularly when sufficient to cause obesity, plays a critical role in causing T2D by causing insulin resistance and other mechanisms (106). Over recent decades the increasing prevalence of obesity has been closely associated with a marked rise in T2D, and this frequent association has been termed diabetesity (107). In 2017-2018, 78% of Australians between 68-74 years were overweight (BMI 25-29.9 kg/m²) or obese (BMI > 30 kg/m²) as compared to 65% aged 18-64 years (103).

It is expected that by 2030, 51% of the world's population will be obese by BMI criteria (108). Around 50–90% of people with T2D have a BMI > 25 kg/m², with even higher rates reported in older people with T2D (109).

Due to various hormonal and other changes, not always well understood, ageing is associated with a substantial accumulation of body fat (110) and redistribution of fat from peripheral and subcutaneous sites to central and other sites, resulting in visceral and ectopic (hepatic and intramuscular) accumulation of adipose tissue. This is reflected in increased waist circumference and abdominal obesity in older adults (111). According to the World Health Organization (WHO), obesity contributes substantially to the development of 44% cases of T2D (see also 1.3), 23% of heart diseases and 7-41% of all cancers (112).

The combination of age-related muscle loss (see 1.2.1) and fat gain can result in older people developing sarcopaenic obesity. Sarcopaenia has been defined as a skeletal muscle mass two standard deviations below the sex-specific reference for a young, healthy population (49) while obesity is usually defined as a BMI greater than 30 kg/m² (113-115). Sarcopaenic obesity is the combination of both and is associated with greater adverse effects than either obesity or sarcopaenia alone including metabolic diseases, disability, increased institutionalization, inflammation and all risk mortality (116-118).

Lifestyle interventions (diet and exercise) are the first line of treatment for obesity followed by pharmaceutical interventions and/or bariatric surgery (119, 120). Increasing physical activity promotes weight loss (121). However, exercise-induced weight loss is often not dramatic and marked increases in exercise may not be

possible in older people with mobility limitations due to sarcopaenia, arthritis and other medical conditions (53). Energy restricted diets, preferably rich in fibre and micronutrients, along with exercise may be beneficial. When body weight is lost deliberately in response to energy restricted diets, lean tissue, including skeletal muscle, is lost along with adipose tissue (122). Given their age-related reduced muscle mass, older people are therefore at increased risk of developing adverse effects from reduced energy intake, whether it be involuntary or deliberate as part of a weight loss diet. Protein supplementation has been shown to have a role in preventing and treating loss of muscle in older people (3, 8, 123). It has been hypothesized that there is a reduction in the risk of frailty with higher protein intake by increase in muscle anabolism, and improvement in muscle quality (124).

While protein is the most satiating macronutrient in young adults and may aid weight loss, the effect of proteins, including whey, to suppress appetite and reduce food intake is reduced in older, non-obese, people (see 1.2.2). The interacting effects of ageing and obesity on appetite and feeding behaviour are not well understood.

1.2.4 Questions about effect of whey on nutritional factors addressed in this thesis

1. Duration of the effect of whey on appetite and feeding beyond the next meal in healthy younger and older men (chapter 3)
2. Effect of whey on appetite and feeding behaviour in obese, younger and older men participants (chapter 4)

1.3 Effect of whey protein ingestion on glucose metabolism

1.3.1 Type 2 diabetes mellitus (T2D)

1.3.1.1 Type 2 diabetes mellitus: Demographics and impact

The prevalence of T2D in most countries has increased over recent decades as their populations have become older and heavier. T2D accounts for about 85% of all diabetes and is the most rapidly growing chronic disease in Australia (125). According to the International Diabetes Federation, in 2015 the number of people with diabetes worldwide was 415 million, and this is expected to rise to 642 million by 2040 (126). Older people with T2D are at increased risk of cognitive and functional impairment, depression, falls and other diabetes related comorbidities (127, 128). A number of factors contribute to the increasing incidence (and hence prevalence) of T2D with increasing age. These include declining insulin secretion, increased insulin resistance, hormonal impairment, inflammatory and genetic factors, increased oxidative stress and increased use of medication promoting hyperglycaemia (e.g. glucocorticoids). Fasting and postprandial hyperglycaemia is a characteristic feature of T2D (129).

1.3.1.2 Management of T2D: Focus on dietary measures

Due to the complexity of factors associated with poor glycaemic control in T2D, achieving optimal levels of glycaemic control is challenging (130). Lifestyle measures are the first line treatment and should be continued even after medications are started, if needed.

Medications include a variety of oral agents, usually used first, then injected

medications in the form of GLP-1 receptor agonists and/or insulin. Oral metformin is the most widely used first line medication and works by several mechanisms including a reduction in hepatic glucose production; sulphonylureas (for example gliclazide, glipizide) increase insulin secretion; DPP-IV inhibitors (linagliptin, saxagliptin, sitagliptin) and GLP-1 receptor agonists (exenatide, liraglutide, lixisenatide, dulaglutide, semaglutide) increase insulin secretion and reduces glucagon secretion; sodium-glucose cotransporter-2 (SGLT-2) inhibitors (dapagliflozin, empagliflozin) inhibit renal glucose reabsorption, whereas insulin (short acting, immediate-acting and long acting) increases glucose disposal and reduces hepatic glucose production (131).

Lifestyle measures, particularly modifications to diet and increased physical exercise remain important treatment for people with T2D. Recommendations for the dietary management of T2D vary between recommending bodies, but generally include an emphasis on energy intake reduction to promote weight loss in overweight people with T2D, and reduction in the intake of unprocessed, high glycaemic index carbohydrates. They contain little mention of dietary protein intake (132).

The multiple determinants of postprandial glycaemia include pre-prandial blood glucose concentrations, the rate of gastric emptying, meal composition, carbohydrate absorption in the small intestine, insulin and glucagon secretion, incretin release and hepatic and peripheral glucose disposal (133). Protein ingestion alone usually has little effect on blood glucose concentrations, due to counteracting stimulatory effects on release of both insulin and glucagon (134, 135). However, when combined with carbohydrates, protein ingestion has been shown to reduce blood glucose levels significantly compared to ingestion of the carbohydrates alone, in younger and

middle-aged adults (86, 95, 136). Ingestion of protein appears to mainly stimulate insulin secretion by direct effects on the pancreas (137, 138). Oral protein ingestion also stimulates the gastrointestinal release of the incretin hormones glucagon-like peptide 1 and gastric inhibitory protein, which in turn stimulate insulin release and inhibit glucagon secretion. GLP-1 in particular stimulates the beta cell function, augment insulin release and regulates gastric emptying rate particularly via the vagal afferents that send signals to the brain. In addition, the release of gut peptides such as CCK and PYY is also stimulated by ingestion of whey protein, with the effect of delaying gastric emptying and regulating the gastrointestinal transit of food. Carbohydrates stimulate insulin secretion via sodium-glucose cotransporter-1 (SGLT-1) and intestinal “sweet taste” receptors (139). Variations in the rate of gastric emptying have an impact on postprandial glycaemia in people with and without T2D. Ageing is associated with modest slowing of gastric emptying thereby lower postprandial glucose excursions, while abnormally delayed gastric emptying may be present in people long standing diabetes, particularly if poorly controlled. In contrast, gastric emptying may be more rapid in people with newly developed and well-controlled. The rate of gastric emptying influences postprandial blood glucose concentrations (see 1.5.1). Due to the ability of insulin to cross the blood brain barrier, increased brain insulin concentrations of insulin after protein ingestion may also play a part in its appetite suppressant effects (140) and possibly reduce hepatic glucose production via the brain-liver axis (141). As explained later co-ingestion of proteins such as whey with carbohydrates can result in a more than additive effect on the increase in circulating insulin concentrations (see chapter 5 and 6). This may be because some amino acids act as substrates in the Krebs cycle to create ATP, which acts directly on the beta cells to increase their insulin secretory response to glucose (142).

Previous studies in young and middle-aged adults with and without T2D have shown that co-ingestion of protein with carbohydrate reduces the postprandial increase in blood glucose after carbohydrate ingestion alone with marked protein-induced increases in circulating insulin concentrations (143, 144). Nuttall reported a significant rise in insulin concentration with co-ingestion of 50 g protein with 50 g of carbohydrate as compared to glucose or protein alone ($247 \pm 33 \mu\text{U} \cdot \text{h/ml}$ vs 97 ± 35 , $83 \pm 19 \mu\text{U} \cdot \text{h/ml}$) and the plasma glucose area above the baseline following a glucose meal was reduced 34% when protein was given with the glucose in 9 men with T2D with a mean age of 61 years (68). Similar results have been reported by other studies. However, those studies largely used casein or beef protein (145, 146), rather than whey, our protein of interest. Frid et al. (136) studied the effect on postprandial glucose and insulin concentrations of adding 27.6 g of whey protein to high glycaemic index meals at breakfast and lunch in non-elderly participants with T2D aged 27–69 years and found a whey induced increase in the insulin responses at breakfast and lunch of 31% and 57% respectively, with an associated whey-induced reduction in blood glucose concentrations.

Ma et al. (147) gave 55 g of whey protein in meal or as a preload along with carbohydrate to 8 participants with diet-controlled T2D with a mean age of 58 years and showed a substantial reduction in postprandial glycaemia. The incremental areas under the curve (iAUC) for insulin, GLP-1, GIP, and CCK were significantly greater and for glucose significantly lower when whey was co-administered. Because of this effect to reduce blood glucose concentrations ingestion of whey with or just before a meal has been proposed as a possible strategy in the management of T2D (147-149). Therefore, it is not known if the above effects of whey on postprandial glycaemia in people with and without T2D persist into old age, particularly over 70

years - the age group with the highest prevalence of diabetes – or if 30 g of whey protein (a muscle protecting dose) has this effect. This led our group conduct studies to determine the effect of combining 30 g whey protein with carbohydrate in men > 65 years with and without T2D.

1.3.2 Questions about effect of whey on glucose metabolism addressed in this thesis

1. Impact of ageing on the effects of whey on glucose metabolism in healthy younger and older men (chapter 5)
2. Effect of whey on glucose metabolism in older men with and without T2D (chapter 6).

1.4 Effects of whey on blood pressure and pulse rate in older people

1.4.1 Physiological cardiovascular responses to eating

Ingestion of food has many physiological effects. The cardiovascular and metabolic responses to feeding are driven both peripherally and centrally (150). Meal ingestion results in blood flow redistribution to the gut, splanchnic blood pooling and a rise in mesenteric blood flow. This diversion of blood from the systemic circulation to aid nutrient absorption and digestion results in a blood pressure (BP) fall unless there is an adequate compensatory increase in pulse rate and hence cardiac output (151). Usually these compensatory mechanisms work well enough to prevent excessive post-prandial BP falls in most young people (152-154). However, these compensatory responses to food ingestion are less effective in older people who experience greater food-induced BP decreases than younger adults (155).

1.4.2 Postprandial hypotension

An excessive fall in BP after a meal has been termed postprandial hypotension (PPH) and defined as a decrease in SBP of > 20 mm Hg within 2 hr of a meal (156). The effectiveness of cardiovascular compensatory responses to food intake declines with advancing age and PPH is not uncommon in older people (157-159). The prevalence of PPH has been reported to be 24% to 38% in community-dwelling elderly and residential facilities (160, 161), 20% to 91% in hospitalized geriatric patients (162, 163), 40% in people with T2D (164, 165), and 40% to 100% in people with Parkinson disease (166). While the majority of people with PPH are probably asymptomatic (166), when severe, PPH can lead to nausea, light headedness, weakness, dizziness, falls, fractures, syncope, and even cerebrovascular events (strokes), thus increasing morbidity and mortality rates in older people (158, 160, 167, 168).

The pathophysiology of PPH is multifactorial. Causative factors, which vary between individuals, include baroreceptor dysfunction, gastric distension, variations in the rate at which nutrients are delivered from the stomach to the small intestine, splanchnic blood flow and release of vasodilatory hormones, and altered neural and hormonal mechanisms. After meal ingestion, secretion of neurotensin and insulin may cause excessive vasodilation, particularly of vessels supplying the gastrointestinal tract thus lowering cardiac output (169-171). In healthy young adults, excessive BP decreases after food ingestion are prevented by increased vasoconstriction of blood vessels in the leg and other compensatory mechanisms which act to increase cardiac output mainly by increasing heart rate (172). On average these compensatory mechanisms become less effective with increasing age

(172).

Other factors affecting postprandial BP responses include meal composition (173-175) and temperature, the timing of food intake and the posture of the participant after food ingestion (155). Smaller meals are associated with less diversion of blood to gut, thereby requiring less cardiovascular compensation. In patients with chronic autonomic failure, consumption of 3 large meals has been associated with lower postprandial BP as compared to 6 smaller meals distributed throughout the day SBP; lying, sitting and standing; large vs. small; 131 vs. 151 mm Hg, 109 vs. 124 mm Hg, 89 vs. 103 mm Hg, $P = 0.005$; DBP: 76 vs. 90 mm Hg, $P = 0.02$, 66 vs. 78 mm Hg, $P = 0.07$ and 50 vs. 66 mm Hg, $P = 0.06$ (176). The magnitude of postprandial BP changes varies with the nutrient composition of the food eaten (173, 174). Carbohydrates, particularly, glucose and sucrose, lower BP more than the other macronutrients (168, 177). Carbohydrate rich meals cause greater activation of the sympathetic nervous system and decrease in peripheral resistance than protein rich meals (178). Nevertheless, in older participants all macronutrients appear to have the capacity to decrease BP when given orally or intraduodenally (179-181).

Only a limited number of studies have been performed, with somewhat conflicting results, on the influence of fat and protein ingestion on postprandial BP and these have produced inconsistent results (173, 174, 182-186). Observational studies indicate that increased dietary protein intake may be associated with reductions in BP (187-190). Milk protein has been associated with reduced BP (191). A number of clinical trials have been carried out to evaluate the effect of milk proteins from whole foods and supplements on BP (192-195). A reduction of 3.33 mm Hg in SBP and 1.08 mm Hg in DBP has been observed with milk proteins (196). A randomized double blind controlled trial was conducted to determine the effect of dietary proteins

on BP (PROPRES) and investigate whether a moderately increased protein intake (+10% of energy) compared with iso-calorically increased maltodextrin intake for 4 weeks lowered BP in 99 overweight individuals aged 20–70 years with grade 1 hypertension (SBP of 130–159 mm Hg and/or DBP of 85–99 mm Hg). Participants were supplemented for 4 weeks with 3x 20 g protein per day (20% pea, 20% soy, 30% egg and 30% milk-protein isolate) or control (with 3 × 20 g maltodextrin/d). Office SBP and DBP were 4.9 ± 1.7 mm Hg and 2.7 ± 1.3 mm Hg (both $P = 0.05$) lower in the protein group and daytime SBP was 4.6 ± 1.7 mm Hg lower in the protein group ($P = 0.006$), with no daytime DBP difference between groups ($P = 0.37$). While this result suggests a BP lowering effect of protein no differences in postprandial BP responses between high-protein and high-carbohydrate meals have been found in other studies (181, 197).

The specific effects of whey protein on BP have been studied, but not in elderly people. It has been proposed that bioactive peptides in whey, particularly lactokinins are involved in inhibition of renin-angiotensin aldosterone system by binding with the zinc fingers on the angiotensin converting enzymes (ACE) molecule resulting in the inability of ACE to further cleave angiotensin I into angiotensin II, thus lowering BP (198, 199). Results of studies with whey in non-elderly adults are inconsistent (200) but mostly indicate that it does lower post-prandial BP. Fekete et al. showed a significant reduction in SBP and DBP with 28 g of whey protein/day as compared to 28 g of calcium caseinate/day or 27 g of maltodextrin/day for 2 days in 38 participants for 8 weeks (191). Fluegel et al. also found that whey beverage consumption for 6 weeks significantly decreased SBP and DBP in adults with elevated BP (201). In a randomised controlled trial, a significant decrease in SBP was observed with a diet containing whey protein (40% carbohydrate: 15% mixed

protein: 15% whey protein: 30% fat) as compared to mixed protein (40% carbohydrate: 30% protein: 30% fat) or control (55% carbohydrate: 15% protein: 30% fat) (65) in 18 healthy adults. However, Lee indicated that the regular consumption of a milk drink containing whey peptides for 12 weeks has no effect on BP in mildly hypertensive individuals (202). Given this, it is possible that the BP changes reported in our studies in a single day treatment would not persist and might even be reversed with longer treatment. It is not clear whether whey exerts hypotensive effect in older men and if so if those effects are dose-dependent.

Various treatments have been proposed for people experiencing adverse effects of PPH. These include medications such as alpha glucosidase inhibitors (acarbose) which delay the digestion of carbohydrates, and non-pharmacological/lifestyle measures such as postprandial exercise, consumption of small frequent meals, and measures to slow the gastric emptying of nutrients into the small intestine such as increased water drinking to increase gastric distension (176, 203-208). A better understanding of the effects of ingestion of proteins including whey, when ingested alone and with other macronutrients, on postprandial BP, is likely to help decisions about the dietary treatment of PPH when present.

The effect of whey protein, administered alone and with other macronutrients, on postprandial BP in older people has not been clarified. The relative effects on BP of whey when administered alone and in combination with other macronutrients are also unclear. If oral ingestion of proteins, including whey has the ability to lower postprandial BP in older people, who are more at risk than younger people of PPH and its adverse consequences, the use of whey supplements for muscle-preservation or other reasons, by older people may be associated with adverse excessive BP

lowering effects. Therefore, we conducted studies to see the effect of 0 g, 30 g and 70 g whey protein alone or mixed with other macronutrients on SBP, DBP and HR in younger and older men.

1.4.3 Questions about effect of whey on blood pressure and pulse rate addressed in this thesis

1. The impact of ageing on BP responses to whey protein (0 vs 70 g in older vs younger men, study in chapter 7).
2. Having found that 70 g decreases BP in older men (chapter 7), does a lower, probably more “physiological dose” (30 g) have the same effect (study in chapter 8).
3. The effect of whey, ingested alone and together with other macronutrients on BP and pulse rate in older men (chapter 9).

1.5 Gastric emptying

1.5.1 Control of gastric emptying

Gastric emptying involves a coordinated set of activities of the proximal (fundus and gastric body) and distal stomach (pylorus and antrum) and duodenum. There is a relaxation of proximal stomach as meal intake is commenced, as it is a reservoir for solid foods. The distal stomach then grinds the solid foods to small particles by contractions thereby creating flow across pylorus (70, 209). The gastric emptying rate is regulated by the interstitial cells of Cajal as it is involved in antrum contractions and neuromuscular transmissions (210). Gastric emptying rate is influenced not only by the physical characteristics of the meal such as whether it is solid, semi-solid, liquid, viscosity, volume but also the chemical composition such

as nutrient and energy content, osmolarity and pH (110).

In this thesis, gastric emptying was measured in the studies described in chapters 3, 4 5, 6, and 7. These measures were obtained because the rate of gastric emptying has been shown in previous studies by our group and others to be associated with the outcomes of interest in the studies in this thesis.

Variations in the rate of gastric emptying are known to be associated with

1. Appetite and feeding behaviour (e.g. hunger, fullness, satiety) – Ingestion of nutrients triggers a series of mechanisms both intragastric (gastric distension) and small intestinal (changes in antropyloroduodenal motility and the release of appetite-regulating hormones) that regulates appetite and feeding behaviour (165). When ingested, nutrients interact in the small intestine resulting in generation of neural and humoral signals which controls gastric emptying (211-213). Secreted by stomach, ghrelin stimulates appetite and energy intake. Small intestine secretes PYY CCK and GLP-1 that suppresses food intake. The delayed proximal gastric distention to a meal observed in the elderly might contribute to early satiation (91). Ageing is associated with reduced proximal gastric distension, and greater distension antral area, resulting in slightly slower gastric emptying, that favours reductions in energy intake (25, 214, 215).
2. Blood glucose concentrations after nutrient ingestion – The rate of gastric emptying is a critical determinant of postprandial blood glucose concentrations (216, 217). In healthy individuals the stomach empties at the rate of 1-4 kcal/min after meal ingestion. Rapid gastric emptying is associated with a greater, more rapid increase in post-prandial blood glucose concentrations. Ageing is associated with modest slowing of gastric emptying, while gastric emptying may

be more rapid in people with well-controlled T2D and delayed gastric emptying may be present in long-standing, poorly controlled T2D (149, 215, 218-223). In people with poorly controlled and long-standing diabetes (HbA1c > 8.5% and > 10 years), gastric emptying of solid meal is delayed in 30-50% of participants (220, 224-226).

3. Greater post prandial BP decreases have been associated with more rapid gastric emptying (227, 228). The fall in BP after a meal is greater when the rate of gastric emptying is faster in both healthy older participants (13, 14, 166, 229) and people with T2D (163, 227).
4. Ageing – ageing is associated with modest slowing of gastric emptying of both solid and liquid meals (230) and this slowing may potentially contribute to reduce food intake in older people (214). With ageing, there is a loss of interstitial cells of Cajal and enteric neurons. Slowing of gastric emptying is due to increased pyloric pressure waves and reduction in duodenal pressure wave (210). Various other factors such as higher fasting and postprandial CCK and GLP-1 concentrations and higher satiating effect of intraduodenal glucose infusion are responsible for altered nutrient response with ageing (25, 26, 28, 231, 232). Perceptions of gastric distension are diminished in the healthy elderly, indicating a reduction in visceral sensitivity (91).

1.5.2 Measurement of gastric emptying rate

The gastric emptying rate can be measured by a number of methods and is often expressed as the time taken for 50% of the ingested meal to leave the stomach (218).

Methods used include scintigraphy, breath tests, ultrasound, paracetamol absorption, radiology, magnetic resonance imaging (MRI), computerized tomography (CT) scan, manometry and two-dimensional (2D) or three-dimensional (3D) ultrasonography (70, 165, 233). Scintigraphy is usually considered the “gold standard” and allows measurement of the gastric residue of both solid and liquid food using dual isotopes. Due to the need for complex equipment including a gamma camera and radiation exposure (70, 230) other methods, validated against scintigraphy are often used.

In the studies described in this thesis, a non-invasive method of measuring gastric emptying with a stable isotope breath test, ^{13}C octanoic acid was used. This technique has been validated against and has been shown to be of comparable accuracy to scintigraphy in the measurement of gastric emptying of both solid and liquid meals in people without (234) and with T2D (235). In the validation paper by Ghooos et al., a test meal with egg yolk labelled with ^{14}C octanoic acid was given to participants as scrambled eggs. It was also labelled with another marker $^{99\text{m}}\text{Tc}$ -albumin colloid. Simultaneous measurements of scintigraphy and breath test were performed on 36 participants. Breath sampling was performed over 4 h at 15-minute intervals. Two curves were obtained for each participant: the scintigraphic and the %dose/h curves. A strong correlation between coefficient of gastric emptying and the scintigraphic half-emptying time ($r = -0.88$); breath test half-emptying time and the scintigraphic half-emptying time ($r = 0.89$); and between the scintigraphy and breath test the lag phases ($r = 0.92$) was obtained.

It is possible to replace ^{14}C with ^{13}C in the octanoic acid breath test with results of similar accuracy (234). Studies in adults participants have demonstrated a close correlation between ^{13}C octanoic acid breath test and gastric emptying scintigraphy

($r = 0.74$) (236-238). This method has been used in previous published studies by our group and others (217, 239-241). Essentially the Carbon-13-isotope is emptied from the stomach and oxidized to $^{13}\text{CO}_2$ after transport to the liver via portal vein. The participant ingests a ^{13}C labelled standardized drink/meal. End-tidal breath samples are collected at different time intervals and labelled carbon dioxide content is measured using isotope ratio mass spectrometry in each breath sample. T_{50} ($t_{1/2}$) of ^{13}C excretion is further determined using the Wagner-Nelson method (242).

The following chapter provides further insights into the rational use of whey protein supplements, particularly in older people, with details extending to include the evidence for benefits of nutritional supplements, potential adverse effects, and relevance of gastrointestinal mechanisms along with the role of gut hormones (chapter 2).

**CHAPTER 2: RATIONAL USE OF PROTEIN SUPPLEMENTS
IN THE ELDERLY- RELEVANCE OF GASTROINTESTINAL
MECHANISMS**

Chapman I, Oberoi A, Giezenaar C, Soenen S

Published in *Nutrients*

2021

STATEMENT OF AUTHORSHIP

Title of the paper	Rational use of protein supplements in the elderly- relevance of gastrointestinal mechanisms.
Publication status	Published
Publication details	Chapman I, Oberoi AO, Giezenaar C, Soenen S. <i>Rational use of protein supplements in the elderly- relevance of gastrointestinal mechanisms</i> . <i>Nutrients</i> 2021; 13: 1227. doi: doi.org/10.3390/nu13041227.

Candidate	Avneet Oberoi		
Contribution	Contributed to the overall design of the manuscript, literature review, drafting and revision of the manuscript.		
Overall percentage	50%		
Certification	This paper reports on original research I conducted during the period of my Higher Degree by Research candidature and is not subject to any obligations or contractual agreements with a third party that would constrain its inclusion in this thesis. I am the primary author of this paper.		
Signature		Date	October 2022

Principal Author***Co-Author Contributions***

By signing the Statement of Authorship, each author certifies that:

- i) the candidate's stated contribution to the publication is accurate (as detailed above);
- ii) permission is granted for the candidate to include the publication in the thesis; and
- iii) the sum of all co-author contributions is equal to 100% less the candidate's stated contribution.

Name of Co-Author	Ian Chapman		
Contribution	Conception, design, drafting and revision of the manuscript.		
Signature		Date	October 2022
Name of Co-Author	Caroline Giezenaar		
Contribution	Drafting and revision of the manuscript.		
Signature		Date	October 2022
Name of Co-Author	Stijn Soenen		
Contribution	Conception, design, drafting, revision of the manuscript and overall responsibility of the paper.		
Signature		Date	October 2022

2.1 Abstract

Protein supplements are increasingly used by older people to maintain nutrition and prevent or treat loss of muscle function. Daily protein requirements in older people are in the range of 1.2 gm/kg/day or higher. Many older adults do not consume this much protein and are likely to benefit from more. Protein supplements are probably best taken twice daily, if possible soon after exercise, in doses that achieve protein intakes of 30 gm or more per episode. It is probably not important to give these supplements between meals, as we have shown no suppressive effects of 30 gm whey drinks, and little if any suppression of 70 gm given to older participants at varying time intervals from meals. Many gastrointestinal mechanisms controlling food intake change with age but their contributions to changes in responses to protein are not yet well understood. There may be benefits in giving the supplement with rather than between meals, to achieve protein intakes above the effective anabolic threshold with lower supplement doses, and have favourable effects on food-induced blood glucose increases in older people with, or at risk of developing type 2 diabetes mellitus; combined protein and glucose drinks lower blood glucose compared to glucose alone in older people.

2.2 Introduction

This review will focus on appetite, feeding and gastrointestinal responses to protein ingestion in older people. As a background, the high rates of under-nutrition, sarcopenia and probably also sub-optimal dietary protein intake in older people will be outlined, in support of the logical use of protein or protein-rich supplements by older people. The type, amount and timing of such supplements will be covered and possible side effects of these supplements. Results of studies with whey protein by our group will be used to illustrate a number of gastrointestinal and cardiovascular responses to protein ingestion and to support some preliminary recommendations about beneficial use of protein supplements by older people. A number of issues remain unresolved.

2.2.1 Ageing, weight loss and undernutrition

Healthy ageing is associated with a physiological reduction in appetite and food intake, the so-called “anorexia of aging” (243); people over 80 years consume about 30% less energy per day than those in their 20s (71). Consequently, after about 65 years in developed countries body weight tends to decrease (71). This age-related weight loss, particularly when substantial and involuntary, has been associated with increased mortality (71). Adverse factors prevalent in older people can be superimposed on the age-related changes to cause pathological under-nutrition, with even greater associated increases in morbidity and mortality (32). Rates of under-nutrition increase with age and loss of independence; it is present in up to 45% of community dwelling elderly people, 50-80% in hospital, and 80-100% in residential facilities (32). The major markers of existing or pending under-nutrition in older

people are low body weight (particularly Body Mass Index < 22 kg/m²); loss of weight, particularly when involuntary and > 5%; substantial loss of muscle mass which can lead to sarcopenia (see below); reduced muscle strength and function; reduced appetite and food intake; and frailty (32, 71).

2.2.2 Ageing and sarcopenia

The weight lost with increased ageing is disproportionately made up of lean tissue, particularly skeletal muscle, but also bone mass. In contrast, fat mass increases with ageing, tending to mask the extent of lean tissue loss (71). After approximately age 30 years about 5% of lean muscle mass is lost per decade (71). The loss of lean tissue, when large, can result in sarcopenia, an excessive and damaging loss of muscle. Sarcopenia is prevalent in older people, present in up to 50% of people over 80 years (244). Sarcopenia has functional adverse consequences, with increased morbidity due to falls, fractures, infections and other conditions, and increased mortality (245).

2.3 Prevention and management of under-nutrition and sarcopenia in older people

A greater appreciation of the high rates and significant adverse effects of undernutrition and sarcopenia, which often co-exist in older people, has led to attempts to prevent, and treat these conditions. Both exercise and nutritional measures have been shown to have benefits, particular when combined (246, 247).

2.3.1 Nutritional measures

Nutritional measures often start with encouragement and assistance to consume greater quantities of usual foods to maintain body weight, the so-called “Food First” approach (248). These measures may include providing meals at home, supervising food intake and/or having the person eat with others in aged care settings (to counteract reduced appetite and reduced spontaneous food intake), increasing the nutrient and energy density of the food, adding flavour boosters and fortifying the food with additional fats, protein and carbohydrates (248).

2.3.2 Nutritional supplements

While these “Food First” approaches may be sufficient, they may be difficult to implement due to cost of foods or time/staffing/convenience constraints, and not always successful. Consequently, nutritional supplements are increasingly recommended for, and used by older people, both as a means of increasing total energy intake (when harmful weight loss is a concern) and to increase protein intake. Mixed macronutrient supplements are used for the former, supplements of pure protein or very high in protein more specifically for the latter indication.

The use of such supplements offers the opportunity to more closely tailor the timing and composition of added protein intake to that what is likely to be most beneficial. It is often recommended that older people take nutritional supplement drinks between and well separated from regular meals so as not to reduce energy intake at those meals; maintenance of weight or even weight gain being a desirable outcome in the many older people with, or at risk of, under-nutrition.

Consequently specific nutritional supplements, usually commercial preparations, are used widely by older people. When the aim is to maintain body weight and general nutrition, rather than specifically enhance skeletal muscle and function, these usually take the form of a mixture of macronutrients in a drink containing 1-1.5 kcal/ml of protein, fat and carbohydrates. Frequently used preparations contain about 9-15gm protein (15-17% of total energy) in a 200-240 ml serve.

2.3.3 Protein nutritional supplements

When the aim is to preferentially prevent and/or reverse ageing-associated muscle loss and sarcopenia the use of such mixed macronutrient nutrient supplements may not be sufficient and pure protein or high protein supplements may be used.

2.4 Dietary protein requirements in older people

Recommended dietary protein intakes for adults of all ages, based on nitrogen balance studies, are 0.8 gm/kg body weight in most countries (249), with no adjustments for age or gender. There have been increasing calls, however, for the minimum recommended daily protein intake in older people to be higher than that (97, 250). One reason is concern that the traditional nitrogen balance study methods used to determine appropriate protein requirements in adults of any age underestimate true requirements, with requirements as determined by the newer indicator amino acid oxidation (IAAO) methods up to 25-75% higher (249). The minimal protein intake for healthy younger adults determined by IAAO is approximately 1.0-1.2 gm/kg/day (249).

Secondly, older people are likely to need higher dietary protein intakes than younger adults for the maintenance of good health, for a number of reasons. These include an age-related reduction in muscle anabolic response to ingested protein; while digestion and absorption of dietary protein is not apparently affected by ageing, there is evidence for an age-related reduction in the anabolic muscle response to ingested protein – anabolic resistance (see (250)). This is due to both a redistribution of ingested proteins away from the muscle to splanchnic tissues and a reduced anabolic effect on muscle of the amino acids that do reach the muscle. In addition, catabolic conditions associated with increased muscle breakdown and needs for dietary protein intake, become more prevalent with increasing age. These include chronic diseases such as obstructive airways disease, heart failure, renal failure, malignancies, inflammatory forms of arthritis and polymyalgia rheumatica, and acute conditions such as infections and cardiovascular/cerebrovascular events.

Although not conclusive, there is some evidence from balance studies that older people do have higher dietary protein requirements than young adults. One meta-analysis was reported to show approximately 6% higher protein intakes needed to maintain nitrogen balance in people 60 year and older compared to those younger (251), and another approximately 26% higher intakes needed in those over 55 years compared to those younger (252), although neither difference achieved statistical significance due to small participant numbers. Using the indicator amino acid oxidation method mentioned above to study small numbers (12 or less per group) of participants protein requirements have been reported as 1.24 gm/kg/day in men over 65 years (253), and 1.15-1.29 gm/kg/day in women over 65 years (254).

2.4.1 Do men and women have different dietary protein needs?

Women, on average consume less protein than men, in absolute terms due to lower body weight, and possibly also less compared to body weight. Women over 70 years in the 2003-4 US NHANES study consumed about 10% less protein per kg body weight than men of the same age, with 50% of these older women reporting daily protein intakes ≤ 0.9 gm/kg ideal body weight/day, compared to only 25% of older men (255). These reduced intakes in women are probably in line with reduced needs. Results of several meta-analyses of nitrogen balance studies indicate that adult women need approximately 10% less dietary protein per kg body weight than men to maintain nitrogen balance (251, 252), probably due to lower muscle mass relative to body weight. Nevertheless, there were few older participants in the studies examined in those meta-analyses, and not all studies show lower requirements in women (256). While protein needs/kg body weight of older women may be lower than those of older men, therefore, the difference does not appear to be great. In the interests of simplicity and the absence of evidence that higher protein intakes, if achievable, do much harm (see below) it seems reasonable to maintain gender-neutral protein intake recommendations for older people at this time.

2.4.2 Summary recommendations for dietary protein intake in older people

2.4.2.1 Total Daily Protein Intake

For reasons outlined above, recent recommendations for total daily protein intake in healthy older people are usually in the range of 1-1.5 gm/kg per day. The European PROT-AGE study group for example, has recommended 1.0-1.2 gm/kg dietary protein for healthy older adults (97). Others have suggested that even higher intakes

(> 1.2 gm/kg/day) are needed for maintenance of muscle mass and function (250). Even higher intakes than these are likely to be needed at times of catabolic stress due to acute or chronic illness, with the PROT-AGE group recommending 1.2-1.5g/kg/day for those with acute or chronic illness, and up to 2.0 gm/kg/day for those with severe illness or injury or with marked malnutrition (97).

2.4.3 How many older people need supplements to reach the recommended protein intake?

It is unclear how many older adults consume less than the recommended protein intake (either current RDA or the newer recommendations), but probably a substantial proportion. The US NHANES study of 2003-4 reported dietary protein intake substantially lower in healthy older than young adults; in people over 70 years mean total protein intake was 64.7 gm with a mean intake of 1.0 gm/kg ideal body weight, 27% and 23% respectively below reported intakes in adults 19-30 years (255). Over 75% of people over 70 years reported intakes of less than 1.2 gm/kg ideal body weight (255). Similarly, in the Quebec NuAge study the mean protein intake of older participants was very close to 1 gm/kg/day (256).

2.4.4 Possible adverse effects of protein supplements

Some concerns have been raised about increasing the dietary protein intakes of older people over current recommended levels. These include:

2.4.4.1 Renal effects

Renal function declines with increasing age (257). High protein intakes can increase renal filtration and accelerate the progression of established renal disease; restricted protein intakes of 0.3-0.8 gm/kg/day have been shown to delay the progression of established renal disease and are often prescribed for this indication. The adverse effects of high dietary protein intakes on impaired renal function appear to be increased in those with diabetes mellitus, hypertension or obesity. It is possible, therefore, that increasing the dietary protein intake of older people, whose renal function has already undergone age-related declines, to levels of 1.2 gm/kg/day or higher may induce renal impairment or accelerate the progression of pre-existing renal disease. While this is possible, it is not established that older people with relatively good renal function and without significant risk factors are at such risk. Evidence does not support a relationship between increased dietary protein content and a decline in renal function (258) and high protein diets undertaken for up to 2 years have not been shown to impair renal function in otherwise healthy people (259). Advanced age alone does not, therefore, appear to be a reason to avoid increased dietary protein intake when indicated, although caution should be exercised in those with pre-existing renal impairment or risk factors for renal function deterioration.

2.4.4.2 Bone effects

Previous suggestions that increased dietary protein intake may have adverse effects on bone health by increasing bone mineral loss and increasing the risk of fractures, have not been supported by more recent studies (249, 260), which have, if anything,

shown beneficial effects of dietary protein on bone health.

2.4.4.3 Post-prandial hypotension

Ingestion of nutrients leads to redirection of blood flow from other organs to the splanchnic circulation to aid digestion. As a result blood pressure can drop. This is largely prevented in young, healthy adults by an increase in heart rate and other compensatory mechanisms, but in older people, these are less effective. As a result, older people have greater food-induced BP decreases than younger adults (155). In some this can be excessive and lead to symptoms of dizziness and in some cases falls and cardiovascular events (261). The excessive drop in BP after food ingestion has been termed post-prandial hypotension and defined as a decrease in systolic BP of 20 mm Hg or more within 2 hours of food ingestion (262). Ingestion of all three macronutrients, alone or together causes post-prandial BP falls in older people. Available evidence indicates that carbohydrate and protein have equivalent BP lowering effects, although the BP decreases occur sooner after carbohydrate than protein and fat (263). We have recently reported substantial systolic BP decreases after ingestion of a 70 gm whey protein drink by healthy, older men, with 58% having a decrease of 20 mg Hg systolic or more within three hours of protein ingestion, with maximum decreases occurring between two and three hours after the drink (264). In contrast, maximum decreases after carbohydrate alone, or mixed drinks, appear to occur earlier. Seventy grams of whey is a high dose, higher than is generally recommended or likely to be ingested by older people (see below), and the hypotensive effects of protein and other macronutrients do seem to be at least partly dose-responsive (265). Nevertheless high-energy nutrient supplement drinks decrease BP in older people and the hypotensive effects of pure or high protein drinks

may be quite prolonged and possibly greater in those already on antihypertensive medications. Appropriate advice and precautions are indicated after such drinks, particularly in at-risk individuals.

2.4.5 Evidence for benefits of nutritional supplements in older people

A detailed review of this matter is beyond the scope of this review. Study methods and results are variable. Not all studies show benefits (256). Nevertheless, in our view, a number of conclusions can be drawn; the use of such supplements is generally safe (see above) and relatively easy to implement. Few, if any adverse effects of such supplements have been reported; use of these supplements by older people improves nutritional intakes and is associated with weight gain and an increase in lean body mass in many cases, and in some cases improvements in muscle function, hospitalisation rates and even death rates (266). The greatest benefit of taking nutritional supplements are obtained by the most undernourished older people (266).

For these reasons, and because many older people appear to have a suboptimal dietary protein intake, protein or protein- rich supplements are increasingly recommended to older people.

In summary, there is evidence that:

1. Older people have higher dietary protein needs than young adults, particularly if they have lost weight and/or are undernourished, are sarcopenic, or have acute or chronic medical conditions which contribute to muscle catabolism. These requirements are in the range of 1.2 gm/kg/day or higher.

2. Many, (probably the majority of those > 70 years) older people do not consume these minimum protein requirements. It can be difficult for older people to increase their dietary protein intake by increasing their intake of usual foods due to anorexia and the cost of high protein foods.
3. Protein supplements increase muscle mass and strength, and may also reduce morbidity and mortality, particularly in undernourished and/or sarcopenic older people.

2.4.6 Does the type of protein or amino acid in the supplement matter?

Yes, it probably does. Not all ingested proteins are the same, particularly in terms of their anabolic effects on skeletal muscle (see (249) for review). Available evidence indicates that branched chain amino acids, particularly leucine, are the most effective amino acids in stimulating muscle protein synthesis (MPS). Consistent with this, in one longitudinal study older people (> 65 years) with dietary leucine intakes in the upper quartile had preservation of lean body mass over 6 years whereas those with intakes in the lowest quartile had loss of lean body mass (267). The extent of MPS is proportional to the peak leucine plasma concentration after protein ingestion (268).

Animal derived dietary proteins appear to be stronger in stimulating MPS than plant-based proteins and milk-based proteins in particular are a good source of leucine containing proteins (249). Whey protein, obtained from milk in the cheese making process, is high in leucine, about 10-15%, and more rapidly absorbed than casein, factors which may contribute to its greater stimulatory effect than casein on MPS in older men (7). Maximum stimulatory effects on MPS probably occur after ingestion of 2-2.5 gm leucine in young and middle aged adults, which little additional effect

of higher intakes. Ingestion of 20gm whey protein concentrate at one time is probably sufficient to optimise MPS in young and middle aged adults (269). In contrast, because of anabolic resistance, older adults appear to need 30-35 gm or more of whey to achieve similar anabolic effects (270).

The effects of protein supplements to increase muscle mass and strength and to have functional benefits, appear greatest in those most at risk ie malnourished, sarcopenic, or at risk of sarcopenia. Several interventional studies have demonstrated that combined dietary supplements of whey protein and leucine increase muscle mass and strength and improve function in older sarcopenic adults see (249). A recent meta-analysis reported increased lean body mass without an increase in strength in sarcopenic older adults taking 2-7.8 gm per day of leucine supplements/day (271). It is difficult to separate the effects of leucine supplements from those of other amino acids or proteins such as whey when they are given in combination. It may be, particularly in the case of already leucine rich whey protein supplements, that they have benefits in addition to those of leucine, and that when a sufficient amount is ingested, further leucine fortification has little further benefit.

2.4.7 How much protein should there be in the supplement?

Owing to age-induced “anabolic resistance” older people probably require 30-45 gm protein per serve to stimulate muscle protein synthesis after that meal, whereas lower doses (≤ 20 gm) are sufficient in young adults (see above and (84)). The consumption of 1-2 daily meals with protein content of 30 to 45 g may be an important strategy for increasing and/or maintaining lean body mass and muscle strength with ageing (84). There is evidence that older men and women with more-evenly mealtime

distributed protein intakes have higher muscle strength irrespective of their total protein intake (256). It seems reasonable, therefore, that if protein supplements are being used for their effects on muscle, they be used in a way that provides at least 1.2 gm/kg/protein per day (food plus supplement) with at least two protein intake episodes/day (food plus supplement) containing at least 30 gm protein each. As the post protein BP drop, which could be potentially harmful, appears to be dose-responsive, it is also likely that smaller, twice daily or even more frequent, doses of protein supplement will have more beneficial effects on BP than once daily larger doses.

2.4.8 Timing of protein intake relative to exercise

Both protein ingestion and resistance exercise independently stimulate muscle protein synthesis in older people, although the response to both is blunted in the elderly (272, 273). These two stimuli have synergistic anabolic effects on MPS, particularly when the protein is ingested soon after the resistance exercise (274). While ingestion of approximately 20 gm of a high-quality protein is sufficient to maximize skeletal muscle protein synthesis rates during recovery from resistance-type exercise in younger adults, doses up to 40 gm or possibly even higher, are needed in older adults (272, 273).

2.5 Gastrointestinal responses to protein ingestion: Effects of ageing

Appetite and food intake in free living humans are dependent on a complex interplay of environmental factors and central and peripheral physical mechanisms. The latter mechanisms include intra-gastric and small intestinal sensory and motor functions

and their interactions (71). Their study has been a focus of our group. We have used whey protein drinks and intra-duodenal infusions of whey protein to investigate peripheral responses to protein in young and older adults and have identified a number of age-related differences between these age groups in gastrointestinal responses to protein ingestion. The doses of oral whey used have mainly been 30 gm and/or 70 gm, which is of significance as 30gm appears to approximate the amount required in older people to stimulate muscle protein synthesis (see above), while higher doses are used by some older people.

Older people have reduced appetite compared to young adults and consume less food. There is no change, however, with ageing in the preference for particular macronutrients i.e. the percentage of total energy ingested as protein does not seem to change with increasing age (275).

2.5.1 Effect of ageing on appetite and feeding responses to whey protein

There is evidence that protein is the most satiating of the macronutrients in young adults (276), although this effect may be less in women than men (17). Our studies have demonstrated that healthy ageing is associated with a marked and significant reduction in the satiating effects of whey protein, administered by both intra-duodenal (9) and oral routes (15, 277).

2.5.1.1 Intra-duodenal whey

Healthy ageing is associated with a marked reduction in the suppressive effect of whey protein administered directly into the duodenum, on appetite and subsequent

food intake. Sixty minute intra-duodenal infusions of 8 gm (0.5 kcal/min), 24 gm (1.5 kcal/min) and 48 gm (3 kcal/min) of whey had a dose responsive suppressive effect on subsequent ad libitum food intake in young men, whereas older men experienced suppression of food intake only after the 48gm infusion (~33% vs 17% suppression by 48 gm whey in young vs older, $P < 0.05$) (9). Baseline hunger ratings were lower in the older than young men and were suppressed less by the protein infusions in the older than young men, consistent with the reduced effects of intra-duodenal fat and carbohydrate infusions on appetite in older people (27).

2.5.1.2 Oral whey

We have reported that healthy, non-obese, young men but not women experience significant suppression of hunger ratings and ad libitum food intake three hours after 30 gm and 70 gm whey protein drinks (12, 15, 17). In contrast, and consistent with their reduced responses to intra-duodenal whey, healthy, non-obese, men and women people over 65 years, experience little reduction in hunger and no suppression of ad libitum food intake three hours after either 30gm or 70gm whey drinks (12, 14-16, 278).

This age-related reduction in the satiating effects of whey drinks is observed in our studies largely irrespective of the timing of the whey drink relative to later ad libitum food intake at test meals. Thirty gram whey protein drinks do not suppress appetite ratings or subsequent ad libitum food intake in healthy older people immediately after the drink, or 35 minutes, 1 hour, 2 hours, 3 hours, 265 minutes and 510 minutes after the drink (14, 15, 98, 277, 278). In only one study have we detected any reduction in subsequent food intake by older people after a whey drink (277). In that

study older men received a 30 gm or 70 gm whey drink then ate at libitum at breakfast (30 minutes later), lunch (265 min later) and dinner (510 min later). There was no reduction in food intake compared to the control day at any of the three meals on the 30 gm day. On the 70gm day there was no effect on breakfast intake, but a 15% reduction in lunch energy intake ($P < 0.05$ vs control), followed by a compensatory 7% increase at dinner.

When intake from whey drinks plus subsequent food intake is calculated, absent (usually) or only minor suppression of subsequent food intake by whey drinks has resulted in consistent findings of increases in total energy intake and even greater proportional increases in protein intake after whey drinks in our short-term studies of older people. For example, in the study above where whey drinks were taken before breakfast and men studied for the rest of the day (277), there were non-significant 4% and 3% increases in total energy intake on the 30 gm and 70 gm days respectively compared to the control day. Total daily protein intake was increased significantly by the whey drinks in a dose-responsive manner, with increases almost equal to the protein content of the whey drinks (+31 gm on the 30 gm whey drink day, + 62 gm on the 70 gm whey drink day, $P < 0.001$ vs control (277).

These findings suggest that it should be possible to give enough extra protein to older people to preserve or increase muscle mass and function without suppressing energy intake and promoting weight loss, particularly if they are encouraged to continue their usual non-supplement food (energy) intake.

2.5.2 What is the best timing of the protein supplement use by older people?

Because older men can increase their protein intake in a single episode into the range of 30-40 gm - enough to maximize the protein's anabolic effects on muscle without the timing of that supplement's ingestion making much if any difference to subsequent appetite and food intake, these supplement can probably be ingested as a between-meal supplement, close to, or even with meals. Effects in women are likely to be similar (14), but require further study. The effects of more frequent protein doses across the day also need to be determined.

Indeed, if the supplement is given with a protein-containing meal, instead of between meals, less supplementary protein is needed to reach the 30-40 gm anabolic threshold described above. Older men in the NuAge study for example had a mean daily protein intake of 1 gm/kg (mean of 18 gm at breakfast and 23 gm at lunch and dinner) (256). Their protein intake could be increased to 1.33 gm/kg/day with a total protein intake of 35 gm per meal two meals a day by adding as little as 12 gm protein with each of lunch and dinner. A pragmatic measure, to allow for those with below average dietary protein intakes, would be take 20 gm of a protein supplement with the two meals each day already containing the most protein. This dose is unlikely to have many if any side effects for most older people.

2.5.3 Effect of ageing on gastric function and emptying

Healthy ageing is associated with changes in gastric function. These include reduced perceptions of proximal gastric distension and delayed gastric accommodation (91), probable changes in the intra-gastric distribution of food after its ingestion, greater

stimulation of phasic pyloric pressure waves by intra-duodenal lipid (46), and slowing of gastric emptying. It is unclear, however, how much, if at all, these changes contribute to the age-related reduction in the satiating effect of ingested protein and responses to protein supplements in older people. It is possible, however, that they may contribute to age-related reduced appetite and food intake.

2.5.3.1 Intra-gastric food distribution and antral area

After food is ingested, it is distributed throughout the stomach, with varying amounts in the proximal versus distal stomach (antrum). Antral distension appears to be a greater determinant of satiety and satiation than proximal gastric distension; the more distal the intra-gastric distribution of food, the greater the antral distension, the greater the sensations of fullness, the less the hunger, and the lower the subsequent food intake (25). Owing to impaired receptive relaxation of the gastric fundus (91) and other factors, food is distributed more distally in older people, as indicated by them having larger antral areas than young adults after ingestion of the same mixed macro-nutrient loads (25). As a result, they probably experience greater fullness and lower hunger ratings. While the more distal movement of food after its ingestion thus probably contributes to reduced hunger and increased fullness in older people, it is unclear how these changes could contribute to the reduced suppression of appetite ratings and food intake after oral ingestion of protein, alone or combined with other nutrients.

2.5.3.2 Gastric emptying

The oral ingestion of nutrients, irrespective of their type, slows gastric emptying.

Gastric emptying of non-nutrient drinks is slightly slower in older than in young adults (12, 14-16, 25, 27, 91, 98, 214, 277, 278). Whey protein drinks in doses of 30–70 gm slow gastric emptying in a dose-responsive manner (14, 15, 17), and probably slow it more in older than in young adults; the stomachs of healthy older men emptied whey into the duodenum at approximately 0.8 kcal/min compared with 1.0 kcal for young adult men ($P < 0.05$) in one study (15). Gastric emptying of whey protein drinks is faster in young, non-obese men than women (17), but this sex difference is no longer present in healthy people over 65 years (14). Gastric emptying of whey drinks occurs at a similar rate in obese young and older men (278), with neither age group having suppression of appetite or food intake 3 hours after ingestion of 30 gm whey—a suppressive dose in young, non-obese men (9, 15, 17). The findings suggest that obesity may blunt the effects of whey on both food intake and the slowing of gastric emptying.

Delayed gastric emptying results in more food in the stomach at any given time after food ingestion than would otherwise be the case. This increases gastric distension and, depending on the distribution of the food within the stomach, might be expected to produce greater feelings of fullness and reduced subsequent food intake. Consistent with this, the intentional creation of gastric distension and fullness by implanting gastric balloons has had some success in the treatment of obesity (279). Greater gastric distension after eating, due to slower gastric emptying, in older than in young adults, might thus be expected to cause greater post-eating suppression of appetite and food intake in older than in young adults. Existing evidence does not suggest, however, that this is so. Perceptions of gastric distension are less in older than young adults (91), and healthy older people have fullness ratings (unlike hunger ratings) 3 h after a whey drink, at the start of an ad libitum meal, that do not relate

significantly to energy intake at that meal (15). Furthermore, as outlined above, older people have less, not more, suppression of appetite and food intake by whey drinks than younger adults, despite slower gastric emptying and presumably greater gastric distension at any given time after the drinks. Our groups' studies have largely involved administering the ad libitum test meal 3 hours after the whey test drink, in order to allow the full effect of gastric mechanisms to be studied. The stomach is empty or almost empty by then after both 30 gm and 70 gm whey protein drinks (15). While it is possible that, if the meal had been given earlier, greater suppressive effects on intake due to greater gastric distension would have been present, this is not supported by our finding of a lack of suppression by 30 gm whey of food intake immediately after the drink and hourly up to 3 hours after the drink (98).

2.5.3.3 Small intestinal satiety mechanisms

As ageing has similar qualitative effects on appetite and food intake after intra-duodenal protein (whey) to those of oral whey (see above), and the age-related changes in gastric mechanisms do not appear to explain the reduced suppression of appetite and food intake by protein in older people, it seems likely that the reduced age-related changes in response to protein are mediated mainly by reductions in the satiating effects of the protein after it enters the small intestine. Nevertheless, slower gastric emptying in older people may modify these post-gastric effects by delaying their onset and prolonging their duration. Sixty minute duodenal infusions of 0.5 kcal/min and 1.5 kcal whey/minute do not suppress subsequent food intake in healthy older men, but 3 kcal/min infusions do, albeit less in older than in younger men (9). As healthy older men empty whey drinks from the stomach at a rate of approximately 0.8 kcal/min (15), it is likely that, even if a very high protein load is taken in (drink

form, for most older people, their age-related slowing of gastric emptying and markedly reduced effect of whey once it enters the duodenum combine to result in no suppression of appetite or subsequent energy intake. In contrast, duodenal infusions of whey in doses of 0.5, 1.5, and 3 kcal/in have dose-responsive suppressive effects on energy intake in young men, who empty whey drinks from the stomach faster than older men (~1 kcal/min) (15). Together, these findings explain the suppressive effects of both 30 gm and 70 gm whey drinks on subsequent energy intake in young, but not older adults (15).

2.5.4 Selected hormones

Ingestion of protein, either orally or by infusion directly into the duodenum, results in changes in circulating concentrations of a number of hormones with definite or possible effects on appetite and subsequent food intake. Among them, concentrations of cholecystokinin (CCK), insulin, glucagon, gastric inhibitory peptide (GIP), glucagon-like peptide-1 (GLP-1), peptide tyrosine-tyrosine (PYY), and amino acids increase, while glucose does not change and ghrelin concentrations decrease (12-14, 16, 17).

A number of these small intestinal responses to protein ingestion differ between healthy older and younger adults and these age-related changes in turn possibly affect the responses to protein or protein containing supplements in older people.

2.5.4.1 Cholecystokinin (CCK)

CCK is released by the small bowel after nutrient ingestion and acts to slow gastric

emptying and reduce food intake. Circulating CCK concentrations increase less after oral protein ingestion in young women than in young men, which may be one reason for the lower satiating effect of whey protein in young women than men (17). Circulating CCK concentrations are higher in healthy fasting older than young adults (29, 90), and older adults retain their sensitivity to the satiating effects of exogenous CCK (231). Increased CCK activity may thus be a cause of the anorexia of ageing and reduced hunger pre meals observed in older adults. Increases in circulating CCK concentrations are at least as great (53), if not greater (58), after whey drinks in healthy older than young adult men and women. The cause of this possibly greater rise is not known; it may relate to slower transition of whey through the small bowel in older people and, therefore, more prolonged contact with the CCK releasing cells. Ageing-related increases in CCK secretion and action are unlikely, however, to contribute to the reduced suppression of appetite and food intake by whey protein observed in older people compared with young adults. The opposite might be expected. The exact role of CCK in mediating age-related responses to protein supplements remains to be determined.

2.5.4.2 Glucagon-like peptide 1 (GLP-1)

GLP-1 is released by the small bowel and colon in response to food ingestion. Like CCK, it slows gastric emptying and has satiating effects. Fasting GLP-1 concentrations are higher in healthy older than young ageing (29, 90), which may thus contribute to lower basal (fasting) hunger in older people. Circulating GLP-1 concentrations appear to increase to a similar extent after whey drinks in young and older adults (13, 214, 279), which does not support a role for the lesser suppression of food intake by whey protein in older adults.

2.5.4.3 Gastric inhibitory peptide (GIP)

Along with GLP-1, GIP is an incretin that plays roles in the control of glucagon, insulin, and blood glucose concentrations. It is not clear what role GIP plays in the control of appetite and feeding, but it may have some effect to stimulate food intake (280). Circulating fasting GIP concentrations are not affected by normal ageing, but GIP concentrations increase more after oral whey ingestion in older than in young adults (12, 14, 16, 25, 27, 91, 98, 214, 277, 278). This greater increase might act to reduce the whey-induced suppression of food intake that would otherwise occur. It might also act to limit blood glucose concentration increases in older people after protein is co-ingested with other nutrients.

2.5.4.4 Insulin and glucose

Insulin plays a key role in glucose homeostasis, while its role in appetite and feeding control is less clear. Oral ingestion of protein, including whey, stimulates insulin secretion in a dose-responsive manner (90). While oral ingestion of protein on its own has little, if any, effect on blood glucose concentrations, its co-ingestion with glucose by non-elderly adults with type 2 diabetes results in significantly smaller increases in blood glucose concentrations than ingestion of the same amount of glucose on its own (68). The stimulation of insulin secretion by whey drinks is not apparently affected by ageing, and remains robust (90). We have recently found that co-ingestion of 30 gm whey protein with 30 gm glucose in drink form significantly reduces the increase in blood glucose concentrations compared with ingestion of 30 gm glucose alone (peak glucose 7.4 vs. 9.0 mmol/L, $P < 0.01$) in men over 65 years

(265), and are now extending these studies to older people with type 2 diabetes. These findings suggest that moderately high whey protein intake together with carbohydrate might improve postprandial glycaemia in older people, particularly those with diabetes, and provide an additional benefit of taking a protein supplement with, rather than between, meals.

2.5.4.5 Glucagon

Circulating concentrations of glucagon are not affected by ageing (13, 214, 279). Whey protein drinks act to increase glucagon concentrations, to a similar degree in older and young adults (90).

2.5.4.6 Ghrelin

Ghrelin is an orexigenic hormone secreted by the enteroendocrine cells of the gastrointestinal tract, particularly the stomach. Circulating concentrations are highest in the fasting state and decrease after food ingestion. Fasting circulating concentrations may be slightly lower in older than in young adults, and thus contribute to the anorexia of ageing (26, 281, 282). Ghrelin concentrations are suppressed to a similar degree after whey drinks in healthy older and young adults (90), so it is not clear what role, if any, ghrelin plays in mediating age-related reductions in feeding responses to protein supplements.

2.6 Future directions

Future studies should focus on women as well as men to determine whether our

findings in older men also apply to women. They should also determine whether the results of our short-term studies with protein supplements, such as post-supplement blood pressure drops and failure to suppress appetite and food intake, are replicated in studies of longer term protein supplement use. Further studies, both short- and longer-term, of the effect of protein when co-ingested with carbohydrates on glucose metabolism in older people with and without diabetes mellitus would also be of interest.

2.7 Conclusions

Nutritional supplements, including pure-protein or protein-enriched drinks, are increasingly used by older people to maintain body weight and nutrition, and specifically to prevent or treat loss of muscle function, sarcopenia, and frailty. Daily protein requirements in older people appear to be higher than those of young adults—in the range of 1.2 gm/kg/day or more. Many older adults do not consume this much protein in their usual diet and are likely to benefit from an increase.

Protein supplements appear relatively safe, although care should be taken in those with or at risk of renal impairment, or prone to post-nutrient hypotension. Protein supplements (alone or in a mixed macronutrient supplement) in doses sufficient to reach the above daily intake are likely to be beneficial, perhaps best taken twice daily, if possible soon after resistance exercise, in doses that achieve protein intakes of 30 gm or more per episode. Our study results suggest that it is probably not important to give these supplements between meals, because they have little, if any, suppressive effect on appetite and later food intake in older people, owing to age-related changes in gastrointestinal and other mechanisms that are as yet poorly understood. Adding

protein supplements to usual food intake is very unlikely to reduce energy intake and instead is likely to increase overall energy and protein intake, particularly if encouragement is given to continue usual food intake. There may even be benefits in giving the supplement with meals, to achieve protein intakes above the effective anabolic threshold with lower supplement doses, and have favorable effects on food-induced blood glucose increases in older people with, or at risk of, developing type 2 diabetes mellitus.

Further studies are indicated to determine the following:

1. The acute effects of whey and other proteins, when co-ingested with other macronutrients, with and between meals, on appetite and food intake in older people.
2. The longer term effects of protein ingestion, alone and combined with other macronutrients, on appetite, food intake, and glucose homeostasis, in older people; i.e., whether effects observed in acute studies persist with longer-term administration.

A better understanding of the mechanisms underlying the reduced suppression of appetite and food intake by protein and other macronutrients in older compared with younger adults, whether gastro-intestinal, central, or both, can be used to develop ways of improving nutrition in at-risk older individuals. Although our studies with whey have identified a number of age-related changes in gastro-intestinal responses to protein ingestion, it is not yet clear exactly how they act to mediate age-related changes in appetite and feeding.

2.8 Overall aims and hypotheses

In order to address the questions raised in sections 1.2.4, 1.3.2 and 1.4.3, the overall aims and hypotheses of the thesis are that:

- Suppression of energy intake by whey protein when compared to control will be less in healthy older than younger adults, resulting in an increase in cumulative energy and protein intake in older men (chapter 3).
- The acute suppression of energy intake by whey protein, will be less in older men with obesity as compared to younger men with obesity (chapter 4).
- Insulin concentrations following both glucose and protein drinks would increase and, to determine whether the effect on insulin of the combination drink is synergistic or additive, we compared the effect of combined whey protein and glucose on the rise in plasma insulin concentrations with that of the sum of the effects of glucose and whey protein alone on insulin concentrations using a paired t-test (chapter 5).
- Insulin concentrations following both glucose and protein drinks would increase, and to determine if the effect on insulin of the combination drink is synergistic or additive, we compared the effect of combined whey protein and glucose (GP drink) on the rise in plasma insulin concentrations to that of the sum of the effects of glucose and whey protein alone (G drink + P drink) on insulin concentrations, using a paired t test (chapter 6).
- Older, when compared to younger, men will exhibit a greater fall in blood pressure, and a smaller increase in heart rate, following whey protein ingestion compared to control (chapter 7).

- The decrease in blood pressure in response to ingestion of 30g whey protein will be less compared to 70g whey protein in healthy older men (chapter 8).
- The hypotensive effects of whey protein containing drinks will be dependent on the energy, rather than protein, content of the drink and persist for greater than 2 h (chapter 9).

**CHAPTER 3: WHEY PROTEIN DRINK INGESTION
BEFORE BREAKFAST SUPPRESSED ENERGY
INTAKE AT BREAKFAST AND LUNCH, BUT NOT
DURING DINNER, AND WAS LESS SUPPRESSED IN
HEALTHY OLDER THAN YOUNGER MEN**

**Oberoi A, Giezenaar C, Clames A, Bøhler K, Lange K, Horowitz M,
Jones KL, Chapman I, Soenen S**

Published in *Nutrients*

2020

STATEMENT OF AUTHORSHIP

Title of the paper	Whey protein drink ingestion before breakfast suppressed energy intake at breakfast and lunch, but not during dinner, and was less suppressed in healthy older than younger men
Publication status	Published
Publication details	Oberoi AO, Giezenaar C, Clames A, Bøhler K, Lange K, Horowitz M, Jones KL, Chapman I, Soenen S. <i>Whey protein drink ingestion before breakfast suppressed energy intake at breakfast and lunch, but not during dinner, and was less suppressed in healthy older than younger men</i> . <i>Nutrients</i> 2020; 12: 3318. doi:10.3390/nu12113318

Candidate	Avneet Oberoi		
Contribution	Contributed to the overall design of the manuscript, literature review, data analysis and interpretation, drafting and revision of the manuscript.		
Overall percentage	75%		
Certification	This paper reports on original research I conducted during the period of my Higher Degree by Research candidature and is not subject to any obligations or contractual agreements with a third party that would constrain its inclusion in this thesis. I am the primary author of this paper.		
Signature		Date	October 2022

Principal Author***Co-Author Contributions***

By signing the Statement of Authorship, each author certifies that:

- i) the candidate's stated contribution to the publication is accurate (as detailed above);
- ii) permission is granted for the candidate to include the publication in the thesis; and

- iii) the sum of all co-author contributions is equal to 100% less the candidate's stated contribution.

Name of Co-Author	Caroline Giezenaar		
Contribution	Data collection and interpretation, and drafting of the manuscript.		
Signature		Date	October 2022
Name of Co-Author	Alina Clames		
Contribution	Data collection and interpretation, and drafting of the manuscript.		
Signature		Date	October 2022
Name of Co-Author	Kristine Bøhler		
Contribution	Data collection and interpretation, and drafting of the manuscript.		
Signature		Date	October 2022
Name of Co-Author	Kylie Lange		
Contribution	Conception and design of the manuscript, statistical analysis, data interpretation, drafting and revision of the manuscript.		
Signature		Date	October 2022
Name of Co-Author	Michael Horowitz		
Contribution	Conception and design of the manuscript, drafting and revision of the manuscript.		
Signature		Date	October 2022
Name of Co-Author	Karen L Jones		

Contribution	Conception and design of the manuscript, data interpretation, drafting and revision of the manuscript.		
Signature		Date	October 2022
Name of Co-Author	Ian Chapman		
Contribution	Conception and design of the manuscript, statistical analysis, data interpretation, drafting and revision of the manuscript.		
Signature		Date	October 2022
Name of Co-Author	Stijn Soenen		
Contribution	Conception and design of the study, data interpretation, statistical analysis, drafting of the manuscript and overall responsibility for the study		
Signature		Date	October 2022

3.1 Abstract

Ageing is associated with changes in feeding behavior. We have reported that there is suppression of energy intake three hours after whey protein drink ingestion in young, but not older, men. This study aimed to determine these effects over a time period of 9 h. Fifteen younger (27 ± 1 years, 25.8 ± 0.7 kg/m²) and 15 older (75 ± 2 years, 26.6 ± 0.8 kg/m²) healthy men were studied on three occasions on which they received, in a randomized order, a 30 g/120 kcal, 70 g/280 kcal whey-protein, or control (~2 kcal) drink. Ad-libitum energy intake (sum of breakfast, lunch, and dinner) was suppressed in a protein load responsive fashion ($P = 0.001$). Suppression was minimal at breakfast, substantial at lunch (~16%, $P = 0.001$), no longer present by dinner, and was less in older than younger men ($-3 \pm 4\%$ vs. $8 \pm 4\%$, $P = 0.027$). Cumulative protein intake was increased in the younger and older men (+20% and +42%, $P < 0.001$). Visual analogue scale ratings of fullness were higher and desire to eat and prospective food consumption were lower after protein vs. control, and these effects were smaller in older vs. younger men (interaction effect $P < 0.05$). These findings support the use of whey-protein drink supplements in older people who aim to increase their protein intake without decreasing their overall energy intake.

3.2 Introduction

The number of older people with malnutrition, both under- and over-nutrition, is rising (283). Healthy ageing is associated with a reduction in appetite and food intake, including protein intake, which predisposes older people to loss of body weight and in particular, skeletal muscle mass (10, 71). The latter is associated with a decrease in function and quality of life (284). The causes of the reduction in food intake during healthy ageing are likely to be heterogeneous, including changes in gastrointestinal mechanisms induced by nutrient intake, such as slowing of gastric emptying (70, 285).

A common strategy to increase energy intake and body weight in undernourished older people is the use of > 25–30 g whey protein-enriched supplements (286), which may result in preserved or even increased muscle mass and strength (286, 287). We reported that in healthy older adults, when compared to younger adults, the acute suppression (up to 3 h following ingestion) of energy intake by protein administered orally or infused directly into the duodenum is less, resulting in an increase of overall energy and protein intake in the older adults (9, 15, 98). In healthy, younger adults, protein is considered to be the most satiating macronutrient and protein-rich supplements and diets are often recommended as a weight loss strategy in obese, younger individuals. There is a lack of definitive evidence on their efficacy (288, 289), especially in older adults.

In this study, we aimed to characterize the effect of ageing on the suppression of food intake at breakfast, lunch, and dinner over a time period of 9 h by a pre-breakfast whey protein load (30 g and 70 g) compared to a control drink in healthy younger

and older men. We hypothesized that suppression of energy intake by whey protein when compared to control would be less in healthy older than younger adults, resulting in an increase in cumulative energy and protein intake in the older men.

3.3 Materials and methods

3.3.1 Participants

The study included 15 healthy younger men (mean \pm standard error of the mean (SEM) age: 27 ± 1 years; body weight: 76.1 ± 2.0 kg; height: 1.73 ± 0.02 m; body mass index (BMI): 25.8 ± 0.7 kg/m²) and 15 healthy older men (75 ± 2 years; 80.7 ± 2.9 kg; 1.75 ± 0.01 m; 26.6 ± 0.8 kg/m²). Body weight and BMI of the younger and older men did not differ significantly ($P > 0.05$). Participants were recruited by online advertisement and by flyers placed on notice boards at the University of Adelaide, Adelaide, Australia.

Exclusion criteria included smoking; alcohol intake of > 2 standard drinks on > 5 days per week; being vegetarian; intake of any illicit substance; use of prescribed or non-prescribed medications that may affect appetite, body weight, gastrointestinal function, or energy metabolism; food allergy(s); diabetes mellitus (fasting glucose concentration > 6.9 mmol/L); epilepsy; gallbladder, pancreatic, cardiovascular, or respiratory diseases; significant gastrointestinal symptoms, disease, or surgery; any other illness deemed significant by the investigator; and an inability to comprehend the study protocol. Inclusion criteria included being weight stable ($< 5\%$ fluctuation in their body weight) at study entry, as assessed by their self-reported weight in the preceding 3 months, and maintenance of usual physical activity level.

All participants gave written informed consent for inclusion before they participated in the study. The study was conducted in accordance with the Declaration of Helsinki and the protocol was approved by the Ethics Committee of The Royal Adelaide Hospital (HREC/18/CALHN/132) and registered under trial registration number ACTRN12618000881235.

3.3.2 Protocol

Each participant was studied on three occasions, separated by ~3–10 days. On each occasion, they received, in a randomized order (using the method of randomly permuted blocks; www.randomization.com), a single drink of either flavoured water (control; ~2 kcal), 30 g whey protein (120 kcal), or 70 g whey protein (280 kcal). The drinks were equivolaemic (~450 mL) and contained different quantities of food-grade unflavoured whey protein isolate (Bulk Nutrients, Tasmania, Australia) dissolved in varying amounts of distilled water, sodium chloride, and low-calorie lime cordial (Bickford's "diet lime" cordial) (15).

Volunteers arrived at the laboratory at ~8.00 a.m. after fasting for ~12 h overnight and refraining from strenuous exercise and alcohol for 24 h. The participants were provided with a standard meal the night before each study day (beef lasagne, McCain Foods Pty Ltd., Wendouree, VIC, Australia ~591 kcal). Participants were told that we were assessing perceptions of appetite around the 3 meals, but not that we measured their food/energy intake.

At baseline ($t = -5$ min), perceptions of appetite were assessed by visual analogue scales (VAS) and the antral area of the stomach (cm^2) was measured with a LogiqTM

e-ultrasound machine (GE Healthcare Technologies, Sydney, NSW, Australia). Subsequently, the drink was administered at $t = -2$ min (~8.30 a.m.) and was served in an opaque cup to ensure that the volunteers were blinded. Participants were asked to ingest the drink within 2 min. Following consumption of the drink ($t = 0$ min), palatability of the drink and perceptions of appetite were assessed by VAS. The antral area of the stomach was measured at several time points between the drink and breakfast ($t = 0, 5, 20, 35$ min) and not thereafter. Energy intake was measured at breakfast ($t = 35\text{--}65$ min; ~9 a.m.), lunch ($t = 275\text{--}305$ min; ~1 p.m.), and dinner ($t = 515\text{--}545$ min; ~5 p.m.). Breakfast and lunch consisted of a cold buffet-style meal (Table 3.1) and dinner consisted of a warm meal and a small variation of buffet items (Table 3.2). Participants were instructed to consume food until they were comfortably full. Before and after consumption of the meals, perceptions of appetite, in terms of hunger, fullness, desire to eat, and prospective food consumption, were assessed ($t = 0, 5, 20, 35, 65, 80, 95, 275, 305, 320, 335, 515, 545, 560, 575$ min). Participants were not permitted to consume any food or drink between ingesting the study drink and the end of the study day, except at the breakfast, lunch, and dinner meals provided during the study day. Water intake in between meals was allowed, but not within 30 min before their next meal.

Table 3.1: Composition of the cold buffet-style breakfast and lunch meal

Food items	Amount served (g)	Energy Content (kcal)	Protein (g)	Carbohydrate (g)	Fat (g)
Wholemeal bread, 4 slices *	125	308	13.8	54.8	4.9
White bread, 4 slices *	125	304	11.1	61.4	2.7
Cheese, sliced †	85	346	22.6	0.9	29.2
Ham, sliced ‡	100	95	17.1	3.5	1.8
Chicken, sliced §	100	104	19.4	3.7	1.7
Margarine	20	108	0.0	0.0	12.4
Mayonnaise ¶	20	137	0.4	0.7	15.2
Tomato, sliced	100	13	1.0	2.0	0.1
Cucumber, sliced	100	11	0.5	2.0	0.1
Lettuce	100	5	0.9	0.4	0.0
Apple	170	89	0.5	2.0	0.1
Banana	190	166	3.3	39.0	0.2
Fruit salad **	140	81	0.4	17.7	1.3
Strawberry yogurt ††	175	162	9.1	25.0	3.4
Chocolate custard ‡‡	100	105	3.3	16.9	3.1
Milky Way §§	12	52	0.3	9.0	1.9
Orange juice, unsweetened	300	117	1.9	22.6	2.7
Iced coffee ¶¶	375	254	12.4	38.3	6.6
Water	600	0	0.0	0.0	0.0
<i>Total</i>		<i>2,457</i>	<i>19%</i>	<i>49%</i>	<i>32%</i>

* Sunblest, Tiptop, George Weston Foods Ltd, Enfield, NSW, Australia. † Coon Tasty Cheese slices, Australian Cooperative Foods Ltd, Sydney Olympic Park, NSW, Australia. ‡ KR

Castlemaine boneless leg ham, George Weston Foods Ltd, Enfield, NSW, Australia. § Inghams chicken breast, Inghams Enterprises Pty Ltd, Burton, SA, Australia. || Vita-Lite canola, Peerless Holdings Pty Ltd, Braybrook, VIC, Australia. ¶ MasterFoods, Mars Food Australia, Berkeley Vale, NSW, Australia. ** Goulburn Valley, SPC, Ardmona Operations Ltd, Shepparton, VIC, Australia. †† Yoplait, LD&D Foods Pty Ltd, Docklands, VIC, Australia. ‡‡ Yogo, LD&D Foods Pty Ltd, Docklands, VIC, Australia. §§ Mars Chocolate Australia, Wendouree, VIC, Australia. ||| Golden Circle Orange juice, Golden Circle Limited, QLD, Australia ¶¶ Farmers Union, LD&D Foods Pty Ltd, Docklands, VIC, Australia.

Table 3.2 Composition of the dinner meal

Food items	Amount served (g)	Energy Content (kcal)	Protein (g)	Carbohydrate (g)	Fat (g)
Pasta with Meatballs α	500	720	27.7	78.4	35.0
Whole meal bread, 4 slices *	125	308	14.0	55.5	4.9
Margarine \parallel	20	108	0.0	0.0	12.5
Philadelphia cream cheese $^{\circ}$	68	175	3.8	2.1	17.3
Apple	170	89	0.5	2.0	0.1
Banana	190	166	3.3	39.5	0.2
Fruit salad **	140	81	0.4	17.9	1.4
Strawberry yogurt $\dagger\dagger$	175	162	9.2	25.3	3.4
Chocolate custard $\ddagger\ddagger$	100	105	3.3	17.1	3.1
Muesli bar \ominus	35	185	5.6	12.5	13.1
Orange juice, unsweetened \lll	300	117	1.9	22.9	2.7
Water	600	0	0.0	0.0	0.0
<i>Total</i>		<i>2,216</i>	<i>13%</i>	<i>49%</i>	<i>38%</i>

α Man Size Spaghetti and Meatballs, McCain Foods Pty Ltd, Wendouree, VIC, Australia. * Sunblest, Tiptop, George Weston Foods Ltd, Enfield, NSW, Australia. \parallel Vita-Lite canola, Peerless Holdings Pty Ltd, Braybook, VIC, Australia. $^{\circ}$ Philadelphia Spreadable Cream Cheese snack tubs, Consumer Advisory Service, Melbourne, VIC, Australia. ** Goulburn Valley, SPC, Ardmona Operations Ltd, Shepparton, VIC, Australia. $\dagger\dagger$ Yoplait, LD&D Foods Pty Ltd, Docklands, VIC, Australia. $\ddagger\ddagger$ Yogo, LD&D Foods Pty Ltd, Docklands, VIC, Australia.

3.3.3 Measurements

The primary outcome of the study was ad libitum energy intake at the buffet-style meal and secondary outcomes include antral area and appetite.

3.3.3.1 Energy intake

To quantify the amount eaten, the weights of the food items were recorded before and after they were offered to the participants (15). Energy intake and macronutrient composition was calculated using commercially available software (Foodworks 3.01, Xyris Software, Highgate Hill, QLD, Australia). Absolute (kcal) and percentage suppression of energy intake (expressed as % of energy intake of the control day) by protein were calculated.

3.3.3.2 Antral area

Gastric emptying (gastric retention) was determined by measuring the antral area of the stomach. The circumference of the antral area was measured with a Logiq™ e-ultrasound machine (GE Healthcare Technologies, Sydney, NSW, Australia) by using a 3.5 C broad spectrum 2.5–4 MHz convex linear array transducer. Antral area (cm²) was determined with the use of a caliper and calculation program built into the ultrasound machine. Volunteers were seated on a chair and were asked to be still during the measurement. The transducer was positioned vertically to obtain a parasagittal image of the antrum, with the superior mesenteric vein and the abdominal aorta in a longitudinal section. If gastric contractions were observed, the acquisition was paused until the contraction wave had passed. To calculate meal

retention in the whole stomach, the fasting antral area (measured at baseline) was subtracted from subsequent measurements performed after ingestion of the drinks (290). Gastric retention was then calculated at a given time point as:

$$\text{Retention (\%)} = [\text{AA}(t) - \text{AA}(f)] / [\text{AA}(\text{max}) - \text{AA}(f)] \times 100,$$

where AA(t) = antral area measured at a given time point, AA(f) = fasting antral area, and AA(max) = maximum antral area recorded after drink ingestion (15).

3.3.3.3 Perceptions of appetite and palatability

Perceptions of appetite in terms of hunger, fullness, desire to eat, and prospective consumption were assessed by use of a VAS questionnaire (291). The questionnaire consisted of 100 mm horizontal lines, where 0 represented that the sensation was “not felt at all” and 100 represented that the sensation was “felt the greatest.” Volunteers placed a vertical mark on each horizontal line to signify the strength of each sensation at the specified time points. Baseline fasting ratings were calculated as the mean of the three study days. Total area under the curve (AUC) was calculated over 0–180 min (15). Palatability of the drink was assessed by ratings of pleasantness, intenseness, full of taste, sweetness, saltiness, sour, bitterness, umami, and creaminess immediately after drink intake; palatability of the meal was assessed by like of taste, like of aftertaste, and enjoyability of the meal by use of a VAS questionnaire.

3.4 Data and statistical analysis

Statistical analyses were performed using SPSS software (version 24; IBM, Armonk, NY, USA). Power calculations were performed for the primary outcome of energy intake using measures of variance obtained from previous data (standard deviation (SD) of 181 kcal) (15) to detect a minimum difference in suppression of energy intake by the treatment condition compared with the control of 251 kcal between younger and older participants. Age and protein load main effects and the age by protein load interaction on outcomes were determined by using two-way repeated-measures analysis of variance (ANOVA). Residuals from all models were checked for normality and constant variance and all assumptions were found to be met. When significant treatment and/or interaction effects were present, Bonferroni corrected post hoc tests were performed to determine which specific drink conditions were different between age groups. Statistical significance was accepted at $P < 0.05$. All data are presented as means \pm SEMs.

3.5 Results

The study protocol was well tolerated by all participants.

3.5.1 Energy intake

Energy intake after the drink (sum of breakfast, lunch, and dinner; Figure 3.1) was suppressed by whey protein compared to control (protein load main effect on energy intake $P = 0.012$), driven by the suppression of the 70 g whey protein drink (young: -251 ± 117 kcal, $-8 \pm 4\%$; older: -184 ± 96 kcal, $-5 \pm 4\%$; post-hoc test $P = 0.023$),

which was greater ($P = 0.027$) when compared with the 30 g protein drink (young: -88 ± 108 kcal, $-3 \pm 4\%$; older: -5 ± 99 kcal, $0 \pm 4\%$; Table 3.3). Suppression of energy intake by the 70 g whey protein compared to control (protein load main effect, $P = 0.007$) was greatest at lunch (young: -181 ± 83 kcal, $-17 \pm 8\%$; older: -154 ± 49 kcal, $-15 \pm 5\%$; $P = 0.001$; Figure 3.2). Protein intake of the drink, before breakfast, did not affect ad libitum energy intake at dinner in either age group. Suppression of energy intake (sum of breakfast, lunch, and dinner) by whey protein was less in healthy older men: -94 ± 82 kcal when compared to younger men -169 ± 100 kcal (there was a main effect of age on suppression of energy intake by protein compared to control $P = 0.027$).

Cumulative energy intake (sum of energy in test drink, breakfast, lunch, and dinner) was not significantly different between study days and age groups (young: control: 2929 ± 131 kcal, 30 g whey protein: 2961 ± 161 kcal and 70 g whey protein: 2958 ± 163 kcal; older: 2878 ± 165 kcal, 2993 ± 122 kcal and 2974 ± 148 kcal, all $P > 0.05$).

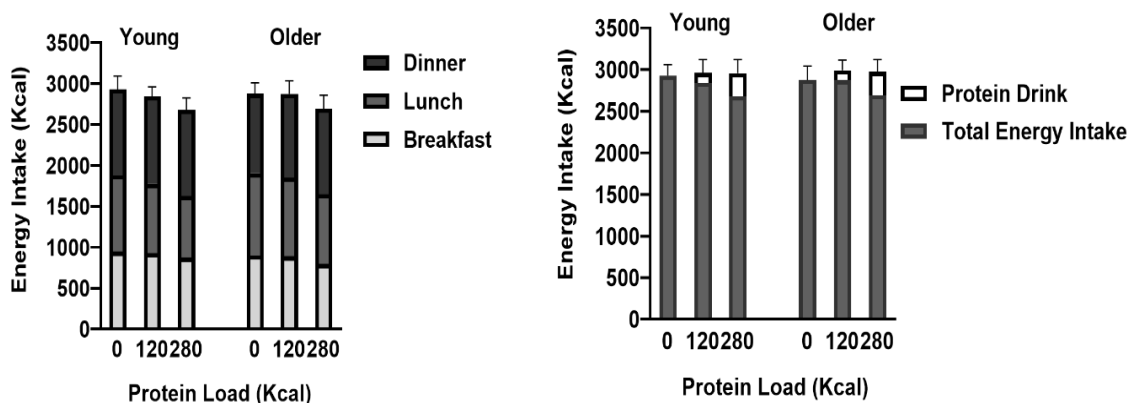


Figure 3.1: Energy intake at breakfast, lunch, and dinner following whey protein ingestion in healthy young and older men. Mean (\pm SEM) ad libitum energy intake (kcal; left) at breakfast (light grey bars), lunch (dark grey bars), and dinner (black bars) following drink ingestion containing flavored water (control, ~ 2 kcal) or whey protein

(30 g/120 kcal or 70 g/280 kcal) and cumulative energy intake (kcal; right; sum total energy intake at breakfast, lunch, and dinner combined (dark grey bars) and protein drink (white bars)) in young (left; $n = 15$) and older (right; $n = 15$) men. Age and protein load main effects and interaction effects were determined by repeated measures ANOVA. * The 70 g protein drink suppressed energy intake (sum of breakfast, lunch, and dinner) compared with the control (protein load effect $P = 0.012$, post-hoc $P = 0.023$).

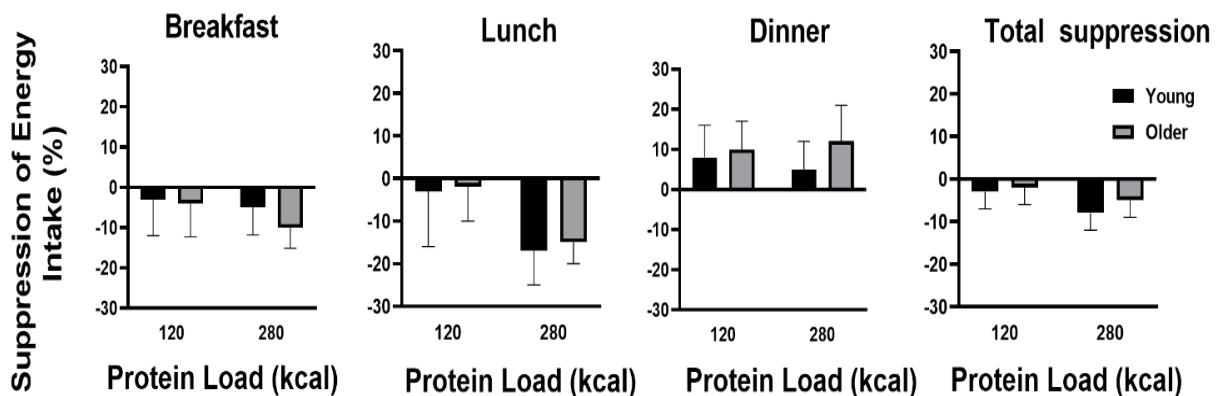


Figure 3.2: Suppression of energy intake by whey protein at breakfast, lunch and dinner and total suppression of energy intake in healthy young and older men. Mean (\pm SEM) suppression of energy intake (kcal) at breakfast lunch and dinner), and total intakes (sum breakfast, lunch and dinner) following drink ingestion containing flavoured water (control, ~ 2 kcal) or whey protein (30 g/120kcal or 70 g/280kcal) in young (grey shading; $n = 15$) and older (black shading; $n = 15$) men. Age and protein load effects and interaction effects were determined by using repeated-measures ANOVA. *Energy intake was suppressed by protein compared to control (protein-load effect $P = 0.012$). Suppression of energy intake by 70g protein compared to control ($P=0.007$) was particularly occurring during lunch ($P = 0.001$). Suppression of energy intake (sum of breakfast, lunch and dinner) by protein was less in healthy older when compared to younger men (age effect $P = 0.027$).

Table 3.3: Energy intake at, and macronutrient composition of, breakfast, lunch and dinner following whey protein ingestion in healthy younger and older men

	Young (n=15)				Older (n=15)			
	Breakfast	Lunch	Dinner	Total	Breakfast	Lunch	Dinner	Total
Control drink								
Energy intake (kcal)	947 ± 64	933 ± 74	1049 ± 68	2929 ± 131	896 ± 74	1007 ± 62	975 ± 79	2878 ± 165
Fat (energy %)	34 ± 1	34 ± 2	36 ± 2		29 ± 2	33 ± 6	39 ± 2	
Carbohydrate (energy %)	43 ± 2	43 ± 2	50 ± 2		51 ± 2	46 ± 2	47 ± 2	
Protein (energy %)	23 ± 1	23 ± 1	14 ± 1		20 ± 1	21 ± 1	14 ± 1	
30 g (120 kcal) protein drink								
Energy intake (kcal)	925 ± 67	848 ± 89	1068 ± 48	2841 ± 161	888 ± 60	962 ± 84	1023 ± 66	2873 ± 122
Fat (energy %)	34 ± 2	30 ± 3	38 ± 2		30 ± 1	34 ± 1	38 ± 2	
Carbohydrate (energy %)	43 ± 2	48 ± 4	47 ± 1		51 ± 3	46 ± 2	48 ± 1	

Whey protein drink ingestion) 23 ± 1 22 ± 2 15 ± 0 19 ± 1 20 ± 1 14 ± 1 Chapter 3

70g (280kcal) protein drink

Energy intake (kcal)	874 ± 70	752 ± 85*	1052 ± 56	2678 ± 163	794 ± 72	853 ± 69*	1047 ± 82	2694 ± 148
Fat (energy %)	34 ± 1	27 ± 2	48 ± 2		30 ± 2	32 ± 1	38 ± 2	
Carbohydrate (energy %)	43 ± 2	54 ± 3	47 ± 1		51 ± 4	46 ± 2	48 ± 1	
Protein (energy %)	23 ± 1	19 ± 2	15 ± 0		19 ± 1	22 ± 1	14 ± 0	

Mean (± SEM) ad libitum energy intake (kcal) at, and macronutrient composition (energy percentage) of, breakfast, lunch and dinner, following drink ingestion containing flavoured water (control, ~2 kcal) or whey protein (30 g/120 kcal or 70 g/280 kcal) in young (left; n = 15) and older (right; n = 15) men. Main age and protein-load effects and interaction effects were determined by using repeated-measures ANOVA. *Energy intake was suppressed by protein compared to control (protein-load effect $P = 0.012$). Suppression of energy intake by 70 g protein compared to control ($P = 0.007$) was particularly occurring during lunch ($P = 0.001$).

3.5.2 Protein intake

1. The sum of breakfast, lunch, and dinner protein intake after the test drinks decreased after the 70 g ($P = 0.023$), but not 30 g, whey protein drink when compared to the control day (protein load main effect $P = 0.009$, main effect of age $P = 0.71$, interaction effect $P = 0.54$).
2. Cumulative protein intake (sum of protein in the drink plus protein intake at the meals) was increased in a protein load responsive fashion (young: control: 143 ± 10 g, 30 g whey protein: +17%, 167 ± 9 g and 70 g whey protein: +36%, 195 ± 9 g; older: control: 133 ± 10 g, 30 g whey protein: +23%, 164 ± 10 g and 70 g whey protein: +47%, 195 ± 9 g; $P < 0.001$) comparably in the healthy younger and older men (main effect of age $P = 0.71$, interaction effect of age x protein load $P = 0.54$; Figure 3.3).

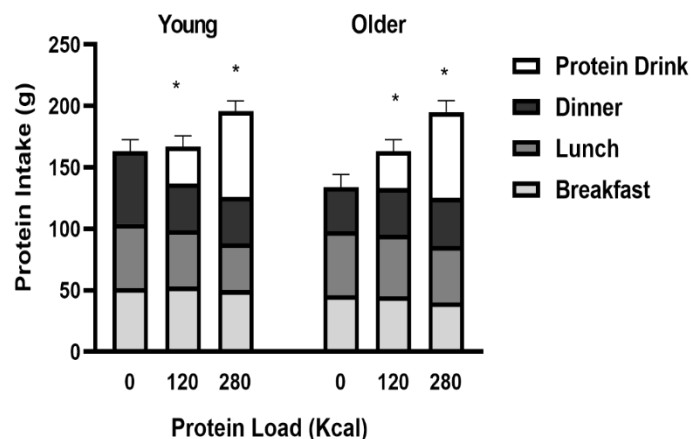


Figure 3.3: Mean (\pm SEM) protein intake (g) at breakfast (light grey bars), lunch (dark grey bars), and dinner (black bars) following drink ingestion containing flavored water (control, ~2 kcal) or whey protein (30 g/120 kcal or 70 g/280 kcal; white bars) in young (left; $n = 15$) and older (right; $n = 15$) men. Age and protein load main effects and interaction effects were determined by using repeated-measures ANOVA. * Cumulative protein intake (sum of protein drink plus protein

intake at meals) was increased in a protein load responsive fashion comparably in the healthy young and older men (main effect of age $P = 0.71$, protein load main effect $P < 0.001$, interaction effect $P = 0.54$).

3.5.3 Gastric emptying

Antral areas following overnight fasting (control: $3.4 \pm 0.8 \text{ cm}^2$; 30 g whey protein: $2.8 \pm 0.7 \text{ cm}^2$; 70 g whey protein: $2.9 \pm 0.8 \text{ cm}^2$; protein load main effect $P = 0.21$) and immediately after drink consumption (control: $15.6 \pm 0.8 \text{ cm}^2$; 30 g whey protein: $16.2 \pm 0.8 \text{ cm}^2$; 70 g whey protein: $16.4 \pm 0.8 \text{ cm}^2$; protein load main effect $P = 0.76$) were comparable between the study days for both the age groups. Gastric retention was greater after both protein drinks compared to control (main effect of age $P = 0.27$, protein load main effect $P < 0.001$, interaction effect $P = 0.091$; Figure 3.4).

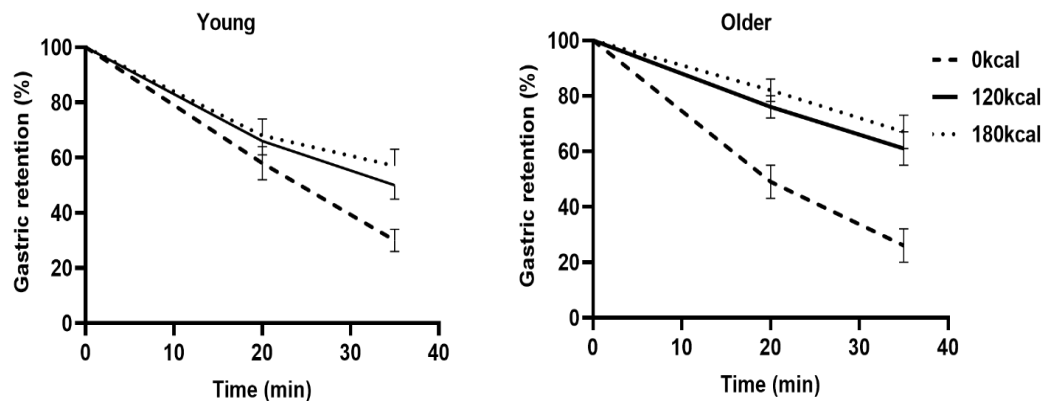


Figure 3.4: Mean (\pm SEM) Gastric Retention (%) of drinks containing flavored water (control, ~ 2 kcal) or whey protein (30 g/120 kcal or 70 g/280 kcal; open bars) in young (left; $n = 15$) and older (right; $n = 15$) men. Age and protein load main effects and interaction effects were determined by using repeated-measures ANOVA. *Gastric Retention, calculated based on the antral areas, were larger after both protein drinks compared to control (main effect of age $P = 0.27$, protein main effect

$P < 0.001$, interaction effect $P = 0.091$).

3.5.4 Appetite

Baseline perceptions of appetite in terms of hunger (young: 61 ± 8 mm; older: 59 ± 9 mm), fullness (13 ± 4 mm; 5 ± 2 mm), desire to eat (61 ± 7 mm; 52 ± 8 mm), and prospective food consumption (67 ± 5 mm; 55 ± 6 mm) were not significantly different between study days and age groups after overnight fasting (all $P > 0.05$). Protein drink ingestion affected fullness (protein main effect $P < 0.001$), desire to eat ($P < 0.001$), and prospective food consumption ($P = 0.002$; Figure 3.5) in a protein load related fashion; fullness was higher (AUC, both $P < 0.001$) and desire to eat (AUC, $P = 0.035$ and $P = 0.009$) and prospective food consumption (immediately before lunch, $P = 0.025$, $P = 0.006$) were lower after the 70 g whey protein drink compared to control and the 30 g protein drink. Older compared to younger men had a lesser desire to eat (main effect of age $P = 0.028$) but also less fullness (main effect of age $P = 0.003$, interaction effect of age x protein load $P < 0.001$) throughout the day (Figure 3.5).

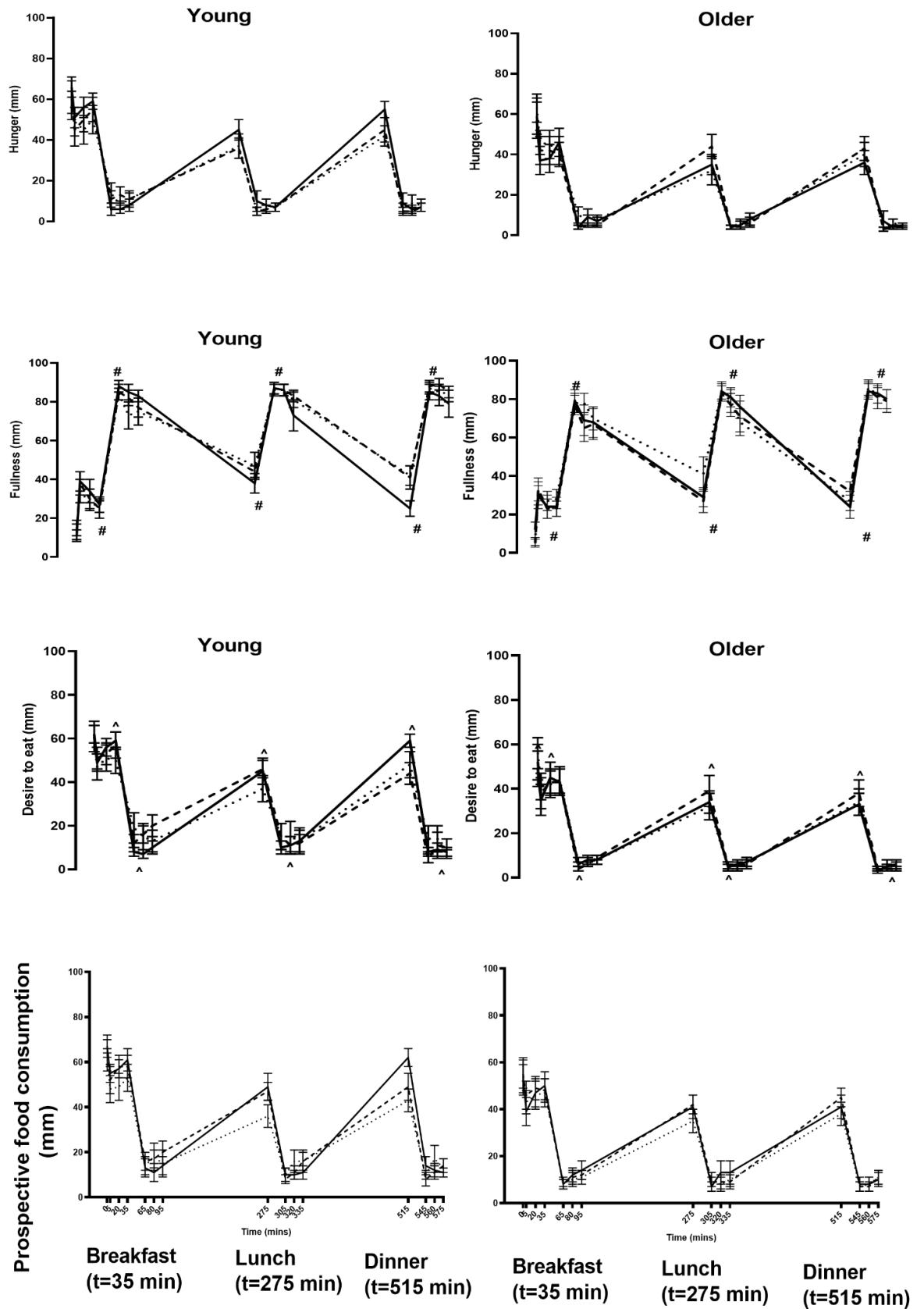


Figure 3.5: Mean (\pm SEM) visual analogue scores (VAS; 0–545 min) of hunger (A, B), fullness (C, D), desire to eat (E, F), and prospective food consumption (G,H)

following overnight fasting ($t = -5$) and after drink ingestion ($t = 0, 5, 20, 35, 65, 80, 95, 275, 305, 320, 335, 515, 545, 560, 575$ min) containing flavored water (control, ~2 kcal) or whey protein (30 g/120 kcal or 70 g/280 kcal; open bars) and immediately before and after breakfast (B), lunch (L), and dinner (D) in young (left; $n = 15$) and older (right; $n = 15$) men. Age and protein load main effects and interaction effects were determined using repeated-measures ANOVA. Protein affected #fullness (protein load main effect $P < 0.001$), ^desire to eat ($P < 0.001$), and *prospective food consumption ($P = 0.002$) in a protein load related fashion. Older compared to younger men had lower desire to eat (main effect of age $P = 0.028$) and fullness ($P = 0.003$, interaction effect $P < 0.001$).

3.5.5 Palatability of drinks and meals

The 70 g whey protein drink was perceived to be creamier when compared to the flavored control drink ($P = 0.016$). Ratings of pleasantness, intenseness, full of taste, sweetness, saltiness, sour, bitterness, umami, and creaminess of the drinks were not significantly different (main effect of protein $P > 0.05$). The healthy younger men rated the drinks as more bitter than the older men (young: 19 ± 4 mm; older: 26 ± 3 mm, main effect of age $P = 0.037$). All other palatability ratings of the drinks were comparable between the age groups: pleasant (young: 47 ± 5 mm; older: 44 ± 4 mm), intense (51 ± 4 mm; 55 ± 3 mm), fullness (59 ± 4 mm; 59 ± 3 mm), sweet (53 ± 3 mm; 48 ± 3 mm), salty (31 ± 6 mm; 37 ± 4 mm), sour (34 ± 6 mm; 39 ± 4 mm), umami (34 ± 5 mm; 35 ± 3 mm), refreshing (40 ± 6 mm; 41 ± 4 mm), creaminess (27 ± 5 mm; 31 ± 3 mm, main effect of age all $P > 0.05$). Palatability of the meals, assessed as ratings of taste, aftertaste, and enjoyability, were comparable between study days and age groups (control, 30 g, 70 g protein: young: taste: 73 ± 5 mm, 75

± 5 mm, 72 ± 6 mm, after taste: 73 ± 5 mm, 72 ± 5 mm, 73 ± 5 mm, enjoyable: 73 ± 5 mm, 75 ± 5 mm, 74 ± 5 mm; older: taste: 72 ± 4 mm, 74 ± 3 mm, 72 ± 4 mm, after taste: 71 ± 3 mm, 71 ± 3 mm, 72 ± 4 mm, enjoyable: 73 ± 4 mm, 76 ± 3 mm, 73 ± 4 mm; main effects of age, protein load main effects and interaction effects all $P > 0.05$).

3.6 Discussion

This study compared the acute effects of ingestion of whey protein drinks containing 30 g and 70 g to those of a flavored control drink consumed 35 min before breakfast on ad libitum energy intake at breakfast, lunch, and dinner, perceptions of appetite throughout the day, and gastric emptying (antral area) in healthy younger and older men. Energy intake (sum of breakfast, lunch, and dinner) was suppressed in a protein load-responsive fashion at breakfast and in particular, at lunch, but not at dinner. Suppression of combined energy intake at breakfast, lunch, and dinner by the protein drink was less in healthy older (-3%) when compared to younger (-7%) men. Cumulative protein intake (sum of protein drink plus protein intake at the meals) was increased in a protein load responsive fashion ($+20\%$ and $+42\%$) in the healthy younger and older men. Gastric emptying of the protein drinks in the 35 min before breakfast was slower than that of the control. Fullness was higher and desire to eat and prospective food consumption lower after protein intake when compared with the control in a protein load related fashion. Older compared to younger men had a lower desire to eat but also lower fullness throughout the day, suggesting that older people experience lower sensitivity of the appetite-suppressing effects of a protein drink and may have a decreased perception of gastric distension as seen in our previous study (12, 91).

Overall, suppression of energy intake by protein was less in healthy older than younger men in this study, confirming the results of our previous studies (14-16, 18, 25), e.g., in a study with a comparable design, suppression of energy intake by oral whey protein ingestion was \sim -15% in healthy young compared to \sim -1% in older men. In the present study, energy intake (sum of breakfast, lunch, and dinner) was suppressed most by the 70 g whey protein load compared to control (\sim 7%) and at lunch, 4 h 35 min after the drink (\sim -20% in young and \sim -15% in older men). In contrast, there was no suppression of energy intake by pre-breakfast protein at dinner time, 8 h 35 min after the drink, in either age group (\sim +7% compared to control dinner). We reported previously that in healthy older people, the timing of a 30 g whey protein drink (3 h, 2 h, 1 h, and immediately before the buffet-style meal) does not affect subsequent energy intake in older people. The effect of the whey protein ingestion on energy intake throughout the day may be associated with the slightly slower gastric emptying, reported by us and others in previous studies measuring gastric emptying for a period of 3 h in healthy older, when compared with younger, people (14-16, 18). Gastric emptying may be associated with postprandial satiety by affecting plasma gut hormone concentrations (90) in healthy younger adults (91, 165, 290, 292).

The cumulative energy intake (sum of drink, breakfast, lunch, and dinner) was comparable between study days while cumulative protein intake was elevated during the protein conditions in both age groups. Cumulative energy intake on the protein days compared to control was slightly higher in older (+4%) than younger men (-1%), as was reported in our previous studies determining ad libitum energy intake 3 h after oral whey protein ingestion (15) and following 1 h whey protein infusions directly into the small intestine (9). The insignificant effect of the whey drink on

cumulative daily energy intake in this study may indicate that the ingestion of a single daily dose of whey protein, in doses up to 70 g, is unlikely to be a successful weight loss strategy to achieve a negative energy balance, without taking the effects on energy expenditure and muscle anabolism into account. Even if whey protein was given more than once a day, we have no evidence that this would have resulted in a greater cumulative energy deficit, particularly in older adults. The energy content of the protein drink would have equalled or outweighed suppression of energy intake produced by the protein drink. Given our finding with one protein drink before breakfast, it is likely that suppression of cumulative energy intake with multiple drinks would have been even less (293). The participants in this study were not aware, however, that we were interested in or measuring their ad libitum meal energy intake throughout the day in response to the different drinks. Young adults using protein supplements to lose weight may have different responses to those in this study. Cumulative protein intake was significantly increased by the 30 g and 70 g whey protein loads, particularly in the older men (young: +17% and +36% and older: +23% and +47%), reaching meaningful amounts sufficient to result in postprandial muscle anabolism in older adults (123, 287)—the 70 g whey protein drink increased protein intake by 62 g, or ~0.8 g/kg body weight, in the older men.

A limitation of the study was that we only studied men. This was to enable comparisons with the results of our previous studies conducted in men which clearly showed the effect of protein load. As men generally show greater variations in appetite and food intake in response to energy manipulation than women (17, 72), the effects of the protein drinks may be different in women and it would be appropriate to perform further studies including women. The healthy older participants were well nourished, unrestrained eaters, had an active lifestyle, and

comparable energy intake on the control day to the younger men. It has been reported numerous times that healthy ageing is associated with reduced food intake (25, 30) and hunger (25, 26, 214) and a blunting of the regulation of food intake (72, 294) as suggested by the findings of this study, i.e., less suppression of energy intake by protein. The suppressive effect of whey protein in younger adults may be affected by having dietary restraints or actively trying to lose body weight (295, 296). Furthermore, the overall suppressive effect of protein supplements may be influenced by protein supplement intake before each meal of the day. The significant increase in cumulative protein intake and slight increase in cumulative energy intake in the older men suggests that whey protein can be given at breakfast, and possibly also at other meals, without decreasing overall daily energy intake, which would benefit malnourished, frail, older people—further studies are warranted. Another possible limitation was that the study was limited to 9 h after drink ingestion. As the effect of the pre breakfast drink on energy and protein intake had worn off by dinner, however, it seems unlikely that it would have had any effect after that.

3.7 Conclusions

Energy intake was suppressed by whey protein drinks in a protein load-responsive fashion at breakfast and particularly, at lunch, but not at dinner, and suppression of energy intake by protein was less in healthy older than younger men. Cumulative protein intake was increased in a protein load responsive fashion. These findings support the use of whey-protein drink supplements in healthy older patients who aim to increase their protein intake without decreasing their overall energy intake.

**CHAPTER 4: ACUTE EFFECTS OF WHEY PROTEIN ON
ENERGY INTAKE, APPETITE AND GASTRIC EMPTYING IN
YOUNGER AND OLDER, OBESE MEN**

**Oberoi A, Giezenaar C, Jensen C, Lange K, Hausken T, Jones K.L,
Horowitz M, Chapman I, Soenen S**

Published in *Nutrition and Diabetes*

2020

STATEMENT OF AUTHORSHIP

Title of the paper	Acute effects of whey protein on energy intake, appetite and gastric emptying in younger and older, obese men
Publication status	Published
Publication details	Oberoi AO, Giezenaar C, Jensen C, Lange K, Hausken T, Jones, K.L, Horowitz M, Chapman I, Soenen S. <i>Acute effects of whey protein on energy intake, appetite and gastric emptying in younger and older, obese men</i> . Nutrition & Diabetes 2020; 10: 37. doi.org/10.1038/s41387-020-00139-8.

Candidate	Avneet Oberoi		
Contribution	Contributed to the overall design of the manuscript, literature review, data analysis and interpretation, drafting and revision of the manuscript.		
Overall percentage	75%		
Certification	This paper reports on original research I conducted during the period of my Higher Degree by Research candidature and is not subject to any obligations or contractual agreements with a third party that would constrain its inclusion in this thesis. I am the primary author of this paper.		
Signature		Date	October 2022

Principal Author***Co-Author Contributions***

By signing the Statement of Authorship, each author certifies that:

- i) the candidate's stated contribution to the publication is accurate (as detailed above);
- ii) permission is granted for the candidate to include the publication in the thesis; and

- iii) the sum of all co-author contributions is equal to 100% less the candidate's stated contribution.

Name of Co-Author	Caroline Giezenaar		
Contribution	Conception and design, drafting and revision of the manuscript.		
Signature		Date	October 2022
Name of Co-Author	Caroline Jensen		
Contribution	Revision of the manuscript.		
Signature		Date	October 2022
Name of Co-Author	Kylie Lange		
Contribution	Conception and design of the manuscript, statistical analysis, data interpretation, drafting and revision of the manuscript.		
Signature		Date	October 2022
Name of Co-Author	Trygve Hausken		
Contribution	Data interpretation and revision of the manuscript.		
Signature		Date	October 2022
Name of Co-Author	Karen L Jones		
Contribution	Conception and design of the manuscript, data interpretation, drafting and revision of the manuscript.		
Signature		Date	October 2022
Name of Co-Author	Michael Horowitz		

Contribution	Conception and design of the manuscript, data interpretation, drafting and revision of the manuscript.		
Signature		Date	October 2022
Name of Co-Author	Ian Chapman		
Contribution	Conception and design of the study, data interpretation, statistical analysis, drafting and revision of the manuscript		
Signature		Date	October 2022
Name of Co-Author	Stijn Soenen		
Contribution	Conception and design of the study, data interpretation, statistical analysis, drafting of the manuscript and overall responsibility for the study.		
Signature		Date	October 2022

4.1 Abstract

Background: Obesity is becoming more prevalent in older people. A management strategy in obese, young adults is to increase dietary protein relative to other macronutrients. It is not clear if this is effective in obese, older individuals. Obesity may be associated with diminished sensitivity to nutrients. We have reported that a 30-g whey protein drink slows gastric emptying more, and suppresses energy intake less, in older, than younger, non-obese men. The aim of this study was to determine the effect of a 30 g whey protein drink on energy intake, gastric emptying (GE) and glycaemia in obese, older and younger men.

Methods: In randomized, double-blind order, 10 younger (age: 27 ± 2 years; BMI: 36 ± 2 kg/m²), and 10 older (72 ± 1 years; 33 ± 1 kg/m²), obese men were studied twice. After an overnight fast, participants ingested a test drink containing 30 g whey protein (120 kcal) or control (2 kcal). Postprandial gastric emptying (antral area, 2D Ultrasound) and blood glucose concentrations were measured for 180 min. At $t = 180$ min participants were given a buffet meal and ad libitum energy intake was assessed.

Results: Older participants ate non-significantly less (~20%) than the younger participants (effect of age, $P = 0.16$). Whey protein had no effect on subsequent energy intake (kcal) compared to control in either the younger (decrease $3 \pm 8\%$) or older (decrease $2 \pm 8\%$) obese men (age effect $P > 0.05$, protein effect $P = 0.46$, age \times protein interaction effect $P = 0.84$). Whey protein slowed gastric emptying, to a similar degree in both age groups (50% emptying time: control vs. protein young men: 255 ± 5 min vs. 40 ± 7 min; older men: 16 ± 5 min vs. 50 ± 8 min; protein effect $P = 0.001$, age effect $P = 0.93$, age \times protein interaction effect $P = 0.13$).

Conclusions: Our data suggest that obesity may blunt/abolish the age-related effect

of whey protein on suppression of energy intake.

4.2 Introduction

While the number of older people is increasing worldwide (297, 298), it is not as well appreciated that obesity rates are rising across the age range. Approximately one-third of people over 65 years are obese BMI > 30 kg/m²) in Australia (299) and other developed countries (US, UK etc.) (300, 301) and this proportion is increasing. Although the BMI range associated with lowest morbidity and mortality increases with age (302), a BMI higher than 30 kg/m² in older people is still associated with adverse health outcomes, including diabetes, hypertension and heart disease. Obesity is therefore a serious problem in older people (303-306) and intentional weight loss is often recommended for obese, older adults (307).

Ageing is also associated with changes in feeding behaviour, gut function, and body composition, which impact on life expectancy and quality (9, 10, 13, 14, 16, 90). Skeletal muscle mass decreases, while fat mass increases (308). Insufficient protein intake in obese older individuals exacerbates muscle loss (309). This muscle loss, which may lead to sarcopaenia, is associated with functional impairment, increased rates of falls, increased nursing home admissions, and other adverse outcomes (111, 306, 310-315).

Protein supplements, often rich in whey protein, are commonly part of weight-loss strategies, based on the rationale that protein is more satiating than the other macronutrients (276, 316-318), although that effect may be modest, particularly when the protein is consumed as part of typical, ad libitum eating situations. Whey protein is high in essential amino acids which are rapidly digested, resulting in postprandial amino acid availability, stimulating muscle protein accretion more

effectively than casein and casein hydrolysate in older men (7, 319). We have recently shown that the acute administration of 30 g (120 kcal) and 70 g (280 kcal) oral whey-protein loads suppressed subsequent energy intake by 12–17% in young people without suppression in healthy older men (15). The whey-induced suppression of appetite and energy intake in young people may reflect an increase in pyloric and reduction in antral and duodenal motility, factors important in the regulation of gastric emptying (18, 320). Obesity appears to have minor, inconsistent, effects on gastric emptying (321- 323), whereas gastric emptying is modestly slower in older, than younger, adults (15). The potential influence of gastric emptying and energy/ protein load of an ingested meal on subsequent voluntary food intake is complex and likely to depend on the time of subsequent eating, the degree of gastric distension and initiation of small intestinal mechanisms to stimulate satiety after food leaves the stomach (25, 322, 324).

Information about the effects of whey protein on energy intake in obese, older people is limited. We have reported that the acute suppression of energy intake by whey protein, administered either intraduodenally (9), or orally (15), is less in healthy, non-obese, older than younger men, suggesting that the use of protein supplements to promote weight loss may not be as affective in older as younger adults (15). However, ad libitum energy intake responses to oral whey protein have not been evaluated in older, obese, adults. These may have implications for the use of high protein dietary strategies to manage obesity and maintain muscle mass in older people.

The primary aim of this study was to determine the effects of a 30 g whey protein drink on energy intake in obese older and younger men.

4.3 Materials and methods

Ten younger (Mean \pm standard error of mean (SEM): age: 27 ± 2 years; body weight: 112 ± 8 kg; height: 1.75 ± 2.81 m; BMI: 36 ± 2 kg/m²) and ten older (age: 72 ± 1 years; body weight: 103 ± 4 kg; height: 1.76 ± 2.56 m; BMI: 33 ± 1 kg/m²) obese men were recruited by advertisement. Body weight and BMI of the younger and older participants did not differ significantly ($P > 0.05$).

On the basis of our previous study in lean participants (9), we determined that ten participants/group would be sufficient to detect a difference in the suppression of energy intake by 30 g whey protein of 395 kcal, with standard deviations (SDs) of 316 kcal (younger participants) and 180 kcal (older participants) (9), and in 50% gastric emptying time (T50 min) of 80 min with (SD's) of 38 min (younger participants) and 63 min (older participants) (214, 215, 325), between younger and older participants, with $\alpha = 0.05$ and power of 80%.

Participants were excluded on the basis of smoking, alcohol abuse, diabetes (HbA1C $> 6\%$), significant gastrointestinal surgery, gastrointestinal symptoms (pain, reflux, diarrhoea, or constipation), and the use of medications known to affect energy intake, appetite, or gastrointestinal motor function. For older people, additional exclusion criteria were impaired cognitive function (score < 25 on Mini Mental State (326)) and depression (score ≥ 11 on the Geriatric Depression Questionnaire (327)).

The Royal Adelaide Hospital Human Research Ethics Committee approved the protocol which was conducted in accordance with the Declaration of Helsinki. The study was registered with the Australian New Zealand Clinical Trial Registry

(www.anzctr.org.au, registration number ANZCTR12616001216404). All participants provided written informed consent prior to their study inclusion.

4.3.1 Protocol

Participants were studied twice, separated by ≥ 3 days, to determine the effects of a whey protein drink (30 g/120 kcal) and a control drink (~ 0 g whey protein/ ~ 2 kcal) on energy intake, in randomized order (1 block with balanced permutations; www.randomization.com), double-blind, and cross-over design.

The protein drink (~ 450 mL) was prepared by dissolving whey protein (Bulk Nutrients, Tasmania, Australia) in demineralized water and diet lime cordial (Bickford's Australia, South Australia (SA), Australia) to achieve the desired load i.e. 30 g whey (volume of the powder: 19 mL) in 335 mL distilled water and 85 mL cordial (2.5 kcal/100 mL). The 'control' drink contained 0 g protein, 360 mL water, and 90 mL cordial. Sodium chloride, 0.3 g and 1.2 g, was added to the whey and control drinks so that the osmolarity (88 mOsm/L) was matched. To ensure even mixing drinks were stirred continuously at low speed on a stirring plate. The volumes of the drinks differed slightly (control: 450 mL; 30 g protein: 439 mL). Drinks were prepared by a research officer not involved in data analysis and served in a covered cup, so that both investigators and participants were blinded to the treatment.

Participants were provided with a standardized meal [beef lasagne (McCain Foods Pty Ltd, Wendouree, Victoria (VIC), Australia), ~ 591 kcal] to consume on the night before each study day at ~ 1900 h. They were instructed to fast overnight ~ 12 h from solids and liquids except water, and to refrain from strenuous physical activity until

they attended the laboratory at ~0830 h.

On arrival, participants were seated in a chair, where they remained throughout the study day, and an intravenous cannula was inserted. Measurements of antral area and perceptions of appetite and gastrointestinal symptoms were performed immediately before (during fasting; 0 min), immediately after ingestion of the drink (5 min), and then at 15 min intervals until 180 min. Participants consumed the drink within 2 min. Antral area was measured by 2-dimensional (2D) ultrasonography (15). Perceptions of appetite and gastrointestinal symptoms were assessed using visual analogue scales (VAS) and blood samples were collected to measure blood glucose. At 180 min, participants were given a cold, buffet-style meal, as described (15), in excess of what they were expected to consume (total energy content of 2,457 kcal; 19% protein, 50% carbohydrates, 31% fat) and instructed to eat freely for up to 30 min until comfortably full (180–210 min) (15).

4.3.2 Measurements

4.3.2.1 Energy intake

The amount eaten (g) was quantified by weighing the meal before and after consumption. Energy intake (kcal) and proportions of intake of protein, carbohydrate, and fat were calculated using commercially available software (Foodworks; 3.01, Xyris Software, Highgate Hill, Queensland, Australia). Energy intake was calculated both as intake at the buffet meal and as the cumulative energy intake (sum of energy intake at the buffet meal and from the preload drink). Absolute (kcal) and percentage suppression/change (energy intake as % of control day intake) of energy intake at the buffet meal by 30 g protein compared with control were

calculated (16).

4.3.2.2 Gastric emptying

Gastric emptying was measured at baseline, after overnight fasting ($t = 0\text{min}$), immediately after drink consumption and then every 15min to 180min. Gastric emptying rates were calculated from ultrasound measurements of gastric antral area (cm^2), as previously described (25) (292). Measurements were performed with a Logiq™ ultrasound machine (GE Healthcare Technologies, Sydney, New South Wales, Australia) using a 3.5 C broad spectrum 2.5–4MHz convex linear array transducer. The transducer was positioned vertically to obtain a para-sagittal image of the antrum with the superior mesenteric vein and the abdominal aorta in a longitudinal section. Measurements were performed at the end of inspiration. To calculate meal retention in the stomach, fasting antral area (measured at baseline) was subtracted from subsequent measurements performed after ingestion of the drinks (290). Gastric retention was calculated as

$$\text{Retention (\%)} = [\text{AA}(t) - \text{AA}(f)] / [\text{AA}(\text{max}) - \text{AA}(f)] * 100$$

where $\text{AA}(t)$ = antral area measured at a given time, $\text{AA}(f)$ = fasting antral area, and $\text{AA}(\text{max})$ = maximum antral area recorded after drink ingestion (290). When ultrasound images lacked sufficient clarity, data were imputed by linear interpolation. The time at which 50% of the preload drink had emptied from the stomach (time to halving of post-drink maximum antral area: T_{50} ; min) was calculated for both conditions.

4.3.2.3 Perceptions of appetite and gastrointestinal symptoms

Perceptions of hunger, desire to eat, prospective consumption, fullness, nausea, and bloating were rated using a 100mm visual analogue scale (VAS) questionnaire at $t = 0, 5, 15, 30, 45, 60, 75, 90, 105, 120, 135, 150, 165, 180,$ and 210 min as previously described (291). Area under the curve (AUC) was calculated for appetite ratings (0–180 min) using the trapezoidal rule.

4.3.2.4 Blood glucose

Blood glucose concentrations (mmol/L) were determined by the glucose oxidase method using a portable glucometer (Optium Xceed, Abbott Laboratories, Australia). Area under the curve (AUC) was calculated for blood glucose 0–180 min using the trapezoidal rule.

4.4 Data and statistical analysis

Statistical analyses were performed using SPSS software (version 25; IBM, Armonk, NY, USA). Effects of age and treatment and their interaction effect were determined using a repeated measures mixed-effects model. An unstructured covariance structure was used to account for the repeated treatments by subject. Suppression of energy intake by protein and outcomes of the control condition were compared between age groups with a paired t-test. Statistical significance was accepted at $P < 0.05$. All data are presented as mean values \pm SEM.

4.5 Results

The study protocol was well tolerated by all participants.

4.5.1 Energy intake

Older participants consumed ~20% less energy after the drinks than younger participants, although this difference was not statistically significant ($P = 0.16$). There was no difference in the amount of energy consumed after the protein drink compared to control, in either younger (control 1173 ± 130 kcal vs. protein 1114 ± 124 kcal) or older (control 926 ± 99 kcal vs. protein 892 ± 127 kcal) men (age effect $P = 0.16$, protein effect $P = 0.46$, age \times protein interaction effect $P = 0.84$; Figure 4.1). There was no suppression of energy intake by whey protein in either age group (younger $-3.0 \pm 7.7\%$ vs. older $-2.3 \pm 8.4\%$, $P = 0.95$).

Furthermore, cumulative energy intake (intake at buffet meal plus test drink) during the protein day did not differ with age or treatment (age effect $P = 0.16$, protein effect $P = 0.25$, age \times protein interaction effect $P = 0.84$).

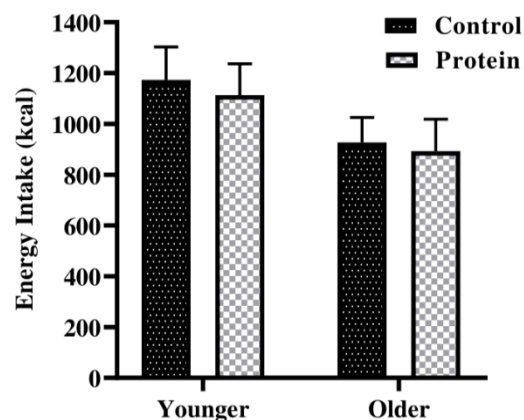


Figure 4.1: Mean \pm standard error of mean (SEM) energy intake at the buffet meal

(kcal) in younger ($n = 10$) and older ($n = 10$) men after drinks containing water (control-black) and 30 g whey protein (black border). Main age and protein effects and age \times protein interaction effect were determined by using mixed model analysis. The protein drink did not suppress subsequent energy intake at the buffet meal compared with control. The main effects of age ($P = 0.16$) and protein ($P = 0.46$) and the age \times protein interaction effect ($P = 0.84$) for energy intake were not significant.

4.5.2 Gastric emptying

In one older participant the quality of antral images was insufficient to determine gastric emptying on both days, so data for this participant was excluded from analysis. There was no difference in baseline (after overnight fasting) antral areas between age groups or study treatments (young men control vs. protein: 3.20 ± 0.37 cm² vs. 2.98 ± 0.31 cm²; older men control vs. protein: 2.68 ± 0.29 cm² vs. 3.19 ± 0.29 cm²; age effect $P = 0.64$, protein effect $P = 0.61$, age \times protein interaction effect $P = 0.23$).

The protein drink emptied more slowly than control in both groups (T_{50} young men control vs. protein: 25 ± 5 min vs. 40 ± 7 min; older men control vs. protein: 16 ± 5 min vs. 50 ± 8 min; protein effect $P = 0.001$, age effect $P = 0.93$, age \times protein interaction effect $P = 0.13$; Figure 4.2). Gastric emptying of the control drink was not significantly different between both age groups ($P = 0.21$).

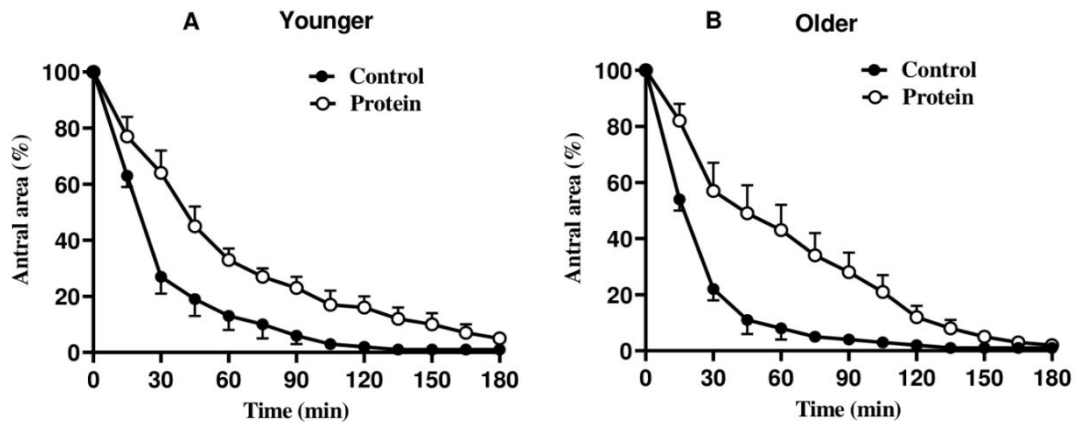


Figure 4.2: Mean \pm standard error of mean (SEM) antral area (%) in younger ($n = 10$) and older ($n = 10$) men after drinks containing water (control) and 30 g whey protein. Main age and protein effects and age \times protein interaction effect were determined by using mixed effects analysis. Gastric emptying (antral area) of the protein drink was slower than control to a similar degree in both the age groups. The main effect of protein for 50% gastric emptying time (T50 min; $P = 0.001$) was significant; the age ($P = 0.93$) and the age \times protein interaction ($P = 0.13$) effects were not significant.

4.5.3 Blood glucose concentrations

Fasting blood glucose concentrations were higher in older (control 6.1 ± 0.2 mmol/L, protein 6.2 ± 0.2 mmol/L) than younger (control 5.4 ± 0.1 mmol/L, protein 5.4 ± 0.2 mmol/L) men (age effect $P = 0.003$, protein effect $P = 0.76$, age \times protein interaction effect $P = 0.58$) and throughout both study days (AUC₀₋₁₈₀ min, young men control: 1003 ± 17 mmol/L, protein: 981 ± 21 mmol/L; older men control: 1112 ± 28 mmol/L, protein: 1108 ± 37 , age effect $P = 0.005$). There was no effect of protein on blood glucose concentrations in either age group (protein effect $P = 0.42$, age \times protein interaction effect $P = 0.54$; Figure 4.3).

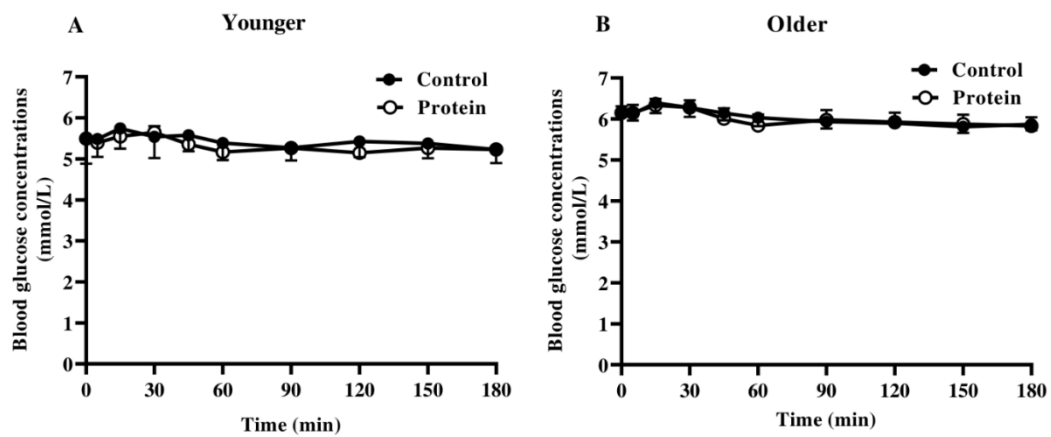


Figure 4.3: Mean \pm standard error of mean (SEM) blood glucose concentrations (mmol/L) in younger ($n = 10$) and older ($n = 10$) men after drinks containing water (control) and 30 g whey protein. Main age and protein effects and age \times protein interaction effect were determined by using mixed effects analysis. There was no effect of protein on the blood glucose concentrations in either age group. The main effect of age ($P = 0.005$) was significant; the protein ($P = 0.42$) and age \times protein interaction ($P = 0.54$) effects were not significant.

4.5.4 Perceptions of appetite and gastrointestinal symptoms

4.5.4.1 Baseline

Baseline ratings of hunger [younger men control (YC) 53 ± 9 mm, younger men protein (YP) 39 ± 9 mm; older men control (OC): 37 ± 9 mm, older men protein (OP) 44 ± 6 mm], fullness (YC 19 ± 5 mm, YP 20 ± 6 mm; OC 16 ± 6 mm, OP 9 ± 5 mm), nausea (YC 14 ± 6 mm, YP 8 ± 4 mm; OC 4 ± 1 mm, OP 5 ± 1 mm) and bloating (YC 8 ± 3 mm, YP 9 ± 5 mm; OC 7 ± 4 mm, OP 5 ± 1.9 mm) were not different between younger and older men or between control and protein days (all main effects $P > 0.05$).

There were significant but modest age \times protein interaction effects for ratings of prospective consumption and desire to eat at baseline. Pairwise comparisons for prospective food consumption (age \times protein interaction effect $P = 0.001$) showed that in the young, scores were higher on the control day than protein day, whereas in the older group, scores were higher on the protein day than control day (YC 62 ± 8 mm, YP 51 ± 8 mm, $P = 0.040$ vs. OC 41 ± 8 mm, OP 58 ± 8 mm, $P = 0.004$).

For desire to eat (age \times protein interaction effect $P = 0.020$), there was no difference between treatments in either age group ($P > 0.05$). Scores were higher in the younger than the older group on the control day ($P = 0.040$), but not on the protein day (YC 52 ± 8 mm, YP 39 ± 7 mm vs. OC 30 ± 7 mm, OP 39 ± 7 mm, $P = 0.98$).

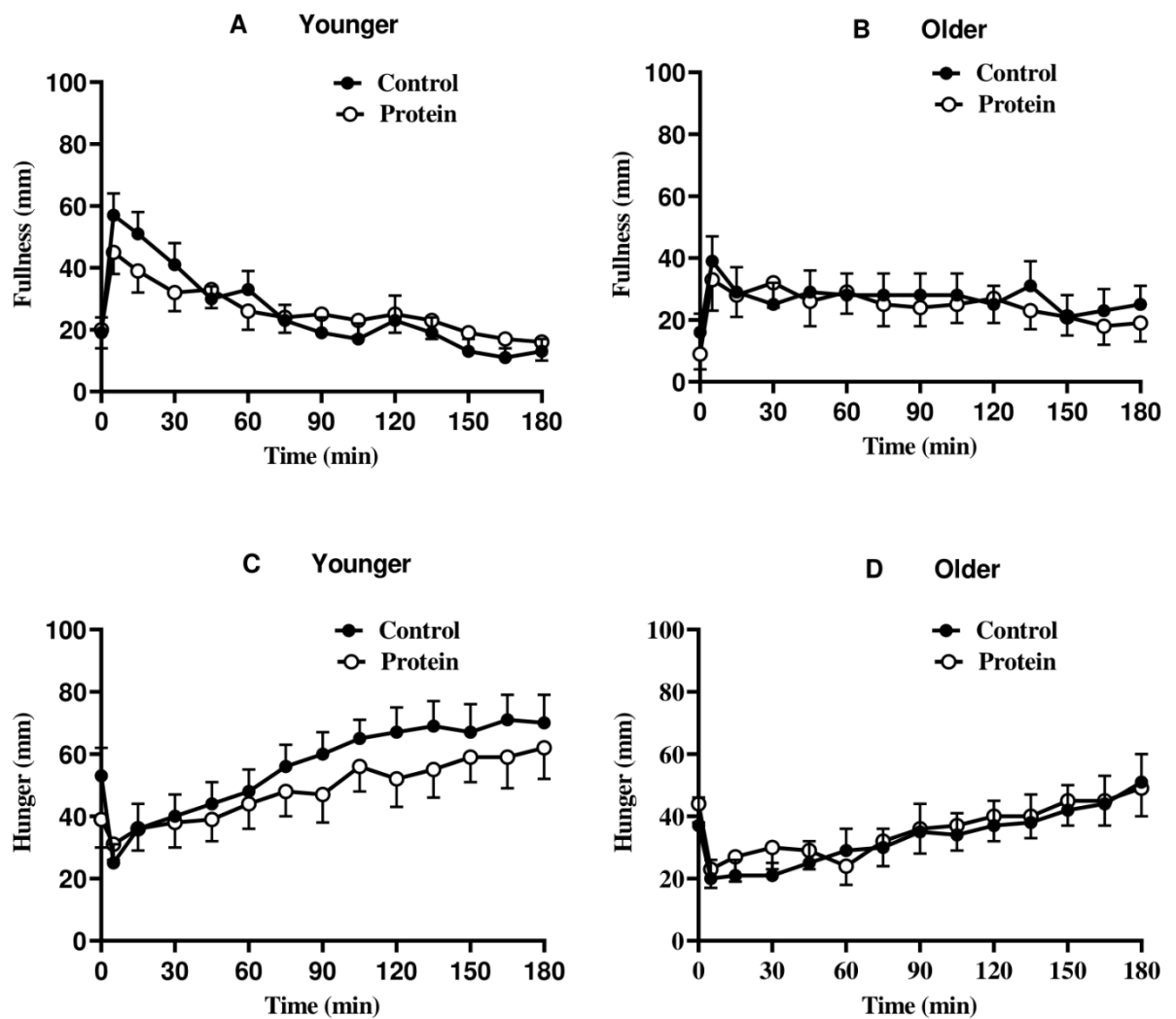
4.5.4.2 After study drink

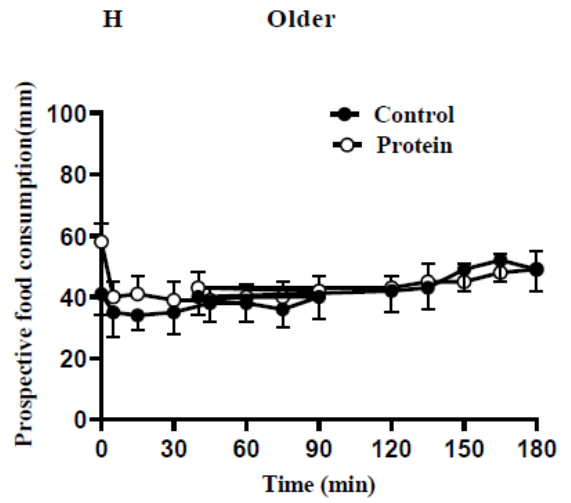
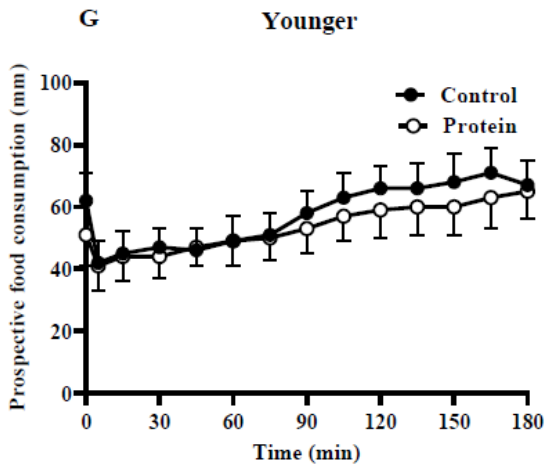
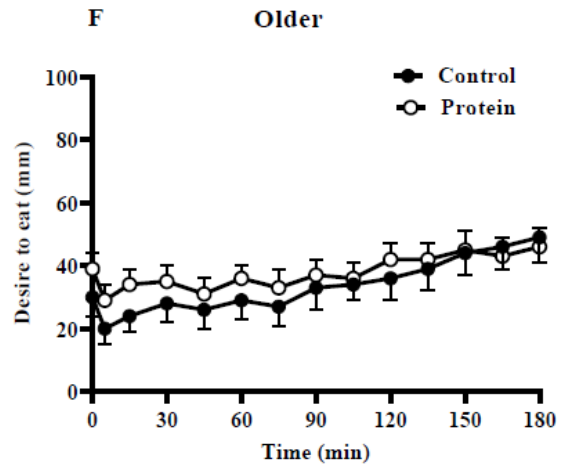
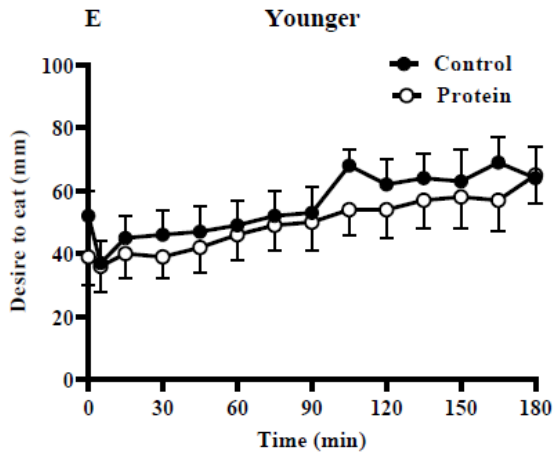
Prospective food consumption

The main age and protein effects for ratings of desire to eat (total AUC; YC $10,019 \pm 1174$ mm.min, YP $8,925 \pm 1156$ mm.min vs. OC $6,003 \pm 1173$ mm.min, OP $6,763 \pm 1156$ mm.min; age effect $P = 0.07$, protein effect $P = 0.71$), hunger (YC $10,041 \pm 1180$ mm.min, YP $8,676 \pm 1289$ mm.min vs. OC $5,873 \pm 1180$ mm.min, OP $6,324 \pm 1289$ mm.min; age effect $P = 0.07$, protein effect $P = 0.35$), fullness (YC $4,705 \pm 897$ mm.min, YP $4,703 \pm 959$ mm.min vs. OC $4,884 \pm 897$ mm.min, OP $4,703 \pm 959$ mm.min; age effect $P = 1.0$, protein effect $P = 0.57$), prospective food consumption (YC $10,279 \pm 1198$ mm.min, YP $9,572 \pm 1143$ mm.min vs. OC $7,341 \pm 1198$ mm.min, OP $7,607 \pm 1143$ mm.min; age effect $P = 0.15$, protein effect $P = 0.57$), bloating (YC $3,151 \pm 875$ mm.min, YP $2,666 \pm 653$ mm.min vs. OC $1,973 \pm 875$ mm.min, OP $1,495 \pm 653$ mm.min; age effect $P = 0.28$, protein effect $P = 0.11$) were

not significant. In younger participants nausea ratings were higher after the test drink than older participants (YC $3,127 \pm 741$ mm.min, YP $2,892 \pm 827$ mm.min vs. OC 771 ± 741 mm.min, OP 708 ± 827 mm.min; age effect $P = 0.04$). The effect of protein was not significant ($P = 0.74$).

The age \times protein interaction effect for ratings of hunger, fullness, prospective food consumption, nausea, and bloating were not significant ($P > 0.05$), although there was a strong trend for desire to eat to be less in the older participants (age \times protein interaction effect $P = 0.051$; Figure 4.4).





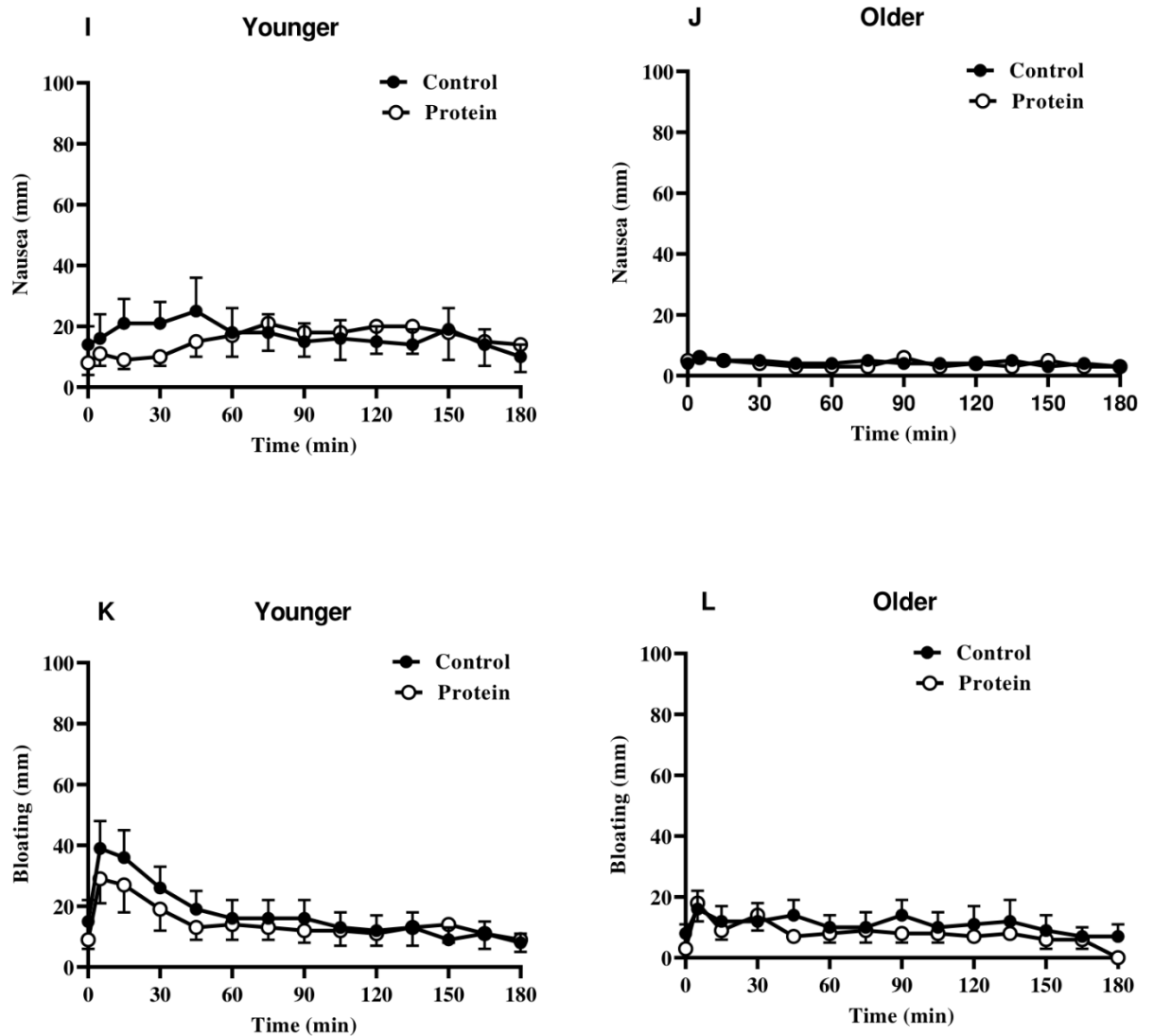


Figure 4.4 Perceptions of appetite and gastrointestinal symptoms. Mean \pm standard error of mean (SEM) visual analogue scores (VAS; mm; 0–180min) of fullness (A, B), hunger (C, D), desire to eat (E, F), prospective food consumption (G, H), nausea (I, J) and bloating (K, L) in younger ($n = 10$) and older ($n = 10$) men immediately before and after drinks containing water (control) and 30 g whey protein. Time (0–180min) effects were determined by using mixed effects analysis. Ratings of fullness, hunger, nausea, and bloating did not change from baseline during all study days in both age groups ($P > 0.05$). Scores for prospective food consumption were significantly higher in the young on the control day ($P = 0.040$), whereas in the older

group, scores were significantly higher on the protein day (protein effect $P = 0.004$; age \times protein interaction effect $P = 0.001$). For desire to eat prospective consumption was significantly higher in the younger than older group on the control day (age effect $P = 0.040$), but not on the protein day (age effect $P = 0.98$; age \times protein interaction effect $P = 0.020$).

4.6 Discussion

The major findings of this study are:

1. There was no suppression of ad libitum energy intake after 30 g whey protein ingestion in either older or young, obese men. This is in contrast to our previous studies in lean, non-obese men, where greater (and significant) suppression of energy intake was observed in younger than older men, after both a 30 g whey protein drink (1% vs. 15% suppression) (15) and intraduodenal whey infusions (1% vs. 19% suppression) (9).
2. The whey protein drink slowed gastric emptying, to a comparable degree in both age groups.
3. There was no effect of the protein drink on hunger, desire to eat, fullness, satiety and bloating.

Numerous studies, including our own, have shown that ingestion of protein, by mouth and directly into the stomach or small intestine, acutely suppresses appetite and ad libitum energy intake in young, non-obese adults. Evidence that protein is the most satiating of the macronutrients in young adults (276, 316), has led to the development of high protein (usually energy-restricted) diets and their recommendation to young, overweight people trying to lose weight. Increased

protein intake in these diets is often at the expense of reduced carbohydrate intake (328-330). Many people, however, fail to achieve substantial, long-term weight loss on such diets. One possible explanation for this is that protein is not as satiating in obese as in non-obese individuals. In the present study we examined the impact of obesity on the satiating effects of protein ingestion (15). In our previous study of young non-obese men (mean BMI 23 kg/m²) we showed that a 30 g whey protein drink significantly suppressed ad libitum energy intake at a test meal 3 h later by 17% compared to control day intake. That suppression was associated with reduced appetite ratings. In contrast, in the present study, using the same whey dose and study protocol, the protein drink suppressed neither energy intake nor appetite ratings in young obese men.

These results contrast with those of Brennan et al. (331), who reported that high protein meals suppress energy intake in both lean and obese younger men 3 h after an energy preload. While the energy preloads in that study were larger than we employed (213 kcal vs. 120 kcal), and administered as a fat, protein and carbohydrate mixture, we have reported previously that pure whey drinks of 280 kcal and mixed macronutrient drink of 504 kcal protein (280 kcal), carbohydrate (112 kcal) and fat (112 kcal) drinks also did not suppress voluntary energy intake at 3 h in non-obese older men (16). The preloads in Brennan et al. were solid food (a meat and pasta dish), compared to a whey drink in our study. There is evidence that drinks are less satiating than solid foods of the same nutrient composition (331). Mourao et al., for example, reported greater ad libitum energy intake after drinks than solid food of comparable energy content (332). These results are consistent with the possibility of a reduced satiating effect of liquid vs. solid protein. This would support the use of increased oral protein in solid rather than liquid form when the intention is to promote

weight loss. Nevertheless, the whey protein drink did suppress appetite and food intake in young, non-obese men in our previous study, an effect not present in the obese young men in this study. These results suggest that obesity may blunt the satiating effects of protein, at least whey protein, and that these effects of obesity may be similar to those of physiological ageing; both healthy ageing and obesity have been associated with a loss of suppression of subsequent food intake by a whey drink.

The mechanism(s) by which obesity blunts the satiating effects of whey protein are not known. Blood glucose concentrations were predictably higher in the older than younger men, but not affected by protein ingestion, as in our previous study of non-obese men (15), suggesting that glucose was not involved.

Gastric emptying was slowed by ingestion of the whey protein in both younger and older men with no difference between them. Despite this, subsequent food intake was suppressed by whey in neither age group. This suggests, at least under these study conditions, that gastric distension due to retention of the protein load in the stomach and delayed entry of test drink into the small intestine, did not affect appetite and food intake. To allow a comprehensive assessment of gastric emptying the test meal was started 180 min after the study drink, by which time the stomach was almost completely empty. It is possible that if the test meal was given earlier, when the difference between the study days in how much drink remained in the stomach was greater, there may have been reduced food intake on the whey drink day and an association between food intake and gastric emptying. Nevertheless, further evidence for the lack of such an association is provided by the results of our previous study in non-obese men (15), in which gastric emptying was markedly slowed by the whey drink in both older and young men, with almost complete emptying of the drink from

the stomach before buffet meal ingestion at 180 min. In that study, there was a reduction in ad libitum energy intake in the non-obese young men, suggesting that slowing of gastric emptying by protein ingestion is not involved in the satiating effect (or lack of) of the protein drinks under these study conditions.

Increasing age is associated with a physiological reduction in hunger and food intake, the so-called “anorexia of ageing” (214, 325). Consistent with that reduction, the older participants consumed about 20% less energy in this study than the younger participants, although that reduction was not statistically significant. On average, body weight decreases after about age 60–70 years. Much of the weight lost is lean tissue, including skeletal muscle and bone (243, 333). Muscle loss can be marked, particularly when pathological processes are superimposed, and can lead to sarcopaenia and frailty. One way to prevent this skeletal muscle loss, and thus preserve function and quality of life, is with dietary protein supplements, that can help maintain lean body mass and improve health (334). In contrast, when body weight is deliberately lost with energy-reduced diets, lean tissue is lost as well as fat. This may have adverse effects in older people with baseline low muscle mass. The results of this and our previous acute studies suggest that protein supplementation as whey protein drinks can be given to older people as an aid to skeletal muscle preservation and even augmentation, without risk of appetite suppression and weight loss. Conversely, such protein drinks are likely to have little effect to promote weight loss in overweight, older people trying to lose weight, and are likely to be even less effective than in younger adults. Longer-term studies in both obese and non-obese older adults evaluating the effects on appetite and food intake of increased protein intake in solid foods would be of interest.

Limitations of this study include the relatively modest participant numbers and the use of only one dose of whey protein. We did not assess the participants' perceptions of taste and/or pleasantness of the drinks. Gastric emptying rate was measured indirectly by 2D Ultrasound. 3D ultrasonography, which we have used previously (15, 230, 335) is probably more accurate in non-obese people, but was not used here as the results are less reliable in obese individuals due to tissue layers. Scintigraphy is the 'gold standard' technique for measurement of gastric emptying but was not available for this study. We studied only men, as they appear to have the greatest ability to regulate energy intake in response to energy manipulation (320). The results do not necessarily apply to women.

In summary a 30 g whey protein drink did not suppress appetite or energy intake in obese younger or older men. We speculate that obesity might mimic the effects of ageing to inhibit responses to protein. The use of liquid whey supplementation is unlikely to be helpful as a weight loss strategy in men of any age. Liquid whey protein supplements, the use of which may be of benefit for the preservation of function in older people, are unlikely to suppress food intake in older people of any weight.

**CHAPTER 5: COMPARATIVE EFFECTS OF CO-INGESTING
WHEY PROTEIN AND GLUCOSE ALONE AND COMBINED
ON BLOOD GLUCOSE, PLASMA INSULIN AND GLUCAGON
CONCENTRATIONS IN YOUNGER AND OLDER MEN**

**Oberoi A, Giezenaar C, Ridga RS, Lange K, Horowitz M, Jones KL,
Chapman I, Soenen S**

Published in *Nutrients*

2022

STATEMENT OF AUTHORSHIP

Title of the paper	Comparative effects of co-ingesting whey protein and glucose alone and combined on blood glucose, plasma insulin and glucagon concentrations in younger and older men.
Publication status	Published
Publication details	Oberoi AO, Giezenaar C, Rigda RS, Lange K, Horowitz M, Jones KL, Chapman I, Soenen S. <i>Comparative effects of co-ingesting whey protein and glucose alone and combined on blood glucose, plasma insulin and glucagon concentrations in younger and older men</i> . <i>Nutrients</i> 2022; 14: 3111. doi:10.3390/nu14153111.

Candidate	Avneet Oberoi		
Contribution	Conducted the clinical trial, contributed to the overall design of the manuscript, literature review, data analysis and interpretation, drafting and revision of the manuscript.		
Overall percentage	75%		
Certification	This paper reports on original research I conducted during the period of my Higher Degree by Research candidature and is not subject to any obligations or contractual agreements with a third party that would constrain its inclusion in this thesis. I am the primary author of this paper.		
Signature		Date	October 2022

Principal Author***Co-Author Contributions***

By signing the Statement of Authorship, each author certifies that:

- i) the candidate's stated contribution to the publication is accurate (as detailed above);
- ii) permission is granted for the candidate to include the publication in the thesis; and

- iii) the sum of all co-author contributions is equal to 100% less the candidate's stated contribution.

Name of Co-Author	Caroline Giezenaar		
Contribution	Drafting and revision of the manuscript		
Signature		Date	October 2022
Name of Co-Author	Rachael S Rigda		
Contribution	Data collection, drafting and revision of the manuscript		
Signature		Date	October 2022
Name of Co-Author	Kylie Lange		
Contribution	Conception and design of the manuscript, statistical analysis, data interpretation, drafting and revision of the manuscript.		
Signature		Date	October 2022
Name of Co-Author	Michael Horowitz		
Contribution	Conception and design of the manuscript, drafting and revision of the manuscript.		
Signature		Date	October 2022
Name of Co-Author	Karen L Jones		
Contribution	Conception and design of the manuscript, data interpretation, drafting and revision of the manuscript.		
Signature		Date	October 2022

Name of Co-Author	Ian Chapman		
Contribution	Conception and design of the manuscript, statistical analysis, data interpretation, drafting and revision of the manuscript.		
Signature		Date	October 2022
Name of Co-Author	Stijn Soenen		
Contribution	Conception and design of the study, data interpretation, statistical analysis, drafting of the manuscript and overall responsibility for the study.		
Signature		Date	October 2022

5.1 Abstract

The ingestion of dietary protein with, or before, carbohydrate may be a useful strategy to reduce postprandial hyperglycemia, but its effect in older people, who have an increased predisposition for type 2 diabetes, has not been clarified. Blood glucose, plasma insulin and glucagon concentrations were measured for 180 min following a drink containing either glucose (120 kcal), whey-protein (120 kcal), whey-protein plus glucose (240 kcal) or control (~2 kcal) in healthy younger ($n = 10$, 29 ± 2 years; 26.1 ± 0.4 kg/m²) and older men ($n = 10$, 78 ± 2 years; 27.3 ± 1.4 kg/m²). Mixed model analysis was used. In both age groups the co-ingestion of protein with glucose (i) markedly reduced the increase in blood glucose concentrations following glucose ingestion alone ($P < 0.001$) and (ii) had a synergistic effect on the increase in insulin concentrations ($P = 0.002$). Peak insulin concentrations after protein were unaffected by ageing, whereas insulin levels after glucose were lower in older than younger men ($P < 0.05$) and peak insulin concentrations were higher after glucose than protein in younger ($P < 0.001$) but not older men. Glucagon concentrations were unaffected by age. We conclude that the ability of whey-protein to reduce carbohydrate-induced postprandial hyperglycemia is retained in older men and that protein supplementation may be a useful strategy in the prevention and management of type 2 diabetes in older people.

5.2 Introduction

Type 2 diabetes mellitus is a major and increasing problem worldwide (336-338). The prevalence of diabetes increases with age; in Australia, from less than 0.3% of those under age 35 years to 14–16% in those 65 years or more (339). Postprandial hyperglycemia is a major determinant of overall glycemic control—when HbA1C is < 8% postprandial glycemic excursions are the dominant determinant of HbA1C (340-343)—and an independent risk factor for cardiovascular disease (344, 345). While ingestion of protein on its own has little effect on blood glucose concentrations (90, 346), co-ingestion of dietary protein with other macronutrients may influence postprandial blood glucose concentrations when combined with carbohydrate. In non-elderly adults with and without type 2 diabetes, the oral ingestion of proteins or their component amino acids substantially reduces the increase in postprandial blood glucose concentrations when compared with carbohydrate ingestion alone (69, 147, 347-350), with different proteins and amino acids varying in their ability to have this effect (44, 346, 351, 352). Whey is a protein with potent glucose-lowering action (44, 86, 138, 350, 351, 353, 354) and there has been increasing interest in the possible benefits of whey protein in lowering blood glucose concentrations when ingested with a meal or as a preload (86, 147).

Human ageing is characteristically associated with a loss of skeletal muscle mass and function; when severe this leads to sarcopenia and other adverse outcomes (63). Increasing protein ingestion can preserve and increase muscle mass and function in older people and protein supplements have been shown to reduce morbidity and mortality in the elderly (63). Whey protein is ideal for this anabolic effect as it is rich in the branched chain amino acids, particularly leucine (249). Due to age-related

anabolic resistance, more protein per serve is required to stimulate muscle protein synthesis in older than younger adults (63) and a minimum of 25–30 g protein per meal has been recommended for older people (97).

Ageing is associated with changes in glucose metabolism, including impaired glucose tolerance and reduced insulin secretion (355-357). There may also be changes in the responses to dietary protein, including differing effects of both oral (12, 15, 90) and intraduodenal (9) whey protein administration on appetite and gut hormone release. Whey protein, when consumed orally alone, slows gastric emptying load dependently in healthy young men (18) and suppresses appetite less in healthy older men than younger men (9). It is possible, therefore, that the capacity of whey protein to attenuate the rise in carbohydrate-induced “postprandial” blood glucose concentrations may differ between older people and young adults. Previous studies examining the effects of protein ingestion on blood glucose concentrations after carbohydrate ingestion have been conducted in young- and middle-aged adults (44, 86, 138, 350, 351, 353, 354). We conducted this study to determine whether the comparative effects of whey protein/carbohydrate co-ingestion on postprandial blood glucose concentrations persists into old age. We used a whey protein dose of 30g because of its tolerability in older people and likely beneficial effects on skeletal muscle.

5.3 Materials and methods

5.3.1 Participants

This was a randomized double-blind cross-over study comprising 10 healthy younger men (age range 18–35 years, mean \pm SEM age: 29 \pm 2 years; body weight: 84 \pm 4

kg; height: 1.75 ± 0.02 m; body mass index (BMI): 26.1 ± 0.4 kg/m²) and 10 healthy older men (68–87 years, 78 ± 2 years; 82 ± 2 kg; 1.76 ± 0.02 m; 27.3 ± 1.4 kg/m²). The body weight and the BMI of the younger and the older men did not differ significantly ($P > 0.05$).

The participants were recruited by online advertisement and by flyers placed on notice boards at the University of Adelaide, Australia.

The exclusion criteria included smoking, alcohol intake of > 2 standard drinks on > 5 days per week, being vegetarian, intake of any illicit substance, use of prescribed or non-prescribed medications which may affect appetite, body weight, gastrointestinal function or energy metabolism, food allergy(s), known diabetes mellitus (or fasting blood glucose concentration > 6.9 mmol/L), epilepsy, gallbladder, pancreatic, known cardiovascular or respiratory disease, significant gastrointestinal symptoms, disease or surgery, any other illness deemed significant by the investigator and inability to comprehend the study protocol. Inclusion criteria included being weight stable ($< 5\%$ fluctuation in weight) at study entry, as assessed by self-reported weight in the preceding 3 months, and willingness to maintain usual physical activity level throughout the study.

The Royal Adelaide Hospital Human Research Ethics Committee approved the protocol, which was conducted in accordance with the Declaration of Helsinki. The study was registered with the Australian New Zealand Clinical Trial Registry (www.anzctr.org.au (2 June 2022), registration number ACTRN12619000420145). All participants provided written informed consent prior to their study inclusion.

5.3.2 Protocol

Each participant was studied on four occasions, separated by ~7–10 days. On each occasion, they received, in randomized order (using the method of randomly permuted blocks; www.randomization.com), a drink of either 30 g glucose (G, 120 kcal) (glucose monohydrate, Sigma-Aldrich, St. Louis, MI, USA), 30 g whey protein (P, 120 kcal) (Bulk Nutrients, Tasmania, Australia), 30 g whey protein plus 30 g glucose (GP, 240 kcal) or flavored water (control, C; ~2 kcal). The effects on blood glucose, plasma insulin, glucagon, gastric emptying, energy intake and perceptions of appetite were evaluated. The drinks were equivolemic (~250 mL) and were prepared by a research assistant who was not involved in the analysis of the study results. They were flavored with varying amounts of distilled water, sodium chloride and light lime cordial (Bickford's Australia Pty Ltd., Adelaide, SA, Australia) and 100 mg [¹³C] sodium acetate to match for taste and served in a covered cup to achieve blinding. To ensure that all ingredients were dissolved evenly throughout, and to minimize the layer of foam on top of the solution, the drinks were stirred continuously at low speed on a stirring plate. Both the investigators conducting the study and the participants were blinded to the drink composition.

Participants were told to consume the same meal on the night before each study day at around 1900 h. They were instructed to fast overnight from solids and liquids and to refrain from strenuous physical activity and alcohol for 24 h prior to their attendance at the laboratory at the Clinical Research Facility, Adelaide Health and Medical Sciences Building, the University of Adelaide at 08:30 am. Upon arrival, participants were seated in a chair, and a cannula was inserted in an antecubital vein for blood sampling. A heated pad was used so that the samples were arterialized.

Once baseline measurements were taken (blood samples, breath sample and appetite ratings), participants were instructed to consume the test drink within 2 min. Gastric emptying (% intragastric retention) of the drink was measured with a ^{13}C -sodium acetate breath test $t = 5, 10, 15, 20, 25, 30, 35, 40, 45, 50, 55, 60, 75, 90, 105, 120, 135, 150, 165$ and 180 min. Perceptions of appetite were assessed with validated visual analog scales and blood samples were collected into ice chilled EDTA-coated tubes for the measurement of blood glucose, plasma insulin and plasma glucagon concentrations at $t = 0, 15, 30, 45, 60, 90, 120, 150$ and 180 min. No inhibitors were added (33). At 180 min, each participant was presented with a standardized cold buffet-style meal, in excess of what they were expected to consume (total energy content of 2457 kcal: 19% protein, 50% carbohydrates, 31% fat) (36), in a room by themselves to limit external distractions, and were allowed to eat freely for 30 min (180 – 210 min) until comfortably full.

5.3.3 Measurements

5.3.3.1 Blood glucose and plasma insulin and glucagon concentrations

Blood glucose concentrations (mmol/L) were quantified by the glucose oxidase method using a glucose analyser (YSI 2900 Stat Plus, Yellow Springs Instruments, Yellow Springs, OH, USA). Intra- and inter-assay coefficient of variations (CVs) were $\leq 2\%$. Plasma was obtained by centrifugation for 15 min, at 3200 rpm at 4 °C and samples were stored at -80 °C for further analysis of insulin and glucagon concentrations. Total plasma insulin (milliunits per liter) was measured by enzyme-linked immunosorbent assay (ELISA) immunoassay (10-1113; Mercodia, Uppsala, Sweden). The sensitivity of the assay was 1.0 mU/L and the coefficient of variation was 2.9% within and 11.6% between assays (358). Plasma glucagon concentrations

were measured by ELISA immunoassay (10-1271-01, Mercodia, Uppsala, Sweden). The lower limit of quantification (LLOQ) was 1.5 pmol/L and the detection limit was 0.75 pmol/L. The coefficient of variation was 9.3% within assays and 7.5% between assays.

5.3.3.2 Gastric emptying

As the stomach empties the test drink containing [¹³C] sodium acetate, the substrate is rapidly absorbed in the proximal small intestine and metabolized in the liver. ¹³CO₂ is produced, which can then be measured in the breath, thus reflecting gastric emptying of nutrients (359). The participants were asked to exhale through a mouthpiece to collect an end-expiratory breath sample into a 100 mL foil bag at certain time intervals. The breath samples were collected and the quantity of ¹³CO₂ (disintegrations per min) was measured by a non-dispersive infrared spectrometer (FANci2, Fischer ANalysen Instrumente, Leipzig, Germany) (360). The gastric half-emptying time (T₅₀) and the intragastric retention were calculated using the Wagner–Nelson method (361). This method has been shown to be of comparable accuracy to scintigraphy in the measurement of the gastric emptying of both solid and liquid meals (242, 362).

5.3.3.3 Energy intake

The energy consumed at the buffet meal (g) was quantified by weighing the food before and after consumption. The energy intake was calculated as absolute (kcal) at the buffet meal using commercially available software (Foodworks version 8; Xyris Software Pty Ltd, Highgate Hill, QLD, Australia (15)).

5.3.3.4 Perceptions of appetite

The perceptions of hunger and fullness were rated using a 100 mm visual analog scale questionnaire immediately before the test drinks and at $t = 15, 30, 45, 60, 90, 120, 150$ and 180 min as described (355).

5.4 Data and statistical analysis

Statistical analyses were performed using SPSS software (version 25; IBM, Armonk, NY, USA). Sample size was based on the statistical power functions of the between-group contrasts of older versus younger with overall $P = 0.05$, statistical power of 0.8 and anticipated drop-out rate of $\sim 10\%$, and significance levels adjusted to account for the 4 comparisons (control, whey protein, glucose, whey protein plus glucose). Calculations were performed for the primary outcome of area under the curve (AUC) of blood glucose concentrations, assuming a within-participant SD of 0.5 mmol/L, and a between-participants SD of 1.4 mmol/L (14, 15, 90) to detect a difference between groups of 1.5 mmol/L and a difference between treatments of 0.4 mmol/L.

The net incremental area under the curve (Net iAUC_{0–180/min}), peak, time to peak and time to return to baseline was calculated for blood glucose, insulin and glucagon. The Net iAUC_{0–180/min} was calculated from baseline using the trapezoidal rule and then divided by time (min) to present a weighted average value over the time interval.

The main effects of age and drink condition, and their interaction effects, on blood glucose, plasma insulin and glucagon concentrations were determined using a mixed-effects model with the drink condition as the within-participant factor and age as the

between- participant factor, including baseline values at each treatment visit as a covariate. Post hoc comparisons, which were adjusted for multiple comparisons with Bonferroni correction, were performed when there were significant drink-condition or interaction effects. Peak concentrations, time to peak and time to return to baseline were determined for blood glucose, plasma insulin and glucagon concentrations in those drink conditions associated with changes from baseline using a repeated measures ANOVA test (G and GP for glucose; G, P and GP for insulin and glucagon). When the time to return to baseline was between 2 blood sampling time points, an interpolated value was estimated assuming a linear relationship between the 2 time points. If a blood glucose ($n = 5$ of 80 study days) or plasma insulin ($n = 8$) or glucagon concentration ($n = 20$) did not return to baseline by 180 min, the time to return to baseline was calculated using a linear extrapolation from the values at 150–180 min. We hypothesized that insulin concentrations following both G and P drinks would increase and, to determine whether the effect on insulin of the combination drink is synergistic or additive, we compared the effect of combined whey protein and glucose (GP drink) on the rise in plasma insulin concentrations with that of the sum of the effects of glucose and whey protein alone (G drink + P drink) on insulin concentrations using a paired t-test. The inhibition of the increase in blood glucose concentrations following glucose ingestion by whey protein when compared with glucose ingestion alone was calculated as the difference between the Net iAUC of G and GP as a percentage of the Net iAUC of G. The increase in plasma insulin concentrations following GP when compared with G ingestion alone was calculated as the difference between the Net iAUC of GP and G as a percentage of the Net iAUC of G. The increase in plasma glucagon concentrations following P when compared with G ingestion alone was calculated as the difference between the Net iAUC of P and G as a percentage of the Net iAUC of G. Statistical significance

was accepted at $P < 0.05$. Data are presented as mean values \pm SEM.

5.5 Results

The study protocol was well tolerated by all participants and no adverse effects were reported.

5.5.1 Blood glucose

Baseline glucose concentrations did not differ between age groups ($P = 0.68$) or study days ($P = 0.24$, Table 5.1).

5.5.1.1 Interaction effects

The time to peak glucose concentrations were longer after G in older than younger men ($P = 0.03$, Table 5.1). The age by drink interaction effect was non-significant for all other outcomes.

5.5.1.2 Drink-condition effects

Blood glucose concentrations increased from baseline following glucose ingestion, alone (G) and when combined with whey protein (GP) (both $P < 0.05$), to peak mean concentrations of 6.3–8.1 mmol/L (Table 5.1) but did not change following the control (C) or protein (P) drinks.

Co-ingestion of whey protein with glucose reduced the peak and the Net iAUC blood

glucose concentrations when compared with glucose ingestion alone (both $P < 0.001$), in both older and younger men, without any effect of age on this inhibitory effect ($P > 0.05$). Peak blood glucose concentrations occurred earlier after GP than G ($P < 0.001$) and returned to baseline at a comparable time after GP and G ($P = 0.83$, Figure 5.1).

The inhibition of the increase in blood glucose concentrations following glucose ingestion by whey protein, when compared with glucose ingestion alone, was $44 \pm 38\%$ over 3 h ($P < 0.001$; $50 \pm 13\%$ in the first hour $P < 0.001$, $66 \pm 33\%$ in the second hour $P = 0.001$ and $-1 \pm 23\%$ in the third hour $P = 0.21$).

5.5.1.3 Age effects

Peak glucose concentrations after GP and G drinks were non-significantly higher in older than younger men (Table 5.1). Peak glucose concentrations occurred later after GP and G in the older than the younger men ($P = 0.007$) and took longer to return to baseline in the older men ($P = 0.03$).

There was no effect of age on the inhibitory effect of whey protein (GP) on the rise in blood glucose compared with after glucose alone (G). The inhibition of the rise in blood glucose concentrations over 3 h was $62 \pm 27\%$ in the older men and $32 \pm 92\%$ in the younger men ($P = 0.75$) and $41 \pm 6\%$ in the older men and $53 \pm 8\%$ in the younger men in the first hour after drink ingestion ($P = 0.46$).

Table 5.1: Blood glucose, plasma insulin and plasma glucagon concentrations following control drink (C), 30g whey protein drink (P), 30g glucose drink (G) and 30g whey protein plus 30g glucose drink (GP) ingestion

	Younger men				Older men				P value age effect	P value drink-condition effect	P value interaction effect
	C	P	G	GP	C	P	G	GP			
Blood glucose (mmol/L)											
Fasted	4.3±0.1	4.3±0.2	4.2±0.2	4.5±0.1	4.5±0.1	4.2±0.2	4.4±0.2	4.5±0.1	0.68	0.24	0.23
Peak	4.7 ±0.1	4.8 ±0.1	7.8±0.3	6.3±0.3	4.9 ±0.1	4.7 ±0.1	8.1±0.3	6.9±0.3	0.27	<0.001	0.54
Time to peak (min)	50±13	68±17	34±3 ¹	30±3	26±13	75±17	51±3 ¹	36±3	0.007	<0.001	0.031
Return to baseline (min)	88±26	41±13	106±15	91±18	60±22	125±23	132±15	153±18	0.03	0.83	0.23
Net iAUC _{0-60min}	-0.1±0.1	0.0±0.1	2.1±0.2	0.9±0.2	0.1±0.1	0.1±0.1	2.1±0.2	1.3±0.2	0.29	<0.001	0.48
Net iAUC _{0-180min}	-0.1±0.1	0.0±0.1	0.7±0.2	0.3±0.1	-0.1±0.1	0.1±0.1	1.1±0.2	0.7±0.2	0.08	<0.001	0.27
Plasma insulin (mU/L)											

*Comparative effects**Chapter 5*

Fasted	5.3±1.0	4.8±0.8	6.9±1.1	6.2±1.0	4.1±1.0	4.5±0.8	4.1±1.1	4.5±1.0	0.22	0.41	0.13
Peak	11.7±1.9	33.2±6.9 ²	55.9±7.3 ²	93.3±16.8	9.7±1.9	31.7±6.9	36.5±7.3	72.7±16.9	0.34	<0.001	0.006
Time to peak (min)	35±13	32±3 ³	33±2 ³	36±3	32±4	48±3 ³	46±2 ³	42±3	0.001	0.91	0.038
Return to baseline (min)	73±26	132±8	112±12	160±11	109 ±18	149 ±8	163±12	166±11	0.011	0.068	0.14
Net iAUC _{0-60min}	1.8±0.5	16.5±3.8	27.1±4.4	46.2±9.3	2.3±0.5	15.5±3.8	20.9±4.4	39.8±9.3	0.59	<0.001	0.26
Net iAUC _{0-180min}	0.2±0.6	8.3±1.9	10.7±2.4	25.8±6.2	1.5±0.6	9.6±1.9	11.6±2.4	26.6±6.2	0.77	<0.001	0.99

Plasma glucagon

(pg/mL)

Fasted	9.7±1.4	10.9±1.6	9.8±1.4	10.3±1.5	8.7±1.4	9.1±1.6	8.1±1.4	9.6±1.5	0.51	0.27	0.69
Peak/Nadir	11.2±1.5	36.4±5.8	10.7±1.4	22.6±3.1	10.2±1.5	33.1±5.8	9.1±1.4	22.1±3.1	0.71	<0.001	0.73
Time to peak (min)	68±22	57±17	68±24	54±9	65±22	51±17	56±24	48±9	0.83	0.65	0.98
Return to baseline (min)	54±16	176±153	71±29	176±51	126±39	391±153	133±29	277±51	0.11	0.011	0.69
Net iAUC _{0-60min}	-0.7±0.0	16.3±2.6	-4.6±0.7	6.3±1.5	-0.1±0.0	13.5±2.6	-2.4±0.7	6.3±1.5	0.98	<0.001	0.33

Net iAUC _{0-180min}	-0.9±0.3	10.8±1.3	-3.9±0.8	5.2±1.3	-0.5±0.3	9.9±1.3	-3.2±0.8	5.1±1.3	0.95	<0.001	0.92
------------------------------	----------	----------	----------	---------	----------	---------	----------	---------	------	--------	------

Blood glucose (mmol/L), plasma insulin (mU/L) and glucagon (pg/mL) concentrations fasted (baseline), peak, time to peak (min), return to baseline (min), Net iAUC_{0-60min} (change from baseline area under the curve during the first hour), Net iAUC_{0-180min} (change from baseline area under the curve during the three hours) following drink ingestion containing (i) flavored water (C, control, ~2 kcal), (ii) 30 g whey protein (P), (iii) 30 g glucose (G) or (iv) 30 g whey protein + 30 g glucose (GP) in younger and older men. Effects of age and drink condition and the interaction effect were determined using a mixed-effect model using G and GP for glucose and P, G and GP for insulin and glucagon. ¹The age × drink-condition interaction, $P = 0.031$ (time to peak glucose is longer after G in older than younger men). ²The age × drink-condition interaction, $P = 0.006$ (peak insulin concentration is higher after G than P in younger but not older men). ³The age drink-condition interaction, $P = 0.038$ (time to peak insulin is longer after P and G in older than younger men). Drink-condition effects: GP vs. G reduced peak ($P < 0.001$) and Net iAUC ($P < 0.001$) glucose concentrations and the peak occurred earlier ($P < 0.001$) in both older and younger men: P, G, GP vs. C increased Net iAUC insulin concentrations ($P < 0.001$); G vs. C decreased and P, GP vs. C, P vs. GP increased Net iAUC glucagon concentrations ($P < 0.001$). Age effects: older vs. younger later peak ($P = 0.007$) and return to baseline ($P = 0.030$) glucose concentrations; older vs. younger later peak ($P = 0.001$) and return to baseline ($P = 0.01$) insulin concentrations.

5.5.2 Plasma insulin

Baseline insulin concentrations did not differ between age groups ($P = 0.22$) or study days ($P = 0.41$, Table 5.1).

5.5.2.1 Interaction effects

The peak insulin concentration was higher after G than P in younger but not older men ($P = 0.006$). Time to peak insulin was longer after P and G in older than younger men ($P = 0.038$). The age by drink-interaction effect was non-significant for all other outcomes.

5.5.2.2 Drink-condition effects

Plasma insulin concentrations (Net iAUC) increased compared with the control following glucose (G), whey protein (P) and their combined ingestion (GP, $P < 0.001$). Co-ingestion of whey protein with glucose (GP) had a more than additive effect on the increase in plasma insulin concentrations (Net iAUC_{0–180/min} GP vs. G plus P: 28.3 ± 4.7 vs. 21.3 ± 3.2 mU/L*min, $P = 0.002$, Figure 5.1). Time to peak and time to return to baseline (see age effects below) did not differ between P, G and GP ($P > 0.05$).

5.5.2.3 Age effects

Peak insulin concentrations were (non-significantly) lower after G and GP in older than younger men ($P = 0.34$). Peak plasma insulin concentrations occurred later in

older than younger men ($P = 0.001$). The time taken to return to baseline after the caloric drinks was longer in older than younger men ($P = 0.01$). GP increased plasma insulin concentrations compared with G plus P; $22 \pm 1\%$ in the older men (Net iAUC_{0–180min} 27 ± 6 vs. 21 ± 5 mU/L*min, $P = 0.02$) and $27 \pm 1\%$ in the younger men (26 ± 6 vs. 19 ± 3 mU/L*min, $P = 0.04$). Peak plasma insulin concentrations were $50 \pm 1\%$ increased by GP compared with G in the older men and $41 \pm 1\%$ in the younger men ($P < 0.001$, Table 5.1).

5.5.3 Plasma glucagon

Baseline glucagon concentrations did not differ between age groups ($P = 0.51$) or during the study days ($P = 0.27$, Table 5.1).

5.5.3.1 Interaction effects

The age by drink-interaction effect was non-significant for all outcomes.

5.5.3.2 Drink-condition effects

Plasma glucagon concentrations (Net iAUC) decreased following glucose when compared with the control ($P < 0.001$), increased following combined GP ($P < 0.001$) and increased even more after whey protein alone ($P < 0.001$, Table 5.1, Figure 5.1). The increase in glucagon concentrations from baseline was greater after P than GP ($P < 0.001$).

Whey protein increased plasma glucagon concentrations by $27 \pm 1\%$ more than G

over 3 h on average ($P < 0.001$): $36 \pm 1\%$ more than G in the first hour ($P < 0.001$), $26 \pm 1\%$ more than G in the second hour ($P < 0.05$) with a comparable increase in the third hour ($P = 0.85$). Peak glucagon concentrations were higher after P than GP in both age groups to a similar degree ($P < 0.001$, age by drink-condition effect, $P = 0.58$, Figure 5.1). Time to peak/nadir after G and GP and nadir after G did not differ between treatments ($P = 0.65$). The time taken to return to baseline after the drinks was longer after P than GP or G ($P = 0.01$).

5.5.3.3 Age effects

Glucagon concentrations were not affected by age ($P = 0.71$).

Younger men

Older men

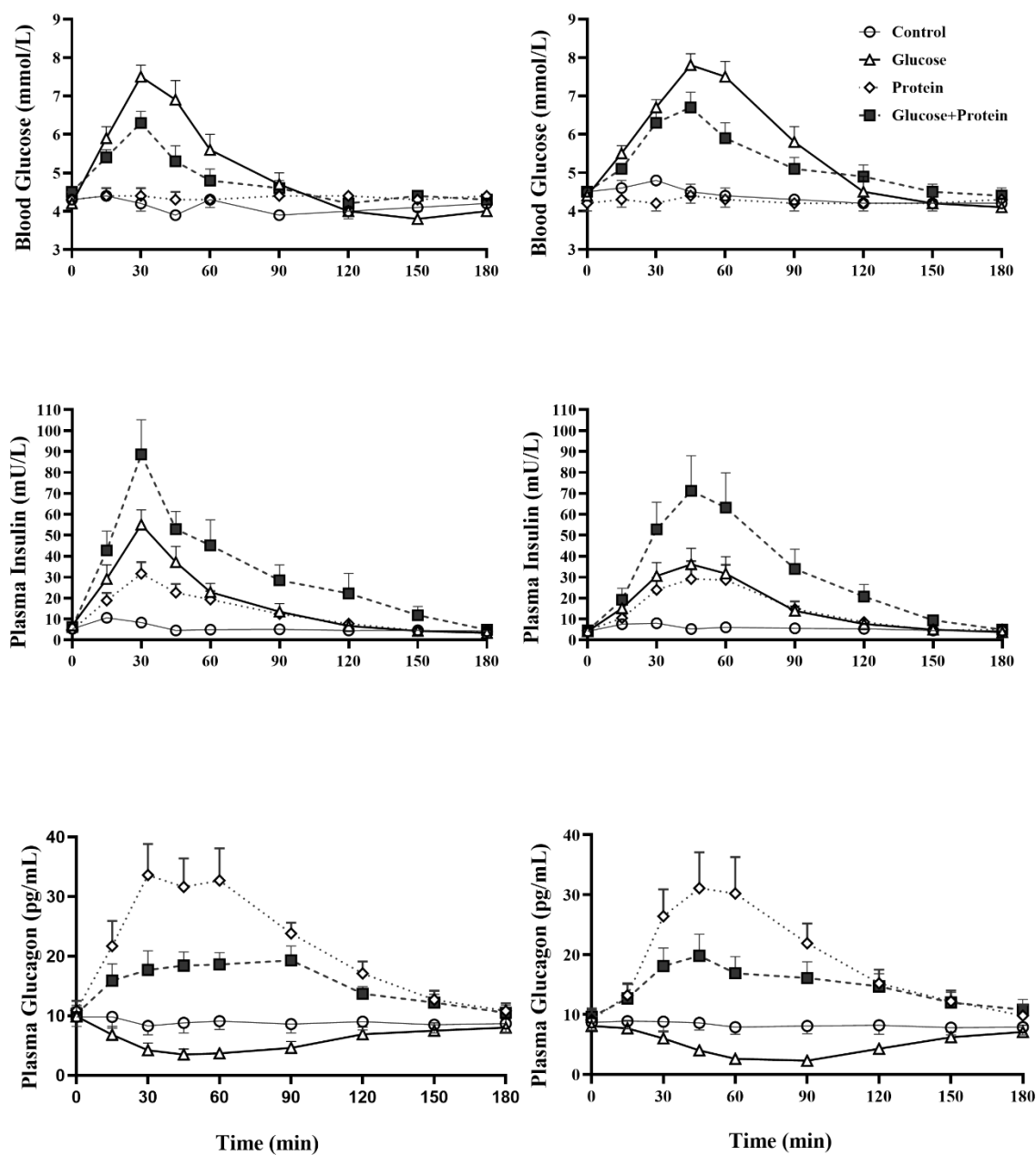


Figure 5.1: Mean (\pm SEM) blood glucose (mmol/L), plasma insulin (mU/L) and glucagon (pg/mL) concentrations following drink ingestion containing (i) flavored water (C, control, \sim 2 kcal), (ii) 30 g whey protein (P), (iii) 30 g glucose (G) or (iv) 30 g whey protein plus 30 g glucose (GP) in younger and older men. Effects of age and drink condition and the interaction effect were determined using a mixed-effect model with baseline concentrations as covariates and post hoc Bonferroni correction.

T = 0 min refers to the point immediately before the drink consumption. Age by drink-condition interaction effects: peak insulin ($P = 0.006$) was higher after G than P in younger but not older men; time to peak glucose ($P = 0.03$) and insulin ($P = 0.038$) was longer after G in older than younger men. Drink-condition effect: $P < 0.001$ Net iAUC glucose GP < G; $P = 0.002$ Net iAUC insulin GP > G; $P < 0.001$ peak glucagon P > GP. Age effect: time to peak glucose ($P = 0.007$) and insulin ($P = 0.001$) occurred later in older than younger men; time to return to baseline glucose ($P = 0.03$) and insulin ($P = 0.01$) occurred later in older than younger men.

5.5.4 Gastric emptying

A number of data points could not be included in the analysis due to poor quality of the samples, particularly in the young men and on the control treatment day, thus the graphs/values obtained were insufficient to determine gastric emptying. Data relating to gastric emptying in these participants were excluded from analysis. After exclusion of these data, gastric emptying data on all three protein and/or glucose drink days were evaluated in 5 younger and 10 older participants who had data from all 3 days.

Gastric emptying was slower after the protein ($\text{AUC}_{0-180/\text{min}}$ gastric retention: $35 \pm 2\% \cdot \text{min}$) than the glucose ($30 \pm 1\% \cdot \text{min}$) drink and slower after the combined glucose and protein drink ($40 \pm 3\% \cdot \text{min}$) than either protein or glucose alone ($P = 0.001$). Gastric emptying was non-significantly slower in older than younger men (T50 GP: 72 ± 13 vs. 53 ± 7 min, P: 47 ± 5 vs. 39 ± 5 min, G: 38 ± 2 vs. 33 ± 2 , $P = 0.29$) (Figure 5.2).

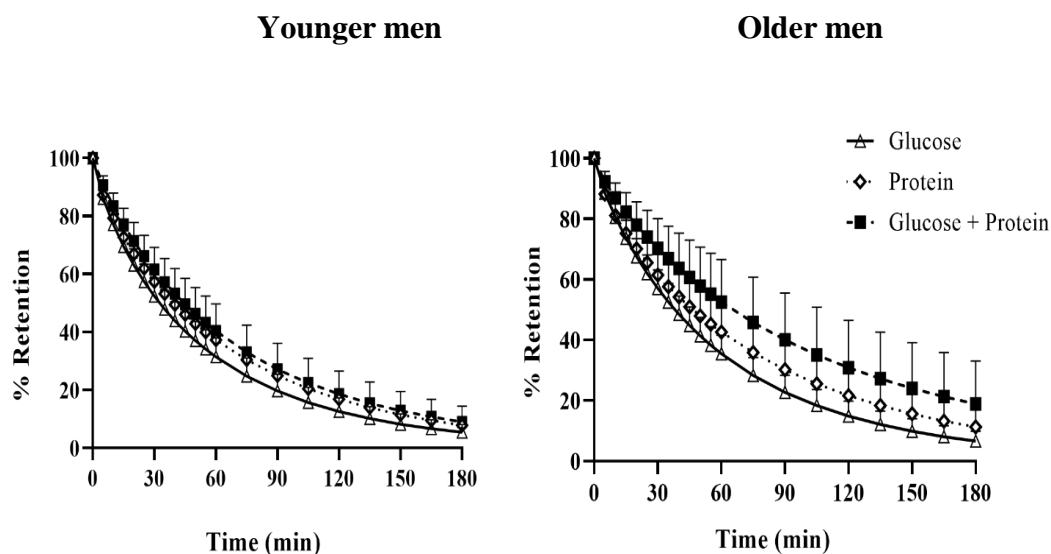


Figure 5.2: Intra-gastric retention (%) following drink ingestion containing (i) 30 g whey protein (P), (ii) 30 g glucose (G) or (iii) 30 g whey protein plus 30 g glucose (GP) in younger ($n = 5$) and older ($n = 10$) men. Effects of age and drink condition and the interaction effect were determined using a mixed-effect model with baseline concentrations as covariates and post hoc Bonferroni correction. $T = 0$ min refers to the point immediately before the drink consumption. Drink-condition effect: $P < 0.001$ gastric emptying of GP was slower than P and both emptied slower than G.

5.5.5 Energy intake

Ad libitum energy intake three hours following preload drink ingestion was comparable between age groups ($P = 0.37$) and drink condition ($P = 0.95$; C, P, G, GP: older: 1008 ± 89 , 1053 ± 106 , 1047 ± 97 , 988 ± 995 kcal; younger: 994 ± 89 , 1003 ± 106 , 960 ± 97 , 939 ± 99 kcal). For GP, the mean energy intake was lower (not significant) compared with all the drinks.

5.5.6 Perceptions of appetite

Mean baseline perceptions of appetite did not differ between age groups (older vs. younger: hunger: 42 ± 9 vs. 49 ± 9 mm, $p = 0.56$; fullness: 6 ± 3 vs. 9 ± 3 mm, $P = 0.55$) or study days ($P > 0.05$).

The protein and protein plus glucose drink suppressed hunger compared with the control during the 3 h following the drinks in the younger ($AUC_{0-180/\text{min}}$ C, P, G, GP: 12 ± 5 , 4 ± 5 , 8 ± 5 , 1 ± 4 mm*min) but not the older (2 ± 5 , 10 ± 5 , 2 ± 5 , 8 ± 4 mm*min) men (age by drink-condition interaction $P = 0.02$, Figure 5.3).

Fullness increased less following the glucose drink when compared with the other drinks in the younger ($AUC_{0-180/\text{min}}$ C, P, G, GP: 4 ± 4 , 5 ± 3 , 0 ± 3 , 7 ± 3 mm*min) but not the older (7 ± 4 , 6 ± 3 , 11 ± 3 , 8 ± 3 mm*min) men (age by drink-condition interaction $P = 0.01$, Figure 5.3).

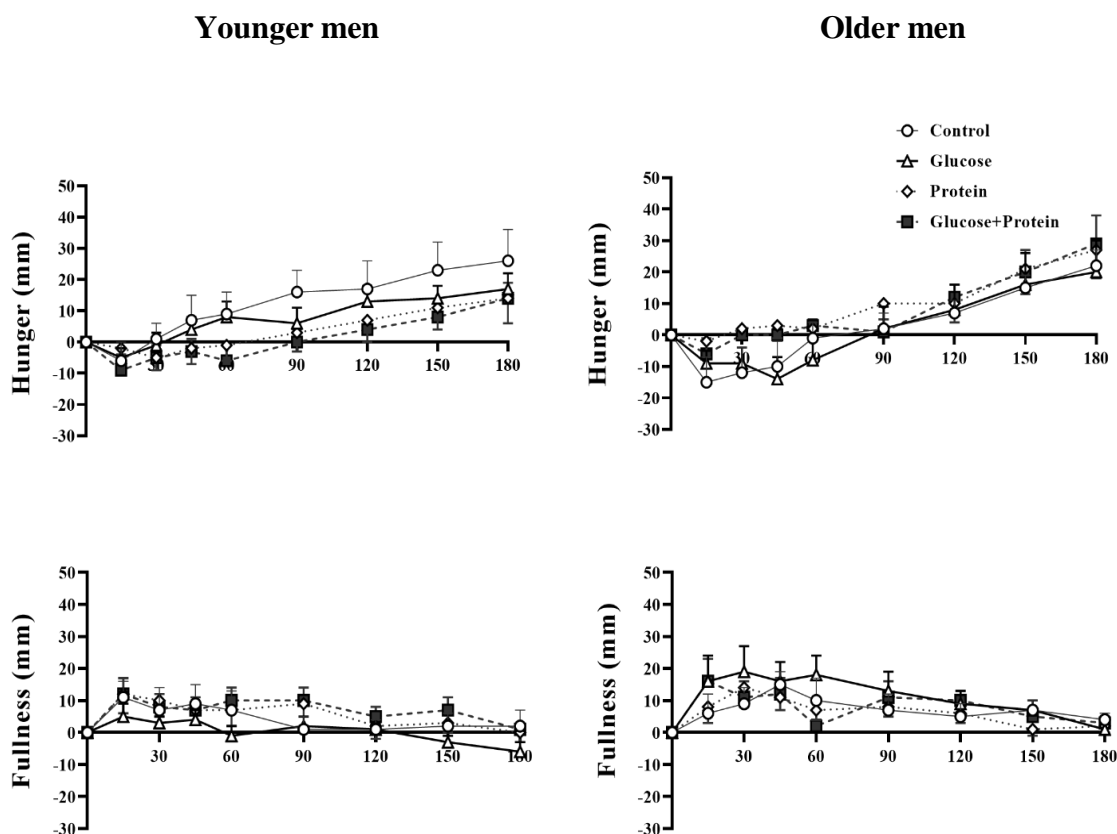


Figure 5.3: Mean \pm SEM visual analogue score (VAS) of hunger (mm) and fullness (mm) following drink ingestion containing (i) flavored water (C, control, \sim 2 kcal), (ii) 30 g whey protein (P), (iii) 30 g glucose (G) or (iv) 30 g whey protein + 30 g glucose (GP) in younger and older men. Effects of age and drink condition and the interaction effect were determined using a mixed-effect model with baseline concentrations as covariates and post hoc Bonferroni correction. T = 0 min refers to the point immediately before the drink consumption. Age by drink-condition interaction effects: $P = 0.02$ hunger was suppressed by P and GP in younger but not older men; $P = 0.01$ fullness was less following G compared with the other drinks in younger but not older men.

5.6 Discussion

The main findings of this study are: (i) the addition of 30 g of whey protein to 30 g of glucose in drink form substantially attenuated the increase in blood glucose concentrations induced by glucose alone; (ii) the magnitude of the whey-induced reduction in blood glucose was not affected by age, with comparable reductions in older to those in younger adult men; (iii) the stimulation of plasma insulin concentrations by whey protein was not reduced by ageing, unlike the insulin response to glucose; (iv) whey protein suppressed hunger less in older than younger men.

These findings are consistent with previous observations, in which the ingestion of various proteins, including oral whey protein in doses ranging from 9 to 55 g, reduced the rise in oral carbohydrate-induced blood glucose concentration in younger adults (44, 86, 138, 350, 351, 353, 354). Those findings are extended by demonstrating that these effects of whey protein are maintained into old age; all older men in this study were 68 years or older with a mean age of 78 years. Previous studies have largely included young and middle-aged adults, with none, as far as we know, studying the effects of whey protein, or other proteins, in any participants much over 60 years. In contrast to this finding, some other responses to dietary protein are affected by ageing. Whey protein ingested alone, in doses of 30–70 g, has a reduced ability to suppress appetite and food intake in older adults compared with young adults (9, 12, 15) and produces greater increases in circulating concentrations of cholecystokinin (CCK) and glucose-dependent insulinotropic polypeptide (GIP) in older adults than younger adults, with comparable increases in circulating insulin (90). Intraduodenal infusions of whey protein, at rates spanning the normal range of gastric emptying, also have different effects on feeding, gut hormone release and glucose metabolism in healthy older adults compared with young adults (9).

The reduced insulin concentrations and slightly higher glucose concentrations observed after ingestion of glucose alone in this study in older adults compared with younger participants are consistent with reported changes in glucose metabolism that accompany human ageing (356-357). There are varying reports as to whether the increase in circulating concentrations of the incretin hormones glucagon-like peptide-1 (GLP-1) and GIP in response to glucose ingestion is affected by ageing (229). Indeed, GIP secretion in response to glucose ingestion may be increased in older people (90, 357, 229), perhaps in compensation for the age-related reduction in the ability of GIP to stimulate insulin secretion (363). Ageing is, however, associated with islet dysfunction and impaired insulin secretion, with disordered pulsatile insulin secretion in both the fasting and hyperglycemic state (357).

Ageing-associated defects in insulin secretion have been demonstrated largely in studies where hyperglycemia has been induced by oral or intravenous glucose; they may not be present in response to protein administration. The results of the present study suggest that they are not. This would be consistent with differing mechanisms of insulin release after glucose and protein ingestion, as also would be the more than additive increases in plasma insulin after combined glucose and protein compared with either alone in this study. Nuttall et al., reported qualitatively similar at-least-additive increases in insulin concentrations after higher, 50 g, doses of protein and glucose. Ingested protein, as with glucose, promotes insulin secretion by stimulating the release of the incretins GLP-1 and GIP. The effects of increasing age on protein-induced increases in circulating incretin concentrations appear similar to those in response to glucose, with equivalent increases in GLP-1 and greater increases in GIP in older and younger participants after whey protein ingestion (90).

Amino acids released by protein digestion also stimulate insulin secretion by direct action on the pancreatic islet cells (142). In contrast to islet responses to glucose, those to amino acids may be less affected, or not at all, by ageing. Some amino acids also stimulate insulin release by acting as substrates in the Krebs cycle to create ATP, which acts directly on the beta cells to increase their insulin secretory response to glucose (142). Support for the existence of this glucose-sensitizing effect of whey protein is provided by greater-than-additive, apparently synergistic, increases in plasma insulin concentrations after combined glucose/whey protein drinks in the present study.

The ingestion of proteins or their component amino acids causes increases in circulating concentrations of both insulin and glucagon, with their opposing effects on blood glucose concentrations and slowing of gastric emptying (90, 346), with resultant slight increases, no effect or slight decreases in blood glucose concentration when ingested on their own, depending on the type of protein or amino acid (346, 351, 352). Circulating glucagon concentrations after the three drinks in this study were not affected by ageing. Glucose alone suppressed plasma glucagon concentrations equally in older and younger participants in this study, consistent with the results of some (142, 364) but not all (365) previous studies. Whey protein alone increased plasma glucagon concentrations markedly and comparably in the two age groups, as previously reported (90). The combination whey protein/glucose drink increased glucagon concentrations equally in both age groups, with the increase approximately half that following whey protein alone. The post whey protein/glucose glucagon increase suggests a stronger stimulatory effect of whey protein on glucagon secretion than the inhibitory effect of glucose, at least with 30 g (120 kcal) of each.

Whey is a dairy protein present in milk along with casein. Relative to casein and most other proteins, whey has higher concentrations of the amino acids leucine, isoleucine, lysine, threonine and tryptophan (93), which are among the most effective amino acids in increasing circulating insulin concentrations and reducing blood glucose when co-ingested with glucose (346, 352), and has greater effects in inhibiting blood glucose increases when co-ingested with carbohydrates (44, 351). In non-elderly people with type 2 diabetes, 17 g whey protein preloads administered alone 15 min before a meal acutely reduced post-prandial glycemia and, when taken twice daily for 12 weeks combined with guar, had sustained effects to slow gastric emptying, reduce postprandial glucose and modestly reduce HbA1C (366). Whey protein supplements are relatively low cost with few, if any, side effects in older people (63). The finding in this study that the increase in circulating insulin concentrations and the reduction in hyperglycemia after whey protein ingestion was unaffected by ageing and persists in those over 70 years and into their 80s, provides encouragement for further studies into the use of whey protein supplements in the prevention and treatment of hyperglycemia in older people.

Whey protein ingestion may have beneficial effects in older people in addition to those on post-prandial glucose concentrations. There is evidence that it can lead to reductions in blood pressure, blood triglyceride levels, inflammation and oxidative stress (128), of which harmfully elevated levels are more prevalent in older adults than younger adults. Another beneficial effect of whey protein in older people is likely to be that on skeletal muscle mass and function. With increasing age there is on average a progressive loss of muscle mass and function, with approximately 5% of muscle mass lost per decade after age 30 years. When excessive, this muscle loss

leads to sarcopenia, with associated substantial increases in morbidity including rates of frailty and mortality (63). Various nutritional measures have been proposed to counteract these adverse effects of ageing, including the use of protein supplements, which has been shown to increase muscle mass and strength and may also reduce morbidity and mortality, with the greatest benefits in the most under-nourished and sarcopenic older people (63).

As with its effects on glucose metabolism, whey protein is a particularly good form of protein to exert this anabolic effect on muscle because of its relatively high content of branched chain amino acids, particularly leucine, which are the most effective amino acids in promoting muscle protein synthesis (249). Some of this effect may be due to the anabolic actions on muscle of the increased insulin secretion following whey protein ingestion. Due to age-related anabolic resistance (63, 270), older adults appear to require 30–35 g/serve of protein to stimulate muscle protein, compared with 20 g or less in younger adults (63), and a minimum of 25–30 g protein intake per meal for older people has been recommended by the PROT-AGE study group (97). This provides the rationale for our use of a 30 g whey protein dose in the present study.

Weight loss in older people, particularly if involuntary, is often associated with adverse outcomes (71) and therefore not necessarily desirable. Protein is the most satiating macronutrient in young adults, and whey protein drinks of 30–70 g suppress appetite ratings and ad libitum energy intake at subsequent meals in healthy young adults. Increasing age is, however, associated with a marked reduction in the satiating effects of whey protein, with no reduction in appetite ratings or ad libitum energy intake by older people at meals 35–510 min after 30 g whey protein drinks by older

people and only minimal effects after 70 g (63). Energy intake at the test meal in this study was not reduced significantly by any of the drinks in either age group, with a slightly greater suppression after the combined whey protein–glucose drink in the younger than the older participants (5.5 vs. 2%). In older people, increasing the dietary content of whey protein may preserve or increase skeletal muscle at the expense of fat gain (128), without reducing appetite and energy intake.

The longer times taken for plasma glucose and insulin concentrations to rise to peak and then return to baseline after the nutrient drinks in the older participants than the younger participants are probably due to a combination of slightly slower gastric emptying and delayed post-gastric effects in the older participants. Gastric emptying of nutrient drinks was non-significantly slower in the older participants than the younger participants, consistent with the results of previous studies, where the slowing of gastric emptying of whey protein with age has been minor when present (63, 229). Post-gastric delays probably played a greater role. Circulating glucose and insulin concentrations increase more slowly in the older men than the younger men, even when the stomach is bypassed by infusing glucose alone (28) directly into the duodenum, and insulin concentrations increase more slowly in the older men than the younger men after intraduodenal whey protein infusion (13). The mechanisms responsible for these post-gastric ageing-related changes are unknown.

Our study has several limitations. The number of participants was relatively small. Nevertheless, the results regarding glucose and insulin results were clearcut and statistically significant. We only studied men, therefore the observation may not apply to women. The results do not necessarily apply for older people ingesting doses of whey protein/glucose other than 30 g, as used in this study, although the

qualitatively similar results observed with a range of protein doses in previous studies of younger participants suggest that they will. While encouraging and suggestive, the ability of co- or pre-ingested whey protein to reduce blood glucose increases in older people after ingestion of carbohydrate in forms other than pure glucose, for example as part of a mixed meal, seems likely but will need to be established (147). Similarly, the effects in older people of non-whey proteins and proteins in non-drink form require further study.

5.7 Conclusions

In conclusion, 30 g of oral whey protein, a dose likely to have beneficial effects on skeletal muscle mass and function in older people, substantially attenuated the rises in blood glucose induced by 30 g of glucose alone in older men without diabetes. This extent of this reduction was not affected by ageing. The plasma insulin and glucagon responses to protein ingestion are preserved in older participants, in contrast to the insulin response to glucose, which is reduced. These findings provide a rationale for future studies of protein supplements/preloads in older people, in whom disorders of glucose metabolism such as type 2 diabetes and of muscle metabolism such as sarcopenia are prevalent and often serious.

**CHAPTER 6: EFFECTS OF CO-INGESTING GLUCOSE AND
WHEY PROTEIN ON BLOOD GLUCOSE, PLASMA INSULIN
AND GLUCAGON CONCENTRATIONS, AND GASTRIC
EMPTYING, IN OLDER MEN WITH AND WITHOUT TYPE 2
DIABETES**

**Oberoi A, Giezenaar C, Ridga RS, Horowitz M, Jones KL, Chapman I,
Soenen S**

Submitted to *BMC Geriatrics*

2022

STATEMENT OF AUTHORSHIP

Title of the paper	Comparative effects of co-ingesting whey protein and glucose alone and combined on blood glucose, plasma insulin and glucagon concentrations in younger and older men.
Publication status	Submitted to BMC Geriatrics
Publication details	Oberoi AO, Giezenaar C, Rigda RS, Horowitz M, Jones KL, Chapman I, Soenen S. <i>Effects of co-ingesting glucose and whey protein on blood glucose, plasma insulin and glucagon concentrations, and gastric emptying, in older men with and without type 2 diabetes.</i>

Candidate	Avneet Oberoi		
Contribution	Conducted the clinical trial, contributed to the overall design of the manuscript, literature review, data analysis and interpretation, drafting and revision of the manuscript.		
Overall percentage	75%		
Certification	This paper reports on original research I conducted during the period of my Higher Degree by Research candidature and is not subject to any obligations or contractual agreements with a third party that would constrain its inclusion in this thesis. I am the primary author of this paper.		
Signature		Date	October 2022

Principal Author***Co-Author Contributions***

By signing the Statement of Authorship, each author certifies that:

- i) the candidate's stated contribution to the publication is accurate (as detailed above);
- ii) permission is granted for the candidate to include the publication in the thesis; and
- iii) the sum of all co-author contributions is equal to 100% less the candidate's

stated contribution.

Name of Co-Author	Caroline Giezenaar		
Contribution	Drafting and revision of the manuscript		
Signature		Date	October 2022
Name of Co-Author	Rachael S Rigda		
Contribution	Data collection, drafting and revision of the manuscript		
Signature		Date	October 2022
Name of Co-Author	Michael Horowitz		
Contribution	Conception and design of the manuscript, drafting and revision of the manuscript.		
Signature		Date	October 2022
Name of Co-Author	Karen L Jones		
Contribution	Conception and design of the manuscript, data interpretation, drafting and revision of the manuscript.		
Signature		Date	October 2022
Name of Co-Author	Ian Chapman		
Contribution	Conception and design of the manuscript, statistical analysis, data interpretation, drafting and revision of the manuscript.		
Signature		Date	October 2022
Name of Co-Author	Stijn Soenen		
Contribution	Conception and design of the study, data interpretation, statistical analysis, drafting of the manuscript and overall responsibility for the study.		

Signature		Date	October 2022
-----------	--	------	--------------

6.1 Abstract

Co-ingestion of dietary protein with, or before, carbohydrate may be a useful strategy to reduce postprandial hyperglycaemia in younger and older men without type 2 diabetes (T2D), but its effect in older people with T2D is not known. Blood glucose, plasma insulin and glucagon concentrations were measured for 180 min following a drink containing either: 30 g glucose (G, 120 kcal), 30 g whey protein (P, 120 kcal), 30 g glucose plus 30 g whey protein (GP, 240 kcal), or control (~2 kcal) in older men with T2D ($n = 10$, 77 ± 1 years; 31 ± 1.7 kg/m²) and without T2D ($n = 10$, 78 ± 2 years; 27 ± 1.4 kg/m²). Mixed model analysis was used. GP vs. G (i) markedly reduced the increase in blood glucose concentrations ($P < 0.001$) and (ii) had a synergistic effect on the increase in insulin concentrations ($P < 0.001$), in men both with and without T2D. Glucose concentrations were higher in men with, when compared to without, T2D whereas insulin and glucagon concentrations were largely unaffected by the presence of T2D. Gastric emptying was faster in men with than without T2D. We conclude that the ability of whey protein to reduce carbohydrate-induced, postprandial hyperglycaemia is retained in older men with, when compared to without, T2D and that whey protein supplementation may be a useful strategy in the prevention and management of T2D in older people.

6.2 Introduction

Type 2 Diabetes mellitus (T2D) becomes more prevalent with increasing age and the impairment of glucose tolerance and hence hyperglycaemia tends to progress over time after diagnosis (340, 367-369). Lifestyle (dietary and exercise) measures are beneficial in improving glycaemic control and metabolic outcomes in T2D, alone and when combined with medications (370-373). Lifestyle measures remain important in older people with T2D (129, 374, 375) and the optimisation of dietary modifications in elderly people with T2D is therefore likely to have substantial benefits in the large number of older people with this condition.

Co-ingestion of protein with carbohydrates increases the insulin secretory response to carbohydrate and so blunts the increase in blood glucose concentrations following ingestion of carbohydrate alone in people both with and without T2D (136, 143, 144, 376, 377). This effect is particularly evident when the ingested protein is whey, a major protein, along with casein, in milk and dairy products (44, 86, 95, 352, 378-385).

We have reported recently the results of a study (135) in older and younger men without T2D, that the inhibitory effect of 30 g whey protein (a dose chosen due to its likely beneficial effects on muscle anabolism in older people) (97) on the increase in blood glucose concentrations after 30 g of glucose ingestion was preserved in older, when compared to younger, men. The stimulation of plasma insulin concentrations by whey protein was not reduced by ageing, unlike the insulin response to oral glucose intake (135). The present study was conducted to determine whether the effects of glucose and whey protein co-ingestion, compared to their ingestion alone,

on blood glucose, plasma insulin and glucagon concentrations, gastric emptying of the drinks, perceptions of appetite and subsequent energy intake present in older men without T2D (135) are also present in those with T2D.

6.3 Materials and methods

6.3.1 Participants

This randomized, double-blind, cross-over study included 10 older men with T2D (age range 69-84 years; 77 ± 1 years; body weight: 89 ± 6 kg; height: 168 ± 3 m; body mass index (BMI): 31.4 ± 1.7 kg/m²) and 10 older men without T2D (age range 68-87 years, 78 ± 2 years; body weight: 82 ± 2 kg; height: 176 ± 2 m; BMI: 27.3 ± 1.4 kg/m²). Body weight and BMI did not differ significantly between groups ($P > 0.05$). Participants were recruited by online advertisement and by flyers placed on notice boards at the University of Adelaide, Australia.

Study exclusion criteria were the following: smokers; vegetarians; intake of any illicit substance; intake of > 20 g alcohol on a daily basis; significant gastrointestinal symptoms or history of gastrointestinal disease including known gastroparesis, or surgery (other than appendectomy or cholecystectomy), proteinuria; any other illness deemed significant by the investigator (including chronic illnesses not explicitly listed above); current use of medications which are likely to affect gastrointestinal function or appetite (e.g. opiates, anticholinergics, levodopa, calcium-channel antagonists, beta blockers, clonidine, nitrates, tricyclic antidepressants, selective serotonin re-uptake inhibitors, phosphodiesterase type 5 inhibitors, sumatriptan, metoclopramide, domperidone, cisapride, tegaserod, or erythromycin); use of non-prescribed medications (including vitamins and herbal supplements) which may

affect appetite, body weight, gastrointestinal function or energy metabolism (e.g. green tea extracts, Astragalus, St John's Wort etc.); individuals who are found to be unable to comprehend the study protocol.

In participants with T2D further exclusion criteria were treatment with insulin or other diabetes medications apart from metformin and/or a dipeptidyl 4 (DPP4) inhibitor.

Inclusion criteria included being weight stable (< 5% fluctuation in their body weight) at study entry, as assessed by self-reported weight in the preceding 3 months, willingness to maintain usual physical activity level throughout the study and for participants with T2D having a HbA1c between 6% and 8.5% at the time of screening. The HbA1c of participants with T2D is presented in Table 6.1. Mean HbA1c of participants without T2D was 5.3%.

Table 6.1: Participant characteristics of older men with T2d (n = 10)

Age (years)	Time since Diabetes Diagnosis (years)	Screening HbA1c (%)	Diabetes medication
77	22	6.1	-
84	17	6.4	-
69	13	7.8	Metformin
78	22	7	Metformin
75	9	8.1	Metformin
82	6	7.9	Metformin/ Sitagliptin

73	16	7.3	Metformin/Empagliflozin
78	8	7.2	-
76	2	6.2	-
80	10	6.1	Metformin

The Royal Adelaide Hospital Human Research Ethics Committee approved the protocol which was conducted in accordance with the Declaration of Helsinki. The study was registered with the Australian New Zealand Clinical Trial Registry (www.anzctr.org.au, registration number ACTRN12619000420145). All participants provided written informed consent prior to their study inclusion.

6.3.2 Protocol

The protocol was identical to that of our previous study that compared younger and older men without T2D, and the results in the older men without T2D have been published previously (135). The study had a randomized (using the method of randomly permuted blocks; www.randomization.com), double blind study design where each participant was studied on four occasions, separated by ~7–10 days, and received a drink of either 30 g glucose (G, 120 kcal) (glucose monohydrate- Sigma-Aldrich, Missouri, USA), 30 g whey protein (P, 120 kcal) (Bulk Nutrients, Tasmania, Australia), 30 g glucose plus 30 g whey protein (GP, 240 kcal), or lime flavoured water (control, C; ~2 kcal) to determine the effects on blood glucose, plasma insulin and glucagon concentrations, gastric emptying, perceptions of appetite and energy intake at a buffet-style meal.

The study drinks were equivolaemic (~250 mL) and prepared by a research assistant

who was not involved in analysis of the study results. The drinks were stirred continuously at low speed on a stirring plate to ensure that all ingredients were dissolved evenly throughout and served in a covered cup to achieve blinding. Both the investigators conducting the study and the participants were blinded to the drink composition. The drinks were flavoured with varying amounts of distilled water, sodium chloride, light lime cordial (Bickford's Australia Pty Ltd, South Australia) and 100 mg [^{13}C] sodium acetate to match for taste.

Participants were instructed (i) to consume the same meal on the night before each study day at around 1900h, (ii) to fast overnight from solids and liquids and, (iii) to refrain from strenuous physical activity and alcohol for 24 hours prior to their attendance at the laboratory at the Clinical Research Facility, Adelaide Health and Medical Sciences Building, the University of Adelaide.

Upon arrival at the facility at 0830am, participants were seated in a chair. A cannula was inserted in an antecubital vein for blood sampling. A heated pad was used so that the blood samples were arterialised. Participants were instructed to consume the test drink within 2 minutes.

6.3.3 Measurements

6.3.3.1 Blood glucose, plasma insulin and plasma glucagon concentrations

Blood samples were collected into ice-chilled, EDTA-coated tubes for the measurement of blood glucose, plasma insulin and glucagon concentrations at $t = 0$ (baseline, after overnight fasting immediately before consumption of the study drink), 15, 30, 45, 60, 90, 120, 150 and 180 min. No inhibitors were added (362).

Plasma was obtained by centrifugation for 15 min, at 3200 rpm at 4°C, and samples were stored at -80°C for further analysis of insulin and glucagon concentrations. Blood glucose concentrations (mmol/L) were measured using a glucose analyser by the glucose oxidase method (YSI 2900 Stat Plus, Yellow Springs Instruments, Yellow Springs, Ohio, USA). Intra- and inter-assay coefficient of variations (CVs) were $\leq 2\%$. Total plasma insulin concentrations (milliunits per liter) were measured by enzyme-linked immunosorbent assay (ELISA) immunoassay (10-1113; Merckodia, Uppsala, Sweden). The sensitivity of the assay was 1.0 mU/L. Intra- and inter-assay CVs were 2.9% and 11.6%. Plasma glucagon concentrations were measured by ELISA immunoassay (10-1271-01, Merckodia, Uppsala, Sweden). The lower limit of quantification (LLOQ) was 1.5 pmol/L, the detection limit was 0.75 pmol/L. Intra- and inter-assay CVs were 9.3% and 7.5%.

6.3.3.2 Gastric emptying

Gastric emptying (% intragastric retention at $t = 0, 5, 10, 15, 20, 25, 30, 35, 40, 45, 50, 55, 60, 75, 90, 105, 120, 135, 150, 165$ and 180 min) of the drink was measured with ^{13}C -sodium acetate breath test (359). Breath samples consisted of the participant's end-expiratory breath, collected into a 100mL foil bag. The quantity of $^{13}\text{CO}_2$ (disintegrations per min) was quantified using a non-dispersive infrared spectrometer (FANci2, Fischer ANalysen Instrumente, Germany) (360). Intragastric retention was calculated using the Wagner-Nelson method (361) which has been shown to be of comparable accuracy to scintigraphy in the measurement of gastric emptying of liquid food ingestion (242, 362).

6.3.3.3 Energy intake

At 180 min, each participant was presented with a standardised, cold, buffet-style meal, in excess of what they were expected to consume and were allowed to eat freely for 30 min (180–210 min) until comfortably full in a room by themselves to limit external distractions. The total energy content of the buffet-style meal was 2457 kcal; 19% protein, 50% carbohydrates, 31% fat. The buffet-style meal consisted of palatable food items including sliced bread, cheese, ham chicken, lettuce, tomato, cucumber, banana, apple, yogurt, custard, margarine, mayonnaise, iced coffee, orange juice, fruit salad, and water (16). Food consumed (g) was quantified by weighing the food before and after consumption. Energy intake was calculated as absolute (kcal) at the buffet meal using commercially available software (Foodworks version 8; Xyris Software Pty Ltd.) (15).

6.3.3.4 Perception of hunger and fullness

Perceptions of hunger and fullness were rated using a 100 mm visual analog scale questionnaire (t = 0, 15, 30, 45, 60, 90, 120, 150 and 180 min).

6.4 Data and statistical analysis

Statistical analyses were performed using SPSS software (version 25; IBM, Armonk, NY, USA). Sample size was based on statistical power functions of between-groups contrasts of older men with and without T2D with overall $P = 0.05$, statistical power of 0.8 and anticipated drop-out rate of ~10%, and significance levels adjusted to account for the 4 comparisons (control, whey protein, glucose, whey protein plus glucose). Calculations were performed for the primary outcome of area under the

curve (AUC) of blood glucose concentrations, assuming a within- participant SD of 0.5 mmol/l, and a between-participants SD of 1.4 mmol/l (14, 15, 90) to detect a difference between groups of 1.5 mmol/L, and between treatments of 0.4 mmol/L.

For measures of blood glucose, plasma insulin and glucagon concentrations, net incremental area under the curve (Net iAUC), peak, time to peak and time to return to baseline was calculated. Net iAUC was calculated from baseline using the trapezoidal rule, and then divided by time (min) to present a weighted average value over the time interval.

Mixed-effects model was used to determine the main effects of age and drink-condition, and their interaction effect, on blood glucose, plasma insulin and glucagon concentrations with the drink-condition as the within- participant factor and age as the between- participant factor, including baseline values at each treatment visit as a covariate. Post hoc comparisons, which were adjusted for multiple comparisons with Bonferroni correction, were performed when there were significant drink-condition or interaction effects. A repeated measures ANOVA (G and GP for glucose; G, P and GP for insulin and glucagon) was used to determine peak concentrations, time to peak and time to return to baseline for blood glucose, plasma insulin and glucagon concentrations in those drink conditions associated with changes from baseline. When the time to return to baseline was between 2 blood sampling time points, an interpolated value was estimated assuming a linear relationship between the 2 time points. If a blood glucose or plasma insulin or glucagon concentration did not return to baseline by 180 minutes the time to return to baseline was calculated using a linear extrapolation from the values at 150-180 mins. We hypothesised that insulin concentrations following both G and P drinks

would increase, and to determine if the effect on insulin of the combination drink is synergistic or additive, we compared the effect of combined whey protein and glucose (GP drink) on the rise in plasma insulin concentrations to that of the sum of the effects of glucose and whey protein alone (G drink + P drink) on insulin concentrations, using a paired t test. The inhibition of the increase in blood glucose concentrations following glucose ingestion by whey protein, when compared to glucose ingestion alone was calculated as the difference between Net iAUC of G and GP as a percentage of Net iAUC of G. The increase in plasma insulin concentrations following GP when compared to G ingestion alone was calculated as the difference between Net iAUC of GP and G as a percentage of Net iAUC of G. The increase in plasma glucagon concentrations following P when compared to G ingestion alone was calculated as the difference between Net iAUC of P and G as a percentage of Net iAUC of G. Statistical significance was accepted at $P < 0.05$. Data are presented as mean values \pm SEM.

6.5 Results

The study protocol was well tolerated by all participants and no adverse effects were reported.

6.5.1 Blood glucose concentrations

6.5.1.1 Interaction effects

Peak blood glucose concentrations occurred earlier after GP than G in older men without T2D ($P = 0.008$), and occurred later in men with T2D than without T2D following ingestion of the GP drink ($P = 0.005$). The group (diabetes status) by drink

interaction effect was non-significant for all other outcomes (Table 6.2).

6.5.1.2 Drink-condition effects

Blood glucose concentrations increased after overnight fasting following glucose ingestion, alone (G, $P < 0.001$) and when combined with whey protein (GP) ($P < 0.001$) compared to control, to peak mean concentrations of 6.9 - 12.5 mmol/L (Table 6.2). Glucose concentrations did not change from baseline following the C and P drinks ($P > 0.05$).

Co-ingestion of glucose with whey protein, when compared to glucose ingestion alone, reduced the peak (GP vs. G: 8.7 ± 0.3 vs. 10.3 ± 0.5 mmol/L, $P < 0.001$) and Net iAUC_{0-60min} (1.9 ± 0.2 vs 2.6 ± 0.2 mmol/L, $P < 0.001$) glucose concentrations, without significant effect of diabetes status on this inhibitory effect ($P > 0.05$).

The inhibition of the increase in blood glucose concentrations following glucose ingestion by whey protein, when compared to glucose ingestion alone was particularly evident during the first two hours following the drink ingestion; 30 ± 7 % in the first hour and 31 ± 9 % in the second hour (Figure 6.1).

6.5.1.3 Group effects

Overnight fasting ($P < 0.001$), peak ($P < 0.001$), Net iAUC_{0-60min} ($P = 0.005$) and Net iAUC_{0-180min} ($P < 0.001$) glucose concentrations were higher in men with T2D than without T2D (both, Table 6.2). Inhibition of the increase in blood glucose concentrations following glucose ingestion by whey protein, when compared to

glucose ingestion alone (see above), did not differ significantly between groups; older men with vs. without T2D 20 ± 11 % vs. 41 ± 6 % in the first hour ($P = 0.12$) and 15 ± 13 % vs. 51 ± 9 % in the second hour ($P = 0.14$).

Table 6.2: Blood glucose, plasma insulin and glucagon concentrations following control drink (C), 30g whey protein drink (P), 30g glucose drink (G) and 30g glucose plus 30g whey protein drink (GP) ingestion

	Older men without T2D				Older men with T2D				P value		
	C	P	G	GP	C	P	G	GP	Group	Drink Condition	Interaction
Blood glucose (mmol/L)											
Fasted	4.5 ± 0.4	4.2 ± 0.4	4.4 ± 0.5	4.5 ± 0.3	6.6 ± 0.4	6.9 ± 0.4	7.1 ± 0.4	6.6 ± 0.3	<0.001	0.52	0.33
Peak	4.9 ± 0.4	4.7 ± 0.1	8.1 ± 0.7	6.9 ± 0.5	7.2 ± 0.4	7.6 ± 0.5	12.5 ± 0.7	10.5 ± 0.5	<0.001	<0.001	0.21
Time to peak (min)	27 ± 7	38 ± 7	51 ± 4 ¹	36 ± 3 ^{1,2}	29 ± 7	32 ± 7	51 ± 4	51 ± 3 ²	0.09	0.049	0.049
Return to baseline (min)	60 ± 22	125 ± 23	132 ± 14	153 ± 49	81 ± 31	44 ± 12	186 ± 14	221 ± 49	0.22	0.69	0.83
Net iAUC _{0-60min}	0.1 ± 0.1	0.1 ± 0.1	2.1 ± 0.2	1.3 ± 0.2	0.0 ± 0.1	0.2 ± 0.1	3.1 ± 0.3	2.4 ± 0.3	0.005	<0.001	0.87

Net iAUC _{0-180min}	0.1 ± 0.1	0.07±0.08	1.1 ± 0.2	0.7 ± 0.2	-0.4 ±0.2	0.0 ± 0.2	2.5 ± 0.3	1.9 ± 0.2	<0.001	0.001	0.65
Plasma insulin (mU/L)											
Fasted	4.1 ± 1.7	4.5 ± 1.4	4.1 ± 1.4	4.5 ± 1.9	7.9 ± 1.7	8.2 ± 1.4	8.2 ± 1.4	8.5 ± 1.9	0.08	0.52	0.95
Peak	9.7 ± 2.9	31.7 ± 7.5	36.5 ± 7.9	72.7 ±14.5	15.1±2.9	41.2 ± 4.5	36.4 ±7.9	81.1 ±14.5	0.67	<0.001	0.07
Time to peak (min)	29 ± 7	48 ± 3	46 ± 6	42 ± 3 ³	39 ± 5	48 ± 3	49 ± 6	61 ± 3 ³	0.12	0.40	0.015
Return to baseline (min)	109 ± 18	149 ± 9	163 ± 12	166 ± 21	95 ± 18	163 ± 9	204 ± 12	222 ± 21	0.009	0.013	0.26
Net iAUC _{0-60min}	2.3 ± 0.5	15.5 ± 3.8	20.9 ± 4.4	39.8 ± 9.3	3.2 ± 0.9	17.5 ± 4.2	16.4 ±4.7	33.6 ± 8.9	0.76	<0.001	0.14
Net iAUC _{0-180min}	1.5 ± 0.6	9.6 ± 1.9	11.6 ± 2.4	26.6 ± 6.2	0.9 ± 0.6	11.9 ± 2.6	12.4± 2.9	31.2 ± 6.9	0.68	<0.001	0.68
Plasma glucagon											
(pg/mL)											
Fasted	8.7 ± 1.3	9.1 ± 1.5	8.1 ± 1.2	9.6 ± 1.3	9.6 ± 1.3	9.9 ± 1.5	9.2 ± 1.2	8.8 ± 1.3	0.78	0.56	0.70
Peak/Nadir	10 ± 1.3	33.1 ± 4.8	9.1 ± 1.4	22.1 ± 2.9	11.7±1.5	35.9 ± 4.8	11.2 ±1.4	25.1 ± 2.9	0.53	<0.001	0.94
Time to peak (min)	65 ± 20	51 ± 10	56 ± 21	48 ± 7	57 ± 20	69 ± 10	34.5 ± 21	37.5 ± 7	0.76	0.11	0.17
Return to baseline (min)	52 ± 14	391 ± 153	133 ± 29	277 ± 51	72 ± 14	185 ± 153	98 ± 34	155 ± 51	0.12	0.042	0.47

Net iAUC _{0-60min}	-0.1 ± 0.2	13.5 ± 2.6	-2.4 ± 0.7	6.3 ± 1.5	0.4 ± 0.2	16.8 ± 2.1	-0.2 ± 0.6	8.8 ± 1.5	0.07	<0.001	0.12
Net iAUC _{0-180min}	-0.5 ± 0.3	9.9 ± 1.3	-3.2 ± 0.8	5.1 ± 1.3	-0.1 ± 0.3	12.1 ± 1.3	-1.7 ± 0.6	7.3 ± 1.4	0.09	<0.001	0.20

Blood glucose (mmol/L) and plasma insulin (mU/L) and glucagon (pg/mL) concentrations fasted (baseline), peak, time to peak (min), return to baseline (min), Net iAUC_{0-60min} (change from baseline area under the curve during the first hour), Net iAUC_{0-180min} (change from baseline area under the curve during the three hours) following drink ingestion containing (i) flavored water (C, control, ~2 kcal), (ii) 30 g whey protein (P), (iii) 30 g glucose (G), or (iv) 30 g glucose plus 30 g whey protein (GP) in older men with and without T2D. Effects of group and drink-condition and interaction effects were determined using a mixed-effect model using G and GP for glucose and P, G and GP for insulin and glucagon. Group x drink-condition interaction: $P = 0.049$ ¹Peak glucose concentrations occurred earlier after GP than G in older men without T2D; ^{2,3}Peak glucose and insulin concentrations occurred later in men with T2D than without T2D. Drink-condition effects: $P < 0.001$ G and GP vs. C higher glucose and G, P and GP vs. C higher insulin concentrations; $P < 0.05$ GP vs. G lower glucose and GP vs. G and P higher insulin concentrations; $P < 0.05$ G vs. C lower GP vs. C and P vs. C and GP higher glucagon concentrations. Group effects: $P < 0.05$ older men with T2D vs. without T2D higher glucose concentrations.

6.5.2 Plasma insulin

6.5.2.1 Interaction effects

Peak blood insulin concentrations occurred later in men with T2D than without T2D following ingestion of the GP drink ($P = 0.015$). The group (diabetes status) by drink interaction effect was non-significant for all other outcomes (Table 6.2).

6.5.2.2 Drink-condition effects

Plasma insulin concentrations (peak, Net iAUC) increased after overnight fasting following G, P and GP, compared to control ($P < 0.001$, Table 6.2). Insulin concentrations (peak and Net iAUC) increased more ($P < 0.001$), and time to return to baseline was greater ($P = 0.013$), following GP than both G and P, with no difference between the increase after G and P over 3 hours ($P = 0.80$). GP increased insulin concentrations more than the sum of the increases after G and P administered separately (Net iAUC_{0-180/min} GP vs. G plus P: 28.9 ± 4.8 vs. 22.7 ± 3.8 mU/L*min, $P = 0.002$).

6.5.2.3 Group effects

Baseline insulin concentrations were non-significantly higher in older men with T2D than without T2D ($P = 0.08$). Older men with T2D vs. without T2D showed non-significantly higher peak insulin concentrations after the P and GP drinks (Table 6.2). Insulin concentration time taken to return to baseline after drink ingestion was longer in men with T2D than without T2D ($P = 0.009$).

The higher increase in insulin concentrations over 3 hours by GP when compared to G plus P (see above) was comparable ($P = 0.68$ in the men with T2D ($23 \pm 1\%$) and without T2D ($22 \pm 1\%$)).

6.5.3 Plasma glucagon

6.5.3.1 Interaction effects

The group (diabetes status) by drink interaction effect was non-significant for plasma glucagon concentrations (Table 6.2).

6.5.3.2 Drink-condition effects

When compared to control, plasma glucagon concentrations decreased after overnight fasting following G, increased following GP and increased significantly more following P than G and GP ($P < 0.001$, Table 6.2, Figure 6.1). P increased glucagon concentrations by $26 \pm 34\%$ more than did G over 3 hours ($P < 0.001$); $14 \pm 22\%$ more in the first hour ($P < 0.001$), $29 \pm 41\%$ more in the second hour ($P < 0.001$), with a comparable increase in the third hour ($P = 0.28$). Glucagon concentrations time taken to return to baseline was longer after P than GP and G ($P = 0.042$).

6.5.3.3 Group effects

Glucagon concentrations were not affected by the presence of diabetes (Table 6.2).

Older men without T2D

Older men with T2D

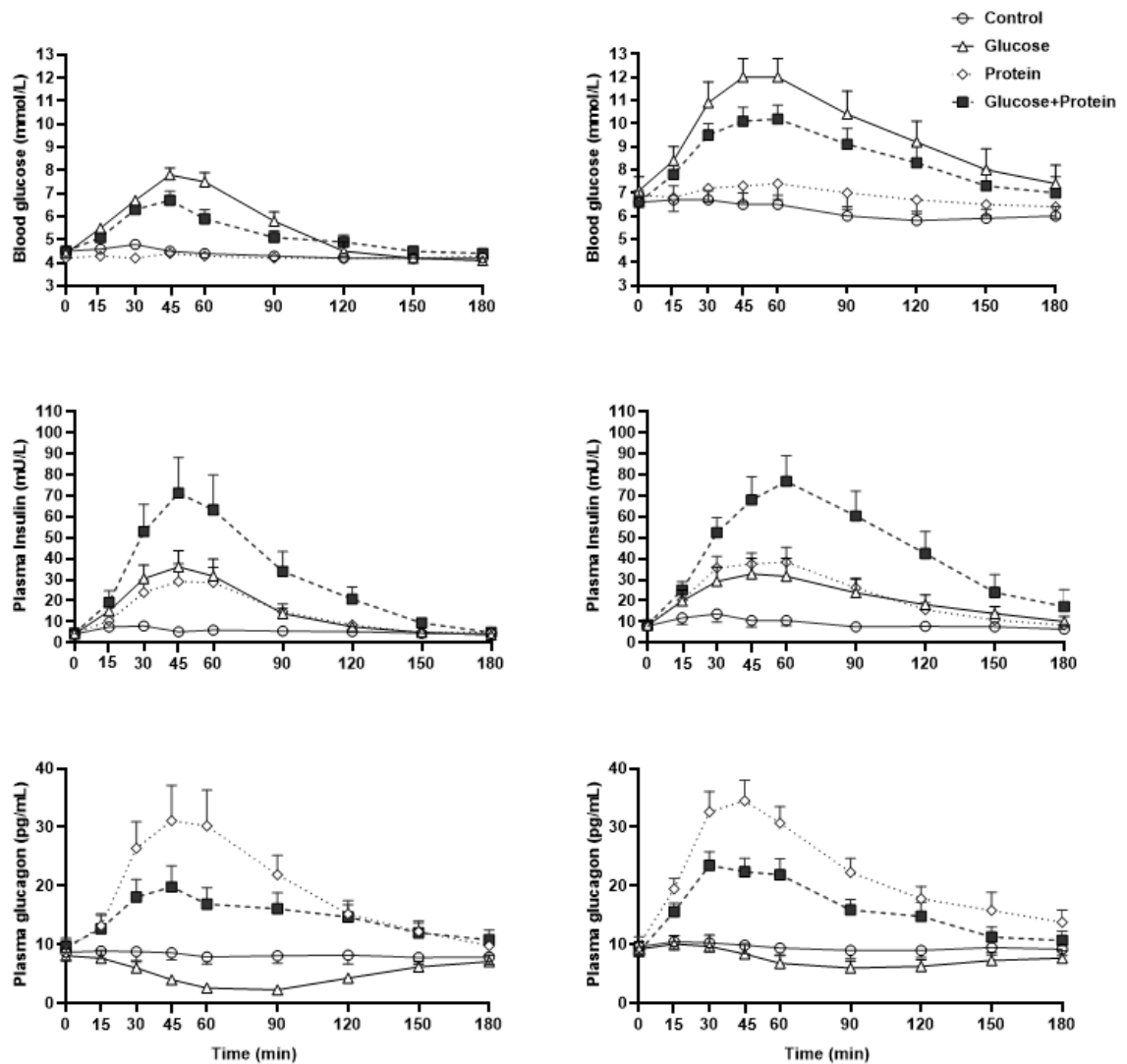


Figure 6.1: Mean (\pm SEM) blood glucose (mmol/L), plasma insulin (mU/L) and glucagon (pg/mL) concentrations following drink ingestion containing (i) flavored water (C, control, \sim 2 kcal), (ii) 30 g glucose (G), (iii) 30 g whey protein (P), or (iv) 30 g glucose plus 30 g whey protein (GP) in men with and without T2D. Effects of group and drink-condition and interaction effects were determined using a mixed-effect model using G and GP for glucose and P, G and GP for insulin and glucagon. Group x drink-condition interaction: $P = 0.049$ Peak glucose concentrations occurred earlier after GP than G in older men without T2D; Peak glucose and insulin concentrations occurred later in men with T2D than without T2D. Drink-condition

effects: $P < 0.001$ G and GP vs. C higher peak and Net iAUC glucose and G, P and GP vs. C higher insulin concentrations; $P < 0.05$ GP vs. G lower glucose and GP vs. G and P higher peak and Net iAUC insulin concentrations; $P < 0.05$ G vs. C lower GP vs. C and P vs. C and GP higher peak and Net iAUC glucagon concentrations. Group effects: $P < 0.05$ older men with T2D vs. without T2D higher peak and Net iAUC glucose concentrations.

6.5.4 Gastric emptying

6.5.4.1 Interaction effects

The group (diabetes status) by drink interaction effect was non-significant for $AUC_{0-180/\text{min}}$ gastric retention ($P = 0.23$).

6.5.4.2 Drink-condition effects

Gastric emptying was slower ($P = 0.007$) after P ($AUC_{0-180/\text{min}}$ gastric retention: $34 \pm 2\% \cdot \text{min}$) than G ($30 \pm 1\% \cdot \text{min}$) and slower after GP ($41 \pm 3\% \cdot \text{min}$) than either P or G (both $P < 0.001$) (Figure 6.2).

6.5.4.3 Group effects

Gastric emptying was faster ($P = 0.04$) in older men with T2D ($31 \pm 2\% \cdot \text{min}$) than without T2D ($38 \pm 2\% \cdot \text{min}$).

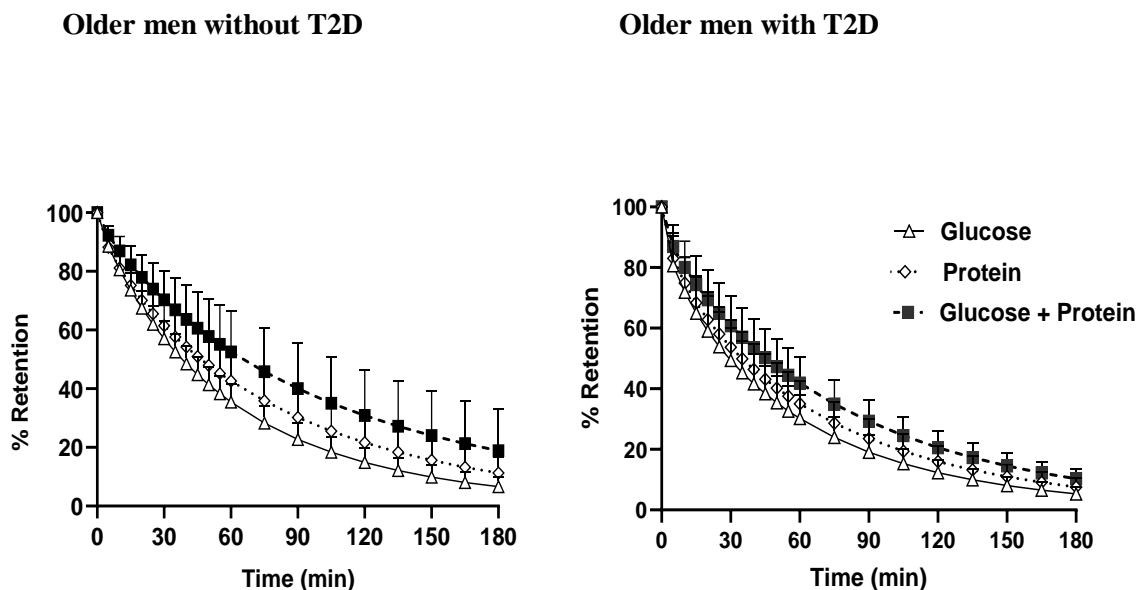


Figure 6.2: Intra-gastric retention (%) following drink ingestion containing (i) 30 g glucose (G), (ii) 30 g whey protein (P), or (iii) 30 g glucose plus 30g whey protein (GP) in men with ($n = 10$) and without T2D ($n = 10$). Effect of group and drink-condition and the interaction effect were determined using a mixed-effect model and post hoc Bonferroni correction. T = 0 min refers to the point immediately before the drink consumption. Drink condition effect: $P < 0.001$ GP vs. P and G, P vs. G slower gastric emptying.

6.5.5 Energy intake

Ad libitum energy intake three hours following drink ingestion was comparable between groups ($P = 0.76$) and drink-condition ($P = 0.88$); older men with T2D: C: 1015 ± 95 kcal, G: 997 ± 99 kcal, P: 997 ± 95 kcal, GP: 965 ± 115 kcal; older men without T2D: C: 1008 ± 89 kcal, P: 1053 ± 106 kcal, G: 1047 ± 97 kcal, GP: 988 ± 995 kcal.

6.5.6 Perception of hunger and fullness

Mean baseline perceptions of appetite did not differ significantly between study days ($P > 0.05$) or groups; older men without T2D vs. with T2D: hunger: 42 ± 9 mm vs. 26 ± 9 mm, $P = 0.21$; fullness: 6 ± 3 mm vs. 8 ± 3 mm, $P = 0.66$.

Hunger decreased immediately following all drinks in older men without T2D to steadily increase during the remainder of the study duration (Net iAUC_{0-180/min} C: 2 ± 5 , P: 10 ± 5 , G: 2 ± 5 , GP: 8 ± 4 mm*min, repeated measures time effects $P < 0.001$), whereas hunger did not decrease following ingestion of the drinks and increased steadily during the three hours in older men with T2D (Net iAUC_{0-180/min} C: 5 ± 3 , P: 7 ± 4 , G: 7 ± 5 , GP: 10 ± 5 mm*min, repeated measures time effects $P < 0.05$, Figure 6.3). Fullness increased immediately following G, P and GP in older men without T2D to decrease back to baseline during the remainder of the study duration (Net iAUC_{0-180/min} C: 7 ± 3 , P: 6 ± 3 , G: 11 ± 5 , GP: 8 ± 3 mm*min, repeated measures time effects $P < 0.05$), whereas fullness did not change significantly over time following drinks ingestion in older men with T2D (Net iAUC_{0-180/min} C: 4 ± 2 , P: 8 ± 3 , G: 5 ± 4 , GP: 9 ± 4 mm*min, repeated measures time effects $P > 0.05$).

Older men without T2D

Older men with T2D

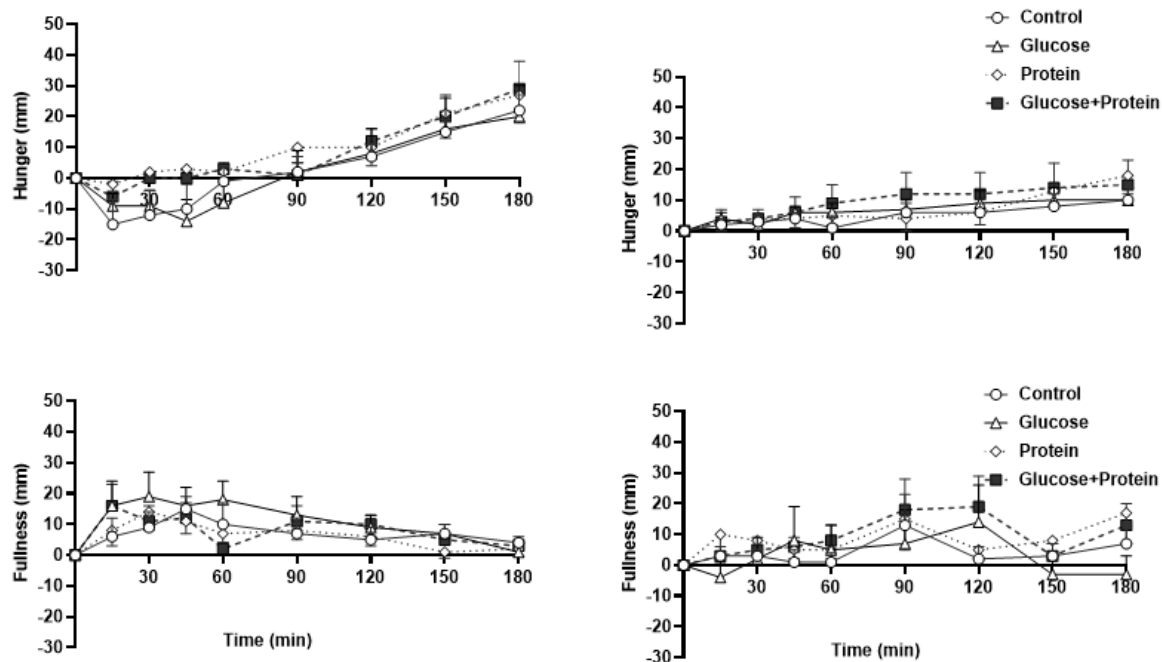


Figure 6.3: Mean \pm SEM visual analogue score (VAS) of hunger (mm) and fullness (mm) following drink ingestion containing (i) flavored water (C, control, \sim 2 kcal), (ii) 30 g glucose (G), (iii) 30 g whey protein (P), or (iv) 30 g glucose plus 30 g whey protein (GP) in men with and without T2D. Repeated measures time effects: Hunger decreased immediately following all drinks in older men without T2D to steadily increase during the remainder of the study duration ($P < 0.001$), whereas hunger did not decrease following ingestion of the drinks and increased steadily during the three hours in older men with T2D ($P < 0.05$). Fullness increased immediately following G, P and GP in older men without T2D to decrease back to baseline during the remainder of the study duration ($P < 0.05$), whereas fullness did not change significantly over time following drinks ingestion in older men with T2D ($P > 0.05$).

6.6 Discussion

The main findings of this study which determined postprandial effects of glucose, whey protein and their combined ingestion are: (i) fasted and postprandial blood glucose concentrations were higher in older men with than without T2D; (ii) the addition of 30 g of whey protein to 30 g of glucose in drink form substantially attenuated the increase in blood glucose concentrations after glucose alone, to a similar extent in older men both with and without T2D; (iii) the stimulation of plasma insulin concentrations by whey protein was not affected by T2D, (iv) time to peak glucose and insulin concentrations occurred later in older men with than without T2D; (v) plasma glucagon concentrations increased after protein and decreased after glucose in older men both with and without T2D; (vi) gastric emptying of the caloric drinks was faster in men with than without T2D.

These results extend and are consistent with those of previous studies. The ability of combined orally ingested carbohydrate and protein ingestion to stimulate insulin secretion more than carbohydrate alone was first suggested in 1960 (146, 386), and later confirmed in people without (69) and with T2D (68, 348, 349). Nuttall *et al.* studied younger participants with T2D than in our study, and similarly reported at-least-additive increases in circulating insulin concentrations, albeit after higher (50 g) doses of protein and glucose than we used and with the protein in the form of a beef hamburger (68). Other studies have reported a 3-4 fold rise in insulin with a protein and carbohydrate meal as compared to glucose alone in people with T2D with a substantial reduction in postprandial glycaemia (143) similar to our study. They have, however, studied younger participants and administered the protein and carbohydrate using more complex study protocols e.g., as protein hydrolysate with

leucine, and phenylalanine together with continuous glucose infusions (376). Our study has investigated the effects of a simpler oral administration of whey protein and glucose in drink form on older men with and without T2D.

We have reported previously that co-ingesting 30 g whey protein with 30 g glucose substantially and significantly attenuated the increase in blood glucose concentration following 30 g glucose alone, to a similar degree in younger and older men without T2D (135). We now report that whey protein also inhibits glucose-induced blood glucose rises in older men with T2D. This is of possible therapeutic significance given the large numbers of older people with T2D.

Of note, although their diabetes was often longstanding (mean duration of known diabetes 12.5 years, range 2-22 years), none of the older men with diabetes in this study were receiving injectable treatment for their T2D i.e., insulin or a GLP-1 analog. It cannot be assumed that the glucose lowering effects of co-administered whey protein will be present in such people with T2D. Indeed this is probably unlikely, particularly in insulin-deficient people with T2D receiving insulin treatment, given that the marked whey-induced increases in plasma insulin concentrations suggest that this is the main mechanism by which whey protein inhibits glucose-induced hyperglycaemia. T2D is characterised by progressive beta cell failure with a substantial proportion of people with this condition eventually requiring insulin treatment (387). The natural history and time course of this failure is, however, variable, as illustrated by the participants in this study; their mean HBA1C was 7%, despite none being on insulin treatment a considerable time after diagnosis. We suspect that the blood glucose lowering effects of whey protein and other proteins will depend on the degree to which insulin secretory capacity and

action is retained, rather than the duration of T2D. There is, however, the possibility that the insulin secretory response to protein will be retained longer than that to carbohydrate in people with T2D, as we have found with the effects of ageing in people without T2D (135). It will be interesting to determine the effect of whey protein on glucose concentrations in older people with T2D on injectable hypoglycaemic medications.

Protein ingestion causes increases in circulating concentrations of both insulin and glucagon with their opposing effects on blood glucose concentrations in men with and without T2D with resultant slight increases, no effect or slight decreases in blood glucose concentration when ingested on their own. Circulating glucagon concentrations after the three caloric drinks in this study were not affected by the presence of T2D. Glucose alone suppressed plasma glucagon concentrations equally in both the participant groups in this study.

We and others have previously shown that gastric emptying is slower in older than younger adults without T2D (15, 215, 222, 325). In the present study the whey protein drink slowed gastric emptying more than an equicaloric glucose drink and the combined drink slowed emptying even more, consistent with previous reports (135, 147, 388). The men with T2D in this study had good glucose control (mean HBA1c 7%) and had faster gastric emptying than those without diabetes. The faster gastric emptying is consistent with previous reports in people with early and well-controlled T2D (218, 222, 389), where there is a positive relationship between the blood glucose increment after a standardized test meal and the rate of gastric emptying, but not necessarily in people with T2D which is poorly controlled or of longer duration (225, 226, 390). The reason for accelerated gastric emptying in early

and well-controlled T2D is not known.

The changes in appetite ratings following the drinks were less in older men with vs. without T2D. Previously a study in 11 people with T2D reported an increase in fullness and decrease in hunger ratings following ingestion of 15g of whey in drink form with a mixed nutrient meal vs. control drink plus meal – the study did not include a group without T2D (391).

Our study has several limitations. Although, the number of participants were relatively small, the glucose and insulin results were clearcut and statistically significant. We studied only men, so the observation may not apply to women. The results do not necessarily apply for doses of whey protein and glucose other than 30 g as used in this study. The ability of co- or pre-ingested whey protein to reduce blood glucose increases in older people with T2D after ingestion of carbohydrate in forms other than pure glucose, for example as part of a mixed meal, seems likely but will need to be established (147). Similarly the effects of non-whey proteins and protein in non-drink form in older people with T2D, require further study.

6.7 Conclusions

In conclusion, 30 g of oral whey protein, a dose likely to have beneficial effects on skeletal muscle mass and function in older people, substantially attenuated the rises in blood glucose concentrations induced by 30 g of glucose alone in older men with and without T2D, with no significant difference between the size of that effect between the two groups. The plasma insulin and glucagon responses to protein ingestion were preserved in older participants with T2D when compared to older men without T2D. These results support the further investigation and use of combined protein ingestion with carbohydrate as a part of the treatment of older people with

T2D.

**CHAPTER 7: EFFECTS OF AGE ON BLOOD PRESSURE AND
HEART RATE RESPONSES TO WHEY PROTEIN IN
YOUNGER AND OLDER MEN**

Giezenaar C, Oberoi A, Jones KL, Horowitz M, Chapman I, Soenen S

Published in the *Journal of American Geriatrics Society*

2021

STATEMENT OF AUTHORSHIP

Title of the paper	Effects of age on blood pressure and heart rate responses to whey protein in younger and older men.
Publication status	Published
Publication details	Giezenaar C, Oberoi AO, Jones KL, Horowitz M, Chapman I, Soenen S. <i>Effects of age on blood pressure and heart rate responses to whey protein in younger and older men</i> . Journal of the American Geriatrics Society 2021; 69:1291–1299. doi: 10.1111/jgs.17083.

Candidate	Avneet Oberoi		
Contribution	Contributed to the drafting and revision of the manuscript.		
Overall percentage	75%		
Certification	This paper reports on original research I conducted during the period of my Higher Degree by Research candidature and is not subject to any obligations or contractual agreements with a third party that would constrain its inclusion in this thesis. I am the primary author of this paper.		
Signature		Date	October 2022

Principal Author

Co-Author Contributions

By signing the Statement of Authorship, each author certifies that:

- i) the candidate's stated contribution to the publication is accurate (as detailed above);
- ii) permission is granted for the candidate to include the publication in the thesis; and
- iii) the sum of all co-author contributions is equal to 100% less the candidate's stated contribution.

Name of Co-Author	Caroline Giezenaar		
Contribution	Conception and design of the manuscript, statistical analysis, data interpretation, drafting and revision of the manuscript.		
Signature		Date	October 2022

Name of Co-Author	Karen L Jones		
Contribution	Conception and design of the manuscript, data interpretation, drafting and revision of the manuscript.		
Signature		Date	October 2022
Name of Co-Author	Michael Horowitz		
Contribution	Conception and design of the manuscript, drafting and revision of the manuscript.		
Signature		Date	October 2022
Name of Co-Author	Ian Chapman		
Contribution	Conception and design of the manuscript, drafting and revision of the manuscript.		
Signature		Date	October 2022
Name of Co-Author	Stijn Soenen		
Contribution	Conception and design of the study, data interpretation, statistical analysis, drafting of the manuscript and overall responsibility for the study.		
Signature		Date	October 2022

7.1 Abstract

Background: Postprandial falls in blood pressure (BP) are more common in older compared to younger individuals. The effects of protein compared to carbohydrates and fat on postprandial BP, and the relation to gastric emptying rates, are poorly studied.

Objective: To determine the effects of a whey protein compared to a control drink on systolic BP (SBP) and diastolic BP (DBP), and heart rate (HR) in healthy younger and older men, and to relate these effects to gastric emptying.

Design: A pooled analyses of two randomized, double-blind, cross-over studies.

Setting: Two acute clinical intervention studies with identical study design.

Participants: Nineteen older (age: 74 ± 1 years, body mass index: 26 ± 1 kg/m²) and 13 younger (23 ± 1 years, 24 ± 1 kg/m²) healthy men.

Intervention: A 70 g/280 kcal whey-protein or control (water with diet cordial, ~2 kcal) drink (450 ml).

Measurements: BP and HR were assessed with an automated device immediately before and at 3-min intervals after drink ingestion (0–180 min). Gastric emptying of the drinks was measured using 3D ultrasonography (0–180 min).

Results: Older versus younger men exhibited a greater fall in SBP (-23 ± 2 vs -15 ± 2 mmHg, $P = 0.001$) after whey-protein versus control, as BP did not change after the two drinks in younger men ($P > 0.05$). The nadir in SBP occurred later in the older than younger men (114 ± 11 vs 62 ± 14 min; $P < 0.001$), with SBP still apparently declining 180 min after whey-protein ingestion in the older men. The magnitude of the rise in HR was greater ($P < 0.05$) in the younger than older men.

Conclusion: Following ingestion of 70 g whey protein, healthy older men exhibited a sustained fall in BP, despite an increase in HR, whereas in younger men there was

no change in BP. BP may need to be monitored after high protein meals in older people at risk of postprandial hypotension.

7.2 Introduction

Falls and syncope are common in older people (155). Dizziness, syncope, and falls may reflect a substantial reduction in blood pressure (BP) induced by energy intake, so-called postprandial hypotension (PPH) (261, 392). PPH occurs often in older people and represents a major cause of morbidity and mortality (262). The prevalence of PPH ranges from ~13% in healthy older people (166, 393) to ~24–36% in nursing home residents (262) (160, 394) and ~43% in hospitalized geriatric patients (395).

We have reported that glucose, when infused directly into the duodenum, at rates within the normal range for gastric emptying (1–4 kcal/min), decreased postprandial systolic BP (SBP) and diastolic BP (DBP) in healthy older, but not younger, men, particularly during the first 60 min (396). Similarly, after an oral glucose load, which is used often in the diagnosis of PPH (155), the magnitude of the fall in SBP is greater when the rate of gastric emptying is relatively more rapid (227). Other groups have similarly reported a greater reduction in SBP and DBP after mixed macronutrient meals in healthy older compared to younger adults (159). The release of gut hormones, including glucagon-like peptide-1 (GLP-1) and glucose-dependent insulinotropic polypeptide (GIP) (397-399), and an impaired sympathetic nervous system response to compensate for the meal-induced splanchnic pooling are likely to be integral to the fall in SBP in older adults (228, 263).

There is increasing emphasis on the importance of protein, especially whey protein, consumption in older people, to prevent and manage malnutrition (97). Information relating to the effects of protein on postprandial BP is limited, with some studies reporting a reduction in BP after protein (174, 263), whereas in other studies no effect

was evident (173, 175).

PPH, defined as a sustained drop in SBP of ≥ 20 mmHg for ≥ 30 min (262), is commonly diagnosed through measurement of BP for 2 h in response to a 75 g glucose load (155). This timeframe, however, may not be sufficient to diagnose PPH following protein ingestion given its slower digestion time compared to glucose (400). For example, administration of protein, carbohydrate, and fat (at a rate of 3 kcal/min) resulted in comparable reductions in SBP and a rise in superior mesenteric artery (SMA) blood flow (changes in SMA flow typically occur after nutrient ingestion) in healthy older men, but these responses were relatively delayed for protein and fat compared to glucose (263, 401).

The current study represents an analysis of secondary outcomes of previously published studies (12, 15), to determine the effect of orally ingested whey protein on SBP, DBP, heart rate (HR), and gastric emptying in healthy older and younger men. We hypothesized that older, when compared to younger, men will exhibit greater falls in SBP and DBP, and a smaller increase in HR, following whey protein ingestion compared to control.

7.3 Methods

7.3.1 Participants

Nineteen healthy older (age: mean \pm standard error of the mean [SEM]: 74 ± 1 years; body weight: 79 ± 2 kg; BMI: 26 ± 1 kg/m², 13 participants from the study described in Giezenaar et al. (12) and six participants from the study described in Giezenaar et al. (15) and 13 younger men (23 ± 1 years; 78 ± 2 kg; 24 ± 1 kg/m² from Giezenaar

et al. (12) were included. Two older participants participated in both studies; only data from the most recent study (12) were included for these two participants. The exclusion criteria were smoking, alcohol abuse, use of illicit substances, (at risk of) diabetes mellitus (HBA1C > 6.0 mmol/L), major disease (i.e., cancer, Parkinson's, multiple sclerosis, etc.), gallbladder, or pancreatic disease, gastrointestinal surgery (apart from an uncomplicated appendectomy), significant gastrointestinal symptoms (abdominal pain, gastro-esophageal reflux, diarrhea, or constipation), use of medications known to potentially affect gastrointestinal motor function, known lactose intolerance, low plasma ferritin levels, or blood donation in the 12 weeks before the study, and cognitive impairment.

Four older men took anti-hypertensive medication (angiotensin-converting enzyme inhibitor, n = 1; antiarrhythmic, n = 1; beta blockers, n = 1; and angiotensin receptor blockers, n = 1). Participants were instructed not to take their medication on the morning of their study visit. The study protocols were approved by the Royal Adelaide Hospital Research Ethics Committee (120503a and 140407, clinical trial registration: ACTRN12612000941864 and ACTRN12614000846628, www.anzctr.org.au), and the studies were conducted in accordance with the Declaration of Helsinki. All participants provided written informed consent.

7.3.2 Protocol

Both studies included in this pooled data analysis had an identical protocol and a randomized double-blind cross-over design (using the method of randomly permuted blocks; www.randomization.com) including 325 or 424 study days, separated by 3–14 days. The sub analysis related to two of these conditions, which were included in

both studies (control and 70 g whey protein).

On study days, participants fasted for ~14 h overnight, after consuming a standardized evening meal (beef lasagne [McCain Foods Pty Ltd, Australia], ~591 kcal), attended the laboratory at ~08:30 h, and ingested test drinks containing 70 g (280 kcal) whey protein or a control drink (~2 kcal) (12, 15). The drinks were prepared by a research assistant who was not involved in the analysis of the study results, flavored with diet lime cordial (Bickford's Australia Pty Ltd, South Australia) to match for taste, and served at room temperature. Drinks were presented in a covered cup to achieve blinding. Upon arrival, participants were seated in a chair, and a cannula was inserted for blood sampling (results not presented in this manuscript). An automated BP monitor (DINAMAP ProCare 100; GE Medical Systems, Milwaukee, WI) was used to measure SBP, DBP, and HR. Participants were studied for 180 min after drink ingestion, and were seated for the duration of the study.

7.3.3 Measurements

7.3.3.1 Blood pressure and heart rate

Measurements were taken three times immediately after the other at baseline (before drink ingestion), and every 3 min in the 3 h following the drink (0–180 min). Baseline BP and HR were calculated as an average of the three baseline measurements. T = 0 min refers to the point immediately after drink consumption.

7.3.3.2 Perceptions of light-headedness and drowsiness

At baseline, T = 0 min, and every 15 min after the drink, perceptions of light-headedness and drowsiness were assessed using a visual analog scale which consisted of a 100-mm horizontal line for each outcome. Participants were asked to place a mark indicating the strength of the sensation at the specified times.

7.3.3.3 Gastric emptying

At baseline, T = 0 min and every 15 min after drink ingestion, gastric volume was determined using 3-dimensional ultrasonography (Logiq 9, GE Healthcare Technologies, Australia), a method that has been validated against the “gold standard” scintigraphy for measurement of gastric emptying (230). Intra-gastric retention was calculated as total gastric volume minus baseline “empty” gastric volume at each time point, expressed as a percentage of the maximal gastric volume (100%), that is, 450 ml volume of the ingested drink. The time at which 50% of the drink was emptied from the stomach (50% gastric emptying time; T₅₀; min) was calculated.

7.4 Statistical analysis

Statistical analyses were performed using SPSS software (version 25; IBM, Armonk, NY). Interaction effects of age and treatment were determined using a two-way repeated-measures analysis of variance (ANOVA), with age as the between-participant factor, and treatment as the within-participant factor. Effects of age, time, and their interaction were determined within treatment groups. For SBP, DBP, and HR, areas under the curve (AUC) were calculated from baseline to 180 min, using the trapezoidal rule. Cumulative changes from baseline were calculated for

SBP, DBP, and HR (0–60 min, 60–120 min, and 120–180 min) by subtracting baseline values from the value at each time point, such that larger negative values indicate larger excursions below baseline, and larger positive values correspond to excursions above baseline. Nadir and time to nadir of SBP and DBP, and peak and time to peak of HR, light-headedness, and drowsiness were determined between 0 and 180 min. Assumptions of normality were verified for all outcomes before statistical analysis. Statistical significance was accepted at $P < 0.05$. Data in the text and tables are presented as mean values \pm SEM.

7.5 Results

The protocol was well tolerated in all participants. Ultrasound images were of poor quality in four older and two younger men and therefore all gastric emptying data were excluded from analyses for these participants.

7.5.1 Systolic blood pressure

Baseline SBP was higher in older than younger men (older vs younger: control, protein drink: 130 ± 3 , 131 ± 3 vs 116 ± 3 , 120 ± 4 mmHg; age main effect $P = 0.005$).

In older, but not younger men, there was a fall in SBP from baseline following the protein drink but not control. As the older men had higher baseline SBPs than younger men, absolute SBPs remained higher after control (older vs younger: 129 ± 3 vs 115 ± 3 mmHg, $P = 0.006$), but were similar after protein (123 ± 3 vs 118 ± 3 mmHg, $P = 0.27$), in older compared to younger men (AUC_{0–180min} interaction

effect of age by drink $P = 0.010$). The greatest difference in the SBP response to treatments between the two age groups occurred in the third hour after drink consumption: there was a fall in SBP (cumulative change from baseline 120 to 180 min; interaction effect of age by drink $P = 0.015$) following the whey protein drink compared to control in the older (protein vs control: -11 ± 2 vs 0 ± 2 mmHg, $P = 0.001$), but not younger (2 ± 3 vs 1 ± 2 mmHg, $P = 0.84$), men. In older (protein vs control nadir: -23 ± 2 vs -15 ± 2 mmHg, $P = 0.001$), but not younger (-13 ± 2 vs -13 ± 2 mmHg, $P = 0.96$), men SBP maximum decrease was larger after the protein compared to control drink (interaction effect of age by drink $P = 0.033$; Figure 7.1). Furthermore, the nadir in SBP occurred later in the older than younger men, independent of the study drink (older vs younger: 114 ± 11 vs 62 ± 14 min; age main effect $P < 0.001$), and SBP was still apparently falling at the end of the study period (3 h after drink ingestion) in the older men.

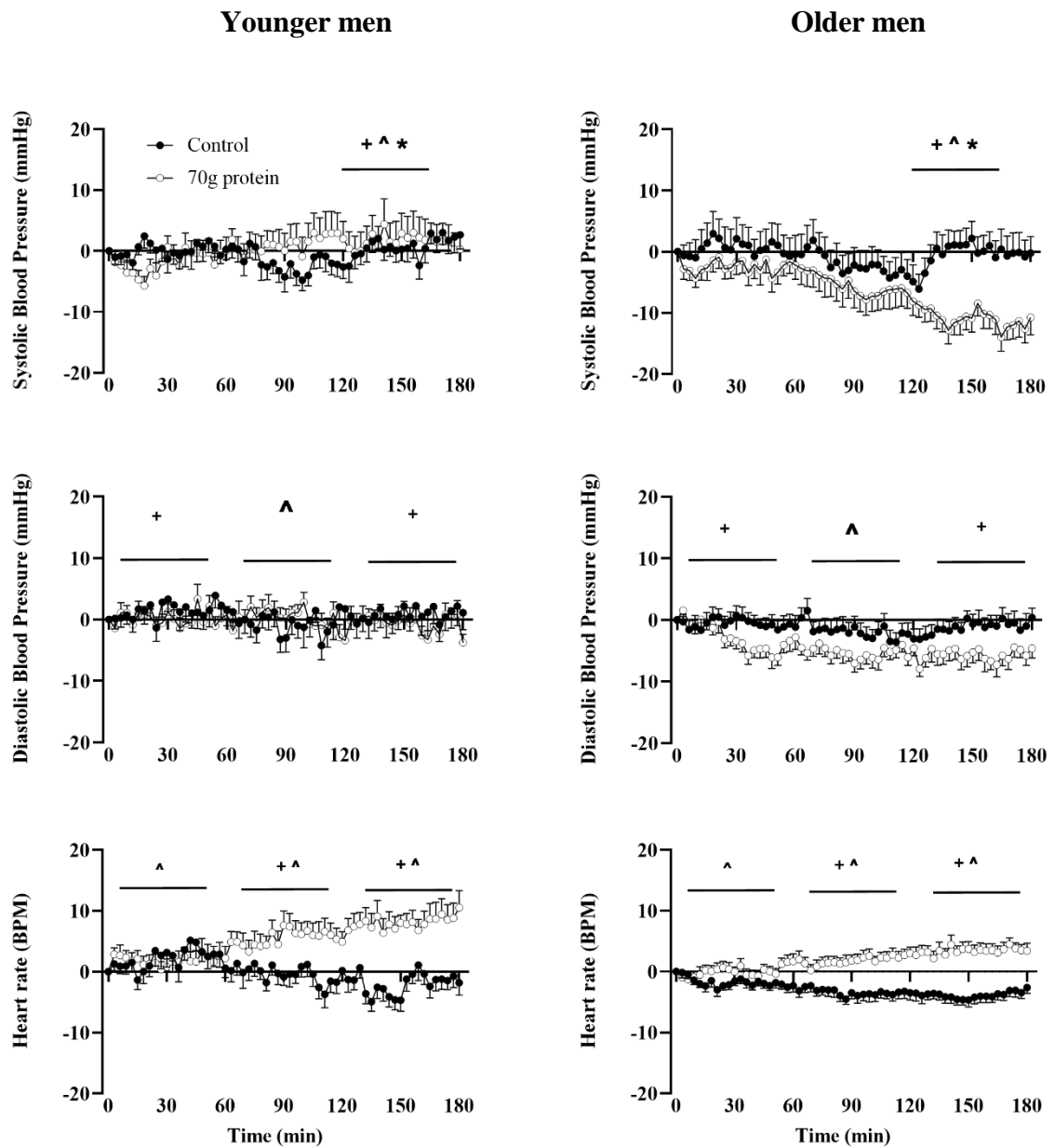


Figure 7.1: Mean (\pm standard error of mean [SEM]) cumulative changes from baseline of systolic and diastolic blood pressure (mmHg) and heart rate (beats per minute [BPM]) in 13 healthy younger and 19 healthy older men following control (\sim 2 kcal), and protein (70 g whey protein, 280 kcal), drinks. Effects of drink condition, age, and the drink by age interaction effect were determined for cumulative changes from baseline between 0–60 min, 60–120 min, and 120–180 min by repeated-measures ANOVA with post hoc Bonferroni corrections. $+P < 0.05$, the

overall effect of drink condition; $^{\wedge}P < 0.05$, the overall effect of age. $*P < 0.05$, post hoc study drink effect within the age group, control is significantly different from protein condition

7.5.2 Diastolic blood pressure

Baseline DBP was higher in older compared to younger men (older vs younger: control, protein drink: 76 ± 2 , 76 ± 2 vs 67 ± 1 , 67 ± 2 mmHg; age main effect, $P < 0.001$).

In older, but not younger men, there was a fall in DBP from baseline in response to the protein drink but not control. Accordingly, absolute DBPs were higher after control (older vs younger: 75 ± 2 vs 67 ± 2 mmHg, $P = 0.005$), but not after protein (71 ± 1 vs 67 ± 2 mmHg, $p = 0.09$), in older compared to younger men (AUC_{0-180/min} interaction effect of age by drink, $P = 0.037$).

The maximum fall in DBP was greater in older than younger men, independent of the study drink (older vs younger: -14 ± 1 vs -11 ± 1 mmHg, age main effect, $P = 0.050$; Figure 7.1). Between 60 and 120 min, there was a further fall in DBP in the older (cumulative change from baseline 60 to 120 min: -4 ± 1 mmHg), but not younger (0 ± 2 mmHg), men, independent of study drink (age main effect, $P = 0.047$).

7.5.3 Heart rate

Baseline HR was not significantly different between age groups (older vs younger: control, protein drink: 59 ± 2 , 59 ± 2 beats per minute (BPM) vs 65 ± 4 , 65 ± 3 BPM;

age main effect, $P = 0.15$).

Independent of the study drink, older compared to younger men had lower absolute HRs (older vs younger: $AUC_{0-180/\text{min}} 58 \pm 2$ vs 66 ± 3 BPM; age main effect, $P = 0.013$). Regardless of the age group, absolute HR was higher after the protein drink compared to the control drink (control vs protein: 60 ± 2 vs 65 ± 2 BPM; drink main effect, $P < 0.001$).

HR increased less from baseline in older than younger men during the first (cumulative change from baseline 0 to 60 min: older vs younger: control: -2 ± 1 vs 2 ± 1 BPM, protein: 0 ± 1 vs 2 ± 1 BPM; age main effect, $P = 0.021$), second (60–120 min: control: -3 ± 1 vs 0 ± 1 BPM, protein: 2 ± 1 vs 6 ± 2 BPM); $P = 0.016$), and third (120–180 min: control: -4 ± 1 vs -2 ± 1 BPM, protein: 3 ± 1 vs 8 ± 2 BPM; $P = 0.029$) hour after drink consumption, independent of the study drink. The maximum increase in HR was less in older than younger men, independent of the study drink (older vs younger: 6 ± 1 vs 15 ± 3 BPM; age main effect, $P < 0.001$, Figure 7.1).

7.5.4 Postprandial hypotension

In response to the control drink, 26% of older (5/19) and 23% of younger (3/13) men experienced a ≥ 20 mm Hg decrease from baseline in SBP of at some time during the following 3 h. Following the definition of PPH as a decrease of ≥ 20 mm Hg sustained for ≥ 30 min in SBP after nutrient ingestion; 10% of older (2/19) and 0% of younger men had PPH after control drink ingestion.

In response to the whey protein drink, 58% older (11/19) and 23% of younger (3/13) men demonstrated a fall of ≥ 20 mm Hg in SBP at some time during the 3 h, and 10% of older (3/19) and 0% of younger men had PPH after whey protein drink ingestion. In two of those three older men, the onset of that fall was more than 120 min after whey drink consumption. In two additional older men, there was a fall of ≥ 20 mm Hg in the six BP measurements before the end of the study period (from T = 165 to 180 min after whey ingestion). As participants were only followed until T = 180 min, these two participants may have had a sustained decrease in SBP ≥ 20 mm Hg for ≥ 30 min.

7.5.5 Perceptions of light-headedness and drowsiness

There were no age, treatment, or interaction effects on AUC or cumulative changes from baseline for perceptions of light-headedness or drowsiness (Figure 7.2). Regardless of the age group, perceptions of drowsiness increased after administration of the protein drink (time main effect, $P = 0.031$), and peak drowsiness was higher after the protein compared to the control drink ($P = 0.011$).

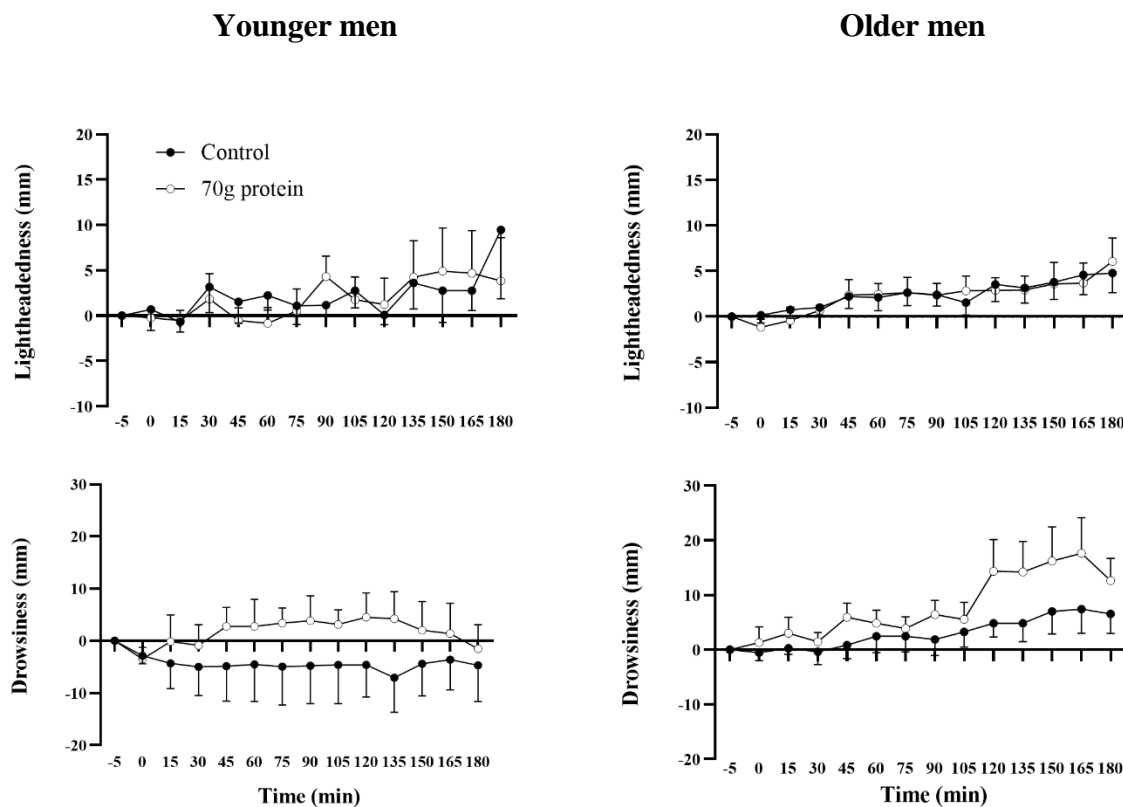


Figure 7.2 Mean (\pm standard error of mean [SEM]) cumulative changes from baseline for perceptions of light-headedness and drowsiness in 13 healthy younger and 19 healthy older men following control (\sim 2 kcal), and protein (70 g whey protein, 280 kcal), drinks. Effects of age, time, and age by time interaction effect within the study drink conditions were determined by repeated-measures ANOVA. Drowsiness: the overall effect of time: $P = 0.031$. Data are mean (\pm standard error of mean [SEM]) [Color figure can be viewed at wileyonlinelibrary.com]

7.5.6 Gastric emptying

In older men, gastric emptying of the protein drink (T_{50} ; older vs younger: 70 ± 6 min vs 58 ± 9 min, $P = 0.019$), but not control (12 ± 1 vs 12 ± 1 min, $P = 0.15$), was

slower, when compared to the younger men (interaction effect of age by study drink, $P = 0.027$; Figure 7.3).

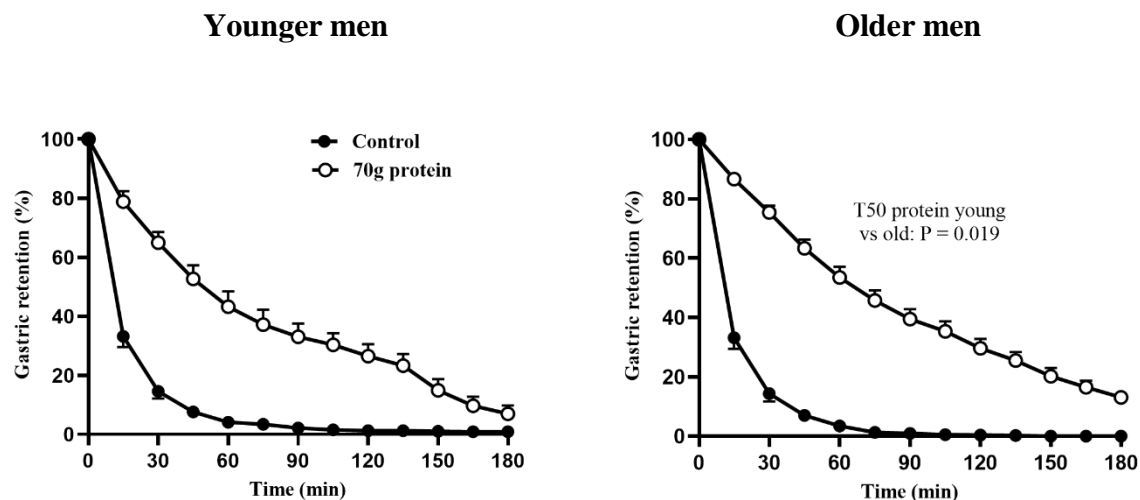


Figure 7.3: Mean (\pm standard error of mean [SEM]) gastric retention (%) in 11 healthy younger and 14 healthy older men following control (~ 2 kcal), and protein (70 g whey protein, 280 kcal), drinks. Effects of drink-condition, age, and the drink-by-age interaction effect were determined by repeated-measures ANOVA with post hoc Bonferroni corrections. Interaction effect of age by drink condition: $P = 0.027$ [Color figure can be viewed at wileyonlinelibrary.com]

7.6 Discussion

Our study has established that, in response to a 280 kcal (70 g) whey protein drink (450 ml) ingestion, healthy older men exhibit a substantial decrease in SBP and DBP, whereas younger men maintained their BP at a level comparable to that after an iso-volumic, non-caloric, drink. Furthermore, in the older participants, the fall in BP was sustained until at least 180 min after consumption of the drink. There was an increase in HR after the protein drink, but this increase in HR was greater in the younger than older men. The perception of drowsiness, but not light-headedness, was greater after

protein irrespective of age. The use of protein supplements to aid undernutrition in older people is increasing, and PPH is particularly common in institutionalized older patients, who are more likely to take nutritional supplements (97). It is certainly not appreciated that PPH is a potential outcome of the use of protein supplements in this group; with emergent increased rates of falls, syncope, and death (155). All our participants were seated, and orthostatic hypotension was not assessed. Posture likely affects postprandial BP (402) and it is possible that the lowering of BP that occurred in the older participants may have been caused partially by the upright seated position during the study.

Our observations also indicate that, in studies evaluating BP after protein administration, BP should be monitored for a minimum of 3 h. PPH is defined as a sustained fall in BP ≥ 20 mm Hg within 2 h of a meal (262). This definition is based on the outcome of studies using high-carbohydrate meals or glucose drinks to diagnose PPH, and our observations suggest that the definition should be modified to increase this time frame—at least after a high protein (i.e., more than 30 g) meal or supplement.

The current findings are consistent with our previous observations, which showed a decrease in BP in response to intraduodenal glucose infusion in older, but not younger, adults (396). We have also reported a more delayed hypotensive response after an intraduodenal infusion of protein compared to glucose in healthy older people (263). This difference may be attributed to differences in time for digestion of carbohydrates and protein (400) to stimulate an increase in superior mesenteric artery (SMA) blood flow response (401); that is, the fall in BP may be triggered by amino acids, the digestion products of protein. In the younger participants, the rise

in HR induced by the protein drink was greater than in the older participants, which is likely to contribute to the maintenance of BP despite a substantial increase in splanchnic blood flow in both age groups (396). The increase in splanchnic blood flow after the protein drink may be reflected by the increase in perceptions of drowsiness which occurs in both age groups. In contrast with our results, another study with a very similar study design, in which BP was measured every 5 min for 150 min after 75 g oral loads of glucose, whey protein, and cream, older hypertensive patients showed a fall in SBP and DBP after glucose, but not after fat and protein. Furthermore, HR increased after glucose and fat but was not affected by protein intake (173). In the current study, participants were seated throughout the study (i.e., 3 h), whereas the participants in the described study (173) were in a semi-recumbent position, reducing effects of potential orthostatic hypotension. In patients with autonomic failure, the fall in postprandial BP was also larger after carbohydrates and fat (403), and in healthy young participants, two iso-caloric loads containing 92 g of either carbohydrates or protein resulted in increased cardiovascular output and ventricular contractility after carbohydrates compared to protein (404). These studies may suggest that even though we reported a fall in BP after protein, this fall may not be as large as a fall in response to carbohydrates.

As shown in this study, gastric emptying of protein was slower in the older compared to younger participants (12, 15). Slower gastric emptying should, intuitively, be associated with increased, and more sustained, gastric distension which is known to attenuate the postprandial fall in BP (228). In older people, this may, in part, represent a compensatory mechanism to reduce the magnitude of a fall in BP after a meal.

This study has several potential limitations. The study represents a pooled data analysis from two separate studies, and whereas the methodology in terms of inclusion criteria, timeline, and drink composition were identical in both studies, there is the inherent potential for bias. We evaluated the effects of a relatively large protein dose (70 g), representative of a normal daily intake of protein— a meal typically contains 15–30 g of protein. Protein supplements used to manage undernutrition in older people typically contain 10–25 g protein, and therefore, use of these may not necessarily be associated with falls in BP as large as those observed in this study. We only studied whey protein, and therefore, the results may not apply to other protein sources. We only studied healthy men, and therefore, the results may not be translatable to women. Given that there was a fall in BP in response to whey protein in healthy older men, it is likely that the fall would be greater in populations at risk of PPH (e.g., diabetic, hospitalized and geriatric patients, and patients with neurological disorders). This warrants investigation as these are patient groups that are most likely to benefit from increased protein intake through the use of protein supplements. We did not measure autonomic nerve function or orthostatic BP, and therefore we were unable to determine whether nerve dysfunction may be an underlying factor in the age-related results on BP and gastric emptying reported in this study. We acknowledge that the inclusion of participants with hypertension may have affected the BP responses in older adults.

In summary, healthy older men exhibited a sustained fall in systolic and diastolic BP, despite an increase in HR, following ingestion of 70 g whey protein, whereas in younger men the increase in HR was greater and there was no change in BP. In future research, it should be investigated whether the substitution of carbohydrates and/or fat by protein slows gastric emptying, and thereby attenuates the postprandial fall in

BP in older people. In conclusion, these observations suggest that postprandial BP after protein supplements or high protein meals may need to be monitored in older people, particularly those at particular risk of PPH.

**CHAPTER 8: BLOOD PRESSURE AND HEART RATE
RESPONSES FOLLOWING DIETARY PROTEIN INTAKE IN
OLDER MEN**

**Oberoi A, Giezenaar C, Lange K, Jones KL, Horowitz M, Chapman I,
Soenen S**

Published in *Nutrients*

2022

STATEMENT OF AUTHORSHIP

Title of the paper	Blood pressure and heart rate responses following dietary protein intake in older men.
Publication status	Published
Publication details	Oberoi AO, Giezenaar C, Lange K, Jones KL, Horowitz M, Chapman I, Soenen S. <i>Blood pressure and heart rate responses following dietary protein intake in older men.</i> <i>Nutrients</i> 2022; 14: 1913. doi:10.3390/nu14091913.

Candidate	Avneet Oberoi		
Contribution	Contributed to the overall design of the manuscript, literature review, data analysis and interpretation, drafting and revision of the manuscript.		
Overall percentage	75%		
Certification	This paper reports on original research I conducted during the period of my Higher Degree by Research candidature and is not subject to any obligations or contractual agreements with a third party that would constrain its inclusion in this thesis. I am the primary author of this paper.		
Signature		Date	October 2022

Principal Author***Co-Author Contributions***

By signing the Statement of Authorship, each author certifies that:

- i) the candidate's stated contribution to the publication is accurate (as detailed above);
- ii) permission is granted for the candidate to include the publication in the thesis; and
- iii) the sum of all co-author contributions is equal to 100% less the candidate's stated contribution.

Name of Co-Author	Caroline Giezenaar		
Contribution	Drafting and revision of the manuscript.		
Signature		Date	October 2022

Name of Co-Author	Kylie Lange		
Contribution	Conception and design of the manuscript, statistical analysis, data interpretation, drafting and revision of the manuscript.		
Signature		Date	October 2022
Name of Co-Author	Karen L Jones		
Contribution	Conception and design of the manuscript, data interpretation, drafting and revision of the manuscript.		
Signature		Date	October 2022
Name of Co-Author	Michael Horowitz		
Contribution	Conception and design of the manuscript, drafting and revision of the manuscript.		
Signature		Date	October 2022
Name of Co-Author	Ian Chapman		
Contribution	Conception and design of the manuscript, statistical analysis, data interpretation, drafting and revision of the manuscript.		
Signature		Date	October 2022
Name of Co-Author	Stijn Soenen		
Contribution	Conception and design of the study, data interpretation, statistical analysis, drafting of the manuscript and overall responsibility for the study.		
Signature		Date	October 2022

8.1 Abstract

Postprandial hypotension (PPH) occurs frequently in older people > 65 years old. Protein-rich supplements, particularly whey protein (WP), are increasingly used by older people for various health benefits. We have reported that 70 g WP drinks cause significant, and in some cases marked, falls in blood pressure (BP) in older men. The effects of lower, more widely used, doses (~30 g) on systolic (SBP) and diastolic (DBP) blood pressure and heart rate (HR) are not known. In a randomized order, eight older men (age: 72 ± 1 years; BMI: 25 ± 1 kg/m²) after overnight fast ingested a drink containing (i) a non-caloric control (~2 kcal), (ii) 30 g of whey protein (120 kcal; 'WP30'), or (iii) 70 g of whey protein (280 kcal; 'WP70'). The BP and HR were measured in this pilot study with an automated device before and at 3-min intervals for 180 min following drink ingestion. Drink condition effects were determined by repeated-measures ANOVA. The SBP decreased after both WP drinks compared to the control ($P = 0.016$), particularly between 120 and 180 min, with no difference in the effects of WP30 and WP70. The SBP decreased by ≥ 20 mm Hg in more than 50% of people after both WP drinks (WP30: 63%; WP70: 75%) compared to 38% after the control. The maximum fall in the SBP occurred during the third hour, with the nadir occurring latest after WP70. The DBP decreased non-significantly by several mmHg more after the WP drinks than after the control. The maximum HR increases occurred during the third hour, with the greatest increase after WP70. The SBP decreased after both WP drinks compared to the control, with the effects most evident between 120 and 180 min. Accordingly, ingestion of even relatively modest protein loads in older men has the potential to cause PPH.

8.2 Introduction

Human ageing is associated with reductions in skeletal muscle mass and strength, which are associated with increased rates of falls and nursing home admissions, as well as other adverse outcomes (111, 306, 310-315). Falls may also be caused by dizziness and/or syncope due to postprandial hypotension (PPH), a substantial reduction in blood pressure (BP) caused by ingestion of nutrients (261, 392), which has been defined as a decrease in systolic blood pressure (SBP) of ≥ 20 mm Hg for ≥ 30 min within 120 min of consumption of a meal (262). PPH occurs frequently in older people and is associated with increased morbidity and mortality (262).

A strategy increasingly adopted to prevent or treat under-nutrition, weight loss, and sarcopenia in older people is to increase consumption of high-energy, protein-rich supplements (97). These supplements are often rich in whey protein (15), a component of milk, which is high in essential amino acids, particularly leucine, which are rapidly digested to increase postprandial amino acid availability and stimulate muscle protein accretion (7, 319).

Oral ingestion of whey protein or whey-protein-rich supplements has the potential to reduce the BP to a degree that is harmful in some older people, predisposing to falls and other adverse effects. Compared to younger adults, older people exhibit greater decreases in the BP after meals (405). Ingestion of all three macronutrients (carbohydrate, protein, fat) decreases the post-prandial BP in older people (263, 264). Carbohydrate and protein lower the BP to a similar degree, although the fall in the BP may occur earlier after carbohydrate ingestion than protein ingestion (263). The hypotensive effects of protein are likely to be mediated by amino acids produced by

digestion, explaining the latency and time of onset of changes in the BP and HR after protein loads. We have recently reported that ingestion of a 70 g whey protein drink is associated with a substantial decrease in the BP in healthy older men; the majority of older men studied had a decrease in the systolic BP (SBP) of 20 mm Hg or more, with the greatest reduction occurring 2–3 h after drink ingestion (264).

Overall, 70 g of protein is more than most older people ingest at one time; amounts of ~30 g are probably sufficient to induce muscle synthesis in older people, particularly if ingested twice a day (63). It is not clear whether the hypotensive effects of whey protein drinks are dose dependent in older people and whether whey doses less than 70 g also cause a substantial fall in the BP. We have reported that ingestion of a whey protein drink as a preload in combination with guar in a much lower dose of 16.4 g has no effects on the BP in healthy older people for up to 2 h (406). Two hours may not, however, have been long enough to detect the maximum hypotensive effects of the drink in that study.

This study compared the effects over 3 hours of 30 g and 70 g whey protein drinks on the BP and heart rate (HR) in older men. This pilot study is a subset (n = 8 males) of a larger study and represents an analysis of secondary outcomes measured in men (BP and HR) in a previously published study (14) that described the effect of orally ingested whey protein on energy intake, gastric emptying, and plasma gut-hormone concentrations in older men (n = 8) and women (n = 8).

8.3 Materials and methods

8.3.1 Participants

Eight older (mean age: 73 ± 1 years; body weight: 77 ± 4 kg; body mass index (BMI): 26 ± 1 kg/m²) men were recruited by advertisement.

Exclusion criteria included alcohol intake of > 2 standard drinks on > 5 days per week; smoking; intake of any illicit substance; use of prescribed or non-prescribed medications that may affect appetite, body weight, gastrointestinal function, or energy metabolism; being vegetarian; known lactose intolerance or food allergies; epilepsy; gallbladder, pancreatic, cardiovascular, or respiratory diseases; significant gastrointestinal symptoms (abdominal pain, gastroesophageal reflux, diarrhea, or constipation) or surgery; any other illness deemed significant by the investigator; low plasma ferritin levels; donation of blood in the 12 weeks prior to the study days; undernourished condition (score < 24 on the Mini Nutritional Assessment (407)); or depression (score ≥ 11 on the Geriatric Depression Questionnaire (327)), impaired cognitive function (score < 25 on Mini Mental State (326)), and inability to comprehend the study.

The Royal Adelaide Hospital Human Research Ethics Committee approved the proto-col, which was conducted in accordance with the Declaration of Helsinki. The study was registered with the Australian New Zealand Clinical Trial Registry (www.anzctr.org.au) (accessed on 15 October 2021; registration number ACTRN12612000941864). All participants provided written informed consent prior to their study inclusion.

8.3.2 Protocol

The protocol was similar to that of our previous study comparing older and younger men (15). Each participant was studied on 3 occasions, separated by 3–14 days, to determine the effects of drinks (~450 mL) containing (i) a non-caloric control (~2 kcal; ‘C’), (ii) 30 g of whey protein (120 kcal; ‘WP30’), or (iii) 70 g of whey protein (280 kcal; ‘WP70’) on the SBP, DBP, and HR in a randomized (using the method of randomly permuted blocks; www.randomization.com; accessed on 3 August 2018), double-blind, cross-over design.

The drinks were prepared by dissolving whey protein isolate (Fonterra Co-Operative Group Ltd., Palmerston North, New Zealand) in varying volumes of demineralized water and diet lime cordial (Bickford’s Australia Pty Ltd., Salisbury South, SA, Australia), to achieve the desired composition, by a research officer who was not involved in the data analysis on the morning of the study day. The drinks were matched for taste and served in a covered cup so that both the participant and the investigator were blinded to the treatment.

Participants consumed a standardized meal (beef lasagne (McCain Foods Pty Ltd., Wen-douree, Victoria, Australia), ~591 kcal) on the night before each study day at ~1900 h. They were instructed to fast overnight ~12 h, taking no solids and liquids except water.

8.3.3 Measurements

Participants were seated in an upright position on a wooden chair with arms, where they remained throughout the study day. They were asked to sit quietly for at least 15 min, and the BP was measured 3 times at 3-min intervals using an automatic BP-measuring device (DINAMAP Pro Care 100; GE Medical Systems, Milwaukee, WI, USA). Participants were then instructed to consume the drink within 2 min, and the BP and HR were measured every 3 min for a further 180 min. When SBP measurements between consecutive samples varied by ≥ 10 mm Hg, repeat measurements were taken.

8.4 Data and statistical analysis

Responses to the drinks were calculated for the first, second, and third hour following drink ingestion as the mean of individual measurements (mean 0–60 min, mean 60–120 min, mean 120–180 min, respectively). All data are presented as mean values \pm standard error of the mean (SEM), and statistical significance was accepted at $P < 0.05$. Statistical analyses were performed using SPSS software (version 24; IBM, Armonk, NY, USA). The maximum decrease from baseline was calculated as the difference between the minimum value and baseline and the time of nadir that was analyzed for the SBP, DBP, and HR.

The baseline blood pressure was calculated as an average of -9 , -6 , and -3 min readings. $T = 0$ min refers to the point immediately after drink consumption. Mixed effects model analysis including treatment as a fixed effect, and an unstructured covariance structure was used to determine the effect of protein load on the SBP,

DBP, and HR baseline levels and each of the three mean outcomes, along with the maximum decrease and the time to nadir. When significant treatment effects were present, Bonferroni corrected post hoc tests were performed to determine which specific drink conditions differed.

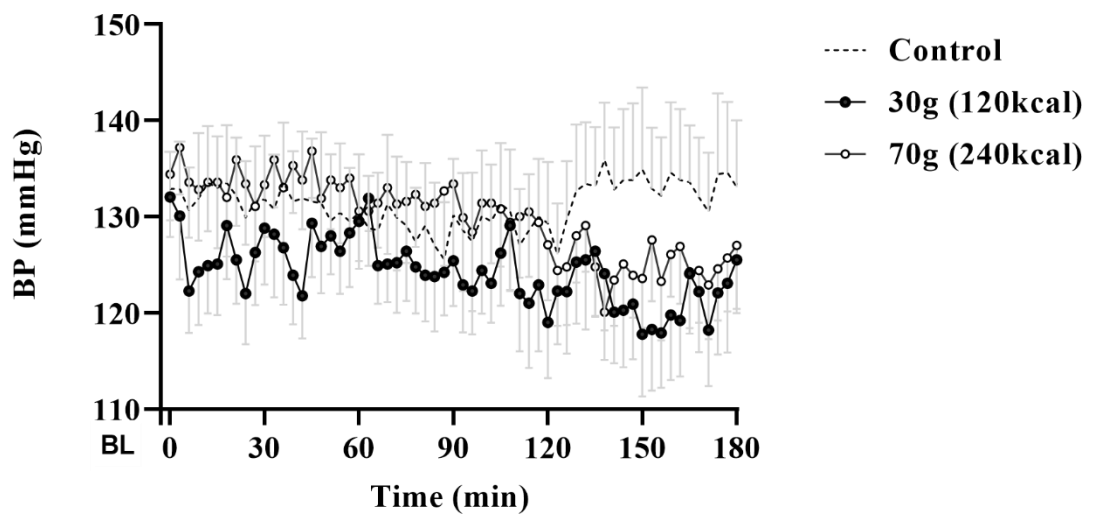
8.5 Results

The study protocol was well tolerated by all participants.

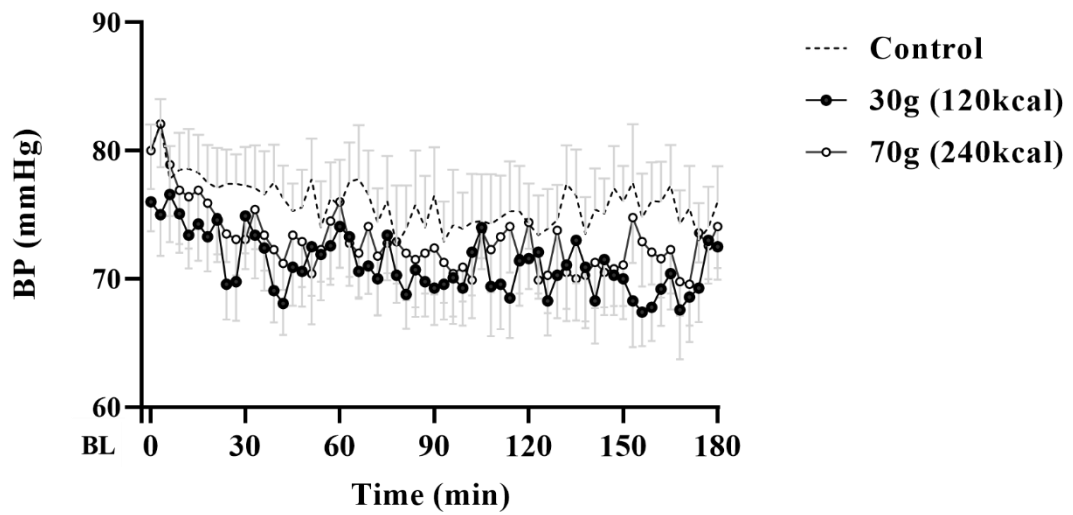
8.5.1 Systolic blood pressure (SBP)

The baseline SBP was comparable on the 3 study days (C: 133 ± 4 mm Hg; WP30: 132 ± 4 mm Hg; WP70: 134 ± 5 mm Hg; $P = 0.95$). During the first 2 hours, SBPs after all drinks were not significantly different (mean 0–60 min, mean 60–120 min; C: 132 ± 6 , 129 ± 6 mm Hg, respectively; WP30: 127 ± 4 , 124 ± 5 mm Hg, respectively; WP70: 134 ± 5 , 131 ± 6 mm Hg, respectively; $P = 0.12$, $P = 0.09$, respectively). In contrast, between 120 and 180 min, the SBP was lower after both whey protein drinks than after the control drink ($P = 0.016$; C vs. WP30 $P = 0.02$; C vs. WP70, $P = 0.20$), with mean SBP changes from baseline of -10 mmHg after both whey protein drinks during the third hour compared to 0 mmHg after the control drink (mean 120–180 min; C: 133 ± 7 mm Hg; WP30: 122 ± 6 mm Hg; WP70: 125 ± 5 mm Hg; Figure 8.1A, Table 8.1).

Systolic Blood Pressure



Diastolic Blood Pressure



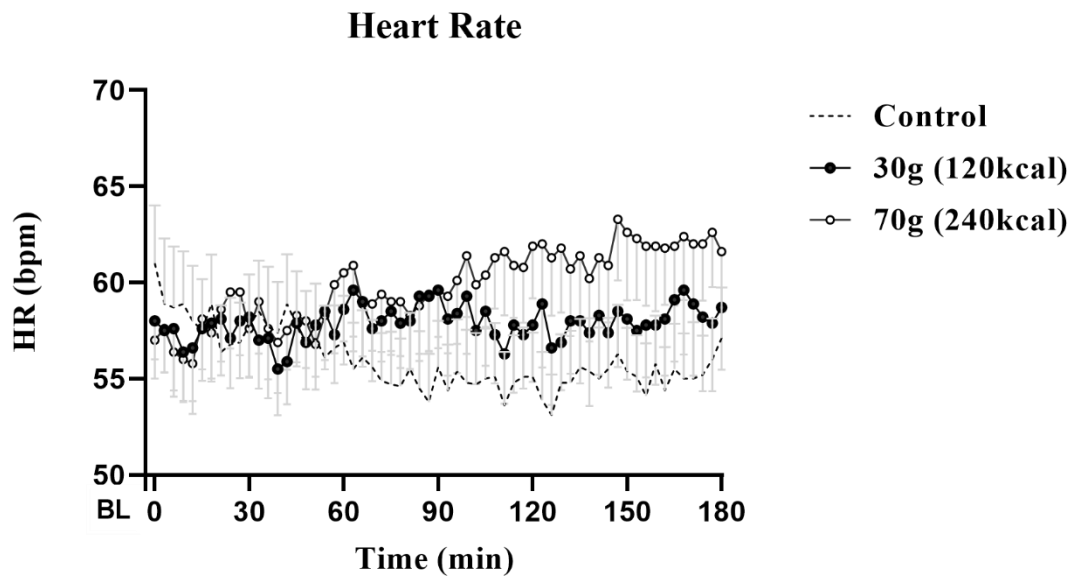


Figure 8.1: Mean (\pm SEM) systolic (A) and diastolic (B) blood pressure (mmHg) and heart rate (bpm) (C) following ingestion of drinks containing (i) flavored water (control, \sim 2 kcal) or (ii) 30 g (120 kcal) of whey protein (WP30) or (iii) 70 g (280 kcal) of whey protein (WP70) in older ($n = 8$) men. The base-line blood pressure represented as ‘BL’ in the figures was calculated as an average of measurements at -9 , -6 , and -3 min. $T = 0$ min refers to the point immediately after drink consumption. Drink condition effects were determined by using mixed model analysis. SBPs were lower after the whey protein drinks when compared to the control drink during the third hour ($p < 0.05$). The mean fall in the DBP was greater after 30 g and 70 g of whey protein drinks than after the control drink. The HR was higher after the whey protein drinks when compared to the control drink during the second ($P = 0.007$) and third ($P = 0.005$) hours, with the greatest increase after WP70 ($P = 0.002$).

Table 8.1: Blood pressure and heart rate after drink ingestion

	Control	WP30	WP70	<i>P</i>
SBP (mmHg)				
Mean 0-60 min	132 ± 6	127 ± 4	134 ± 5	0.123
Mean 60-120 min	129 ± 6	124 ± 5	131 ± 6	0.092
Mean 120-180 min	133 ± 7	122 ± 6	125 ± 5	0.016
Max change from baseline	-15 ± 2	-24 ± 2	-25 ± 4	0.020
Time to Nadir(min)	90 ± 8	111 ± 22	136 ± 11	0.084
DBP (mmHg)				
Mean 0-60 min	77 ± 7	73 ± 2	75 ± 3	0.235
Mean 60-120 min	75 ± 3	71 ± 3	72 ± 3	0.414
Mean 120-180 min	75±3	70±23	72±3	0.202
Max change from baseline	-13 ± 2	15 ± 1	-17 ± 2	0.13
Time to nadir (min)	89 ± 19	98 ± 19	108 ± 11	0.74
HR (bpm)				
Mean 0-60 min	55 ± 3	58 ± 3	59 ± 3	0.057
Mean 60-120 min	53 ± 2	60 ± 3	60 ± 3	0.007
Mean 120-180 min	53 ± 2	58 ± 3	62 ± 3	0.005
Max change from baseline	2 ± 1	6 ± 1	10 ± 2	0.002
Time to nadir (min)	71 ± 25	112 ± 17	120 ± 17	0.181

Mean (± SEM) systolic blood pressure (SBP, mm Hg), diastolic blood pressure (DBP, mm Hg) and heart rate (beats per minute, bpm) following ingestion of drinks containing (i) flavored water (control, ~2 kcal) or (ii) 30g (120 kcal) whey protein (WP30) (iii) 70g (280 kcal) whey protein (WP70) in older (n = 8) men. P value

treatment effect of mixed model analysis.

Maximum decreases in the SBP from baseline (C: -15 ± 2 mm Hg; WP30: -24 ± 2 mm Hg; WP70: -25 ± 4 mm Hg; $P = 0.02$) occurred from ~ 2 h after drink ingestion onward, with the nadir (C: 118 ± 5 mm Hg; WP30: 108 ± 5 mm Hg; WP70: 110 ± 4 mm Hg) occurring latest after the 70 g whey protein drink (C: 90 ± 8 min; WP30: 111 ± 22 min; WP70: 136 ± 11 min; $P = 0.08$). The SBP decreased by ≥ 20 mm Hg in more than 50% of participants after both whey protein drinks (C: 3/8 (38%); WP30: 6/8 (75%); WP70: 5/8 (63%)).

8.5.2 Diastolic blood pressure (DBP)

The baseline DBP was not different on the study days (C: 80 ± 2 mm Hg; WP30: 76 ± 2 mm Hg; WP70: 80 ± 3 mm Hg; $P = 0.35$). The mean fall in the DBP was greater after 30 g and 70 g whey protein drinks than after the control drink, which did not achieve statistical significance during any period. Mean decreases in the DBP from baseline during the third hour were $-6/ -8$ mm Hg after WP30/WP70 compared to -4 mmHg after the control drink (mean 0–60 min, mean 60–120 min, mean 120–180 min; C: 77 ± 7 , 75 ± 3 , 75 ± 3 mm Hg, respectively; WP30: 73 ± 2 , 71 ± 3 , 70 ± 3 mm Hg, respectively; WP70: 75 ± 3 , 72 ± 3 , 72 ± 3 mm Hg, respectively; $P = 0.23$, $P = 0.41$, $P = 0.20$, respectively; Figure 8.1B, Table 8.1). Maximal changes in the DBP from baseline (C: -13 ± 2 mm Hg; WP30: -15 ± 1 mm Hg; WP70: -17 ± 2 mmHg; $P = 0.13$) occurred ~ 1.5 –2 h after drink ingestion, with the nadir occurring latest after the 70 g whey protein drink (C: 89 ± 19 min; WP30: 98 ± 19 min; WP70: 108 ± 11 min; $P = 0.74$).

8.5.3 Heart rate (HR)

The baseline HR was not different between the study days (C: 61 ± 3 bpm; WP30: 58 ± 2 bpm; WP70: 57 ± 2 bpm; $P = 0.56$). The HR was higher after the whey protein drinks when compared to the control drink during the last 2 hours (mean 0–60 min, mean 60–120 min, mean 120–180 min; C: 55 ± 3 , 55 ± 2 , 55 ± 2 bpm, respectively; WP30: 58 ± 3 , 60 ± 3 , 58 ± 3 bpm, respectively; WP70: 59 ± 3 , 60 ± 3 , 62 ± 3 bpm, respectively; $P = 0.05$, $P = 0.007$, $P = 0.005$, respectively), with the greatest increase after WP70 and during the third hour (change from baseline to 120–180 min: C: -5 ± 1 bpm; WP30: 0 ± 1 bpm; WP70: $+5 \pm 1$ bpm; $P < 0.05$; Figure 8.1C, Table 8.1). The maximal increase in the HR from baseline (C: 2 ± 1 bpm; WP30: 6 ± 1 bpm; WP70: 10 ± 2 bpm; $P = 0.002$) occurred ~ 1.5 –2 h after drink ingestion (C: 71 ± 25 min; WP30: 112 ± 17 min; WP70: 120 ± 17 min; $P = 0.181$).

8.6 Discussion

We demonstrated that healthy older men exhibit a decrease in the SBP after ingestion of both 30 g and 70 g whey protein drinks, which was similar in degree after both whey protein drinks and the greatest between 120 and 180 min after their ingestion. Previously, we reported that protein, when administered directly into the duodenum and thereby bypassing gastric effects, lowers blood pressure comparably to glucose and fat, with the onset of the hypotensive response being earlier after glucose ingestion (263). In addition, the type of protein may affect lowering of blood pressure, and certain proteins (e.g., in ancient wheat) have an effect on endothelial reactivity (408).

Protein supplements, usually in drink form, are increasingly administered to older people, particularly the institutionalized elderly, who are at a high risk of postprandial hypotension. While we have shown that protein loads of 70 g can lead to substantial falls in the SBP (264), this amount of protein is greater than usually taken at one time by older people. Lower doses are more likely to be used in supplements for older people, and there is evidence that ~30 g doses, once or twice daily, are likely to be beneficial for weight, nutrition, and muscle preservation (63). Our observation suggests that care may need to be taken to monitor and prevent the hypotensive effects of protein supplements in susceptible older people, even when used in doses as low as 30 g and for 3 hours or even longer after protein supplement ingestion; our observations finished at 3 h when the SBP was possibly still reduced. The heart rate (HR) increased after both whey protein drinks, with the greatest increase after the 70 g drink. These dose-responsive HR increases help to maintain cardiac output and hence the BP in the face of diversion of blood to the gut to aid protein absorption and digestion. The greater compensatory increase in the HR (and thus cardiac output) after the 70 g than the 30 g drink is likely to be a factor in the two doses lowering the BP to a similar degree. This might also suggest that if the compensatory HR increases after protein (or other nutrient) ingestion are impaired in older people, the decrease in BP after nutrient ingestion will be greater and potentially more likely to lead to falls and other adverse effects. This is supported by our previous finding that in young men, there were greater increases in the HR and lesser decreases in the SBP than in older men after 70 g whey protein drinks (264). Older people taking medications such as beta-blockers, which limit the heart rate and heart rate responses, or with cardiac conditions leading to bradycardia may be at even greater risk of post-protein BP falls. Following a standard mixed macronutrient breakfast meal, 16 participants over 75 years old showed a significant fall in blood

pressure, which may, in some less robust elderly persons, contribute to falls, while the younger control group did not show a significant decrease in postprandial blood pressure (159).

Our study has some limitations. The study was only conducted in healthy men, with a relatively small number of participants. The results may not be translatable to women. While our observation period of 3 hours was longer than in most previous studies, the BP was possibly still decreasing and the HR was still higher than on the control day 3 h after the protein drinks. A longer study duration would have helped determine the full time course of the effects of protein drinks. Since only whey protein was studied, the observation cannot be applied to other protein sources. A greater protein-induced BP fall than observed in this study might occur in older people at greater risk of postprandial hypotension than our participants, for example, institutionalized, frail older people. A possible safety issue for older people adopting a program of post-protein supplement or postprandial exercise might be excessive BP drops leading to falls.

Our result suggest that if the intention is to give an older person a nutritional supplement drink containing 30–70 g of whey protein to preserve or even enhance muscle mass and function, excessive post-protein BP is a possibility, particularly in those most at risk. Consideration should be given to monitoring for this and/or advising measures (such as care when standing) to reduce the harmful effects of excessive postprandial BP decreases. It would also be appropriate in future studies to examine the effects on the BP in older people of combining protein and other nutrients with exercise.

8.7 Conclusions

The SBP decreased after both WP drinks but was not dose dependent compared to the control, with the effects most evident between 120 and 180 min. Accordingly, ingestion of even relatively modest protein loads in older men has the potential to cause PPH.

**CHAPTER 9: ACUTE EFFECTS OF WHEY PROTEIN,
ALONE AND MIXED WITH OTHER MACRONUTRIENTS,
ON BLOOD PRESSURE AND HEART RATE IN OLDER
MEN**

**Oberoi A, Giezenaar C, Lange K, Jones KL, Horowitz M, Chapman I,
Soenen S**

Published in *BMC Geriatrics*

2022

STATEMENT OF AUTHORSHIP

Title of the paper	Acute effects of whey protein, alone and mixed with other macronutrients, on blood pressure and heart rate in older men		
Publication status	Published		
Publication details	Oberoi AO, Giezenaar C, Lange K, Jones KL, Horowitz M, Chapman I, Soenen S. <i>Acute effects of whey protein, alone and mixed with other macronutrients, on blood pressure and heart rate in older men</i> . BMC Geriatrics 2022; 22: 535. doi:10.1186/s12877-022-03213-1		
Candidate	Avneet Oberoi		
Contribution	Contributed to the overall design of the manuscript, literature review, data analysis and interpretation, drafting and revision of the manuscript.		
Overall percentage	75%		
Certification	This paper reports on original research I conducted during the period of my Higher Degree by Research candidature and is not subject to any obligations or contractual agreements with a third party that would constrain its inclusion in this thesis. I am the primary author of this paper.		
Signature		Date	October 2022

Principal Author*Co-Author Contributions*

By signing the Statement of Authorship, each author certifies that:

- i) the candidate's stated contribution to the publication is accurate (as detailed above);
- ii) permission is granted for the candidate to include the publication in the thesis; and
- iii) the sum of all co-author contributions is equal to 100% less the candidate's stated contribution.

Name of Co-Author	Caroline Giezenaar		
Contribution	Drafting and revision of the manuscript		
Signature		Date	October 2022
Name of Co-Author	Kylie Lange		
Contribution	Conception and design of the manuscript, statistical analysis, data interpretation, drafting and revision of the manuscript.		
Signature		Date	October 2022
Name of Co-Author	Karen L Jones		
Contribution	Conception and design of the manuscript, data interpretation, drafting and revision of the manuscript.		
Signature		Date	October 2022
Name of Co-Author	Michael Horowitz		
Contribution	Conception and design of the manuscript, drafting and revision of the manuscript.		
Signature		Date	October 2022
Name of Co-Author	Ian Chapman		
Contribution	Conception and design of the manuscript, statistical analysis, data interpretation, drafting and revision of the manuscript.		
Signature		Date	October 2022
Name of Co-Author	Stijn Soenen		
Contribution	Conception and design of the study, data interpretation, statistical analysis, drafting of the manuscript and overall responsibility for the study.		

Signature		Date	October 2022
-----------	--	------	--------------

9.1 Abstract

Background: Caloric supplements are increasingly used by older people, aiming to increase their daily protein intake. These high caloric drinks, rich in glucose and whey-protein in particular, may result in potential harmful decreases in blood pressure (BP). The effect of ingesting whey-protein with glucose and fat on BP is unknown. It has also been assumed that the maximum fall in systolic blood pressure occurs within 2 h of a meal.

Methods: This study aimed to determine in older men, the effects of whey-protein, alone and mixed with other macronutrients, on systolic (SBP) and diastolic (DBP) blood pressure and heart rate (HR) in older men for 3 h. Thirteen older men (age 75 ± 2 yrs; body mass index (BMI) 25.6 ± 0.6 kg/m²) ingested a drink on separate study days: (i) 70 g whey-protein (P₂₈₀); (ii) 14 g whey-protein, 28 g carbohydrate, 12.4 g fat (M₂₈₀); (iii) 70 g whey-protein, 28 g carbohydrate, 12.4 g fat (M₅₀₄); or (iv) a non-caloric control drink (C).

Results: SBP decreased after all three nutrient drinks compared to the C, with the greatest reduction after the M₅₀₄ drink ($P = 0.008$). Maximal decreases in SBP (C: -14 ± 2 mm Hg, P₂₈₀: -22 ± 2 mm Hg, M₂₈₀: -22 ± 4 mm Hg, M₅₀₄: -24 ± 3 mm Hg) occurred about 2 h after drink ingestion and this fall was sustained thereafter (120-180 min: P₂₈₀ and M₅₀₄ vs. C $P < 0.05$). Maximum DBP decreases and HR increases occurred after M₅₀₄, with no differences between the effects of the P₂₈₀ and M₂₈₀ drinks.

Conclusions: The effects of whey-protein containing drinks to lower BP and

increase HR appear to be primarily dependent on their energy content rather than macronutrient composition and may persist for at least 3 h after ingestion. Pure whey-protein drinks may represent the best approach to maximize protein intake without increasing the potential for deleterious BP falls in older people.

9.2 Introduction

Older people are increasingly encouraged to take high protein nutritional supplements to reduce the age-associated loss of muscle mass and function (97). Whey protein is often part of these supplements, given that it is high in essential amino acids and appears to be effective in stimulating muscle protein formation (7).

Ingestion of nutrients can lead to postprandial reductions in blood pressure (BP), in older people (even when apparently healthy) (227, 396), in part, caused by postprandial diversion of blood to the gut, which can lead to syncope, falls and, in some cases, stroke or death (262, 409, 410). This so-called postprandial hypotension (PPH) has been defined as a fall in systolic blood pressure (SBP) greater than 20 mm Hg during the 2 h following nutrient ingestion i.e. it is assumed that the maximum fall in SBP occurs within 2 h. We have recently reported, in a cohort of healthy older men, that ingestion of a 70 g (280 kcal) whey-protein drink decreased SBP substantially when compared to a non-caloric control drink (264). In the majority of men in that study magnitude of the SBP decrease was greater than 20 mmHg after the 70 g whey protein drink (11/19 compared to 5/19 after the control drink) (262). Furthermore, the hypotensive effect of a whey protein drink was prolonged, with a sustained reduction in SBP being evident at 3 h after ingestion. It has been suggested that hypertensive men may be of particular risk of blood pressure falls following food intake (411). The current study aimed to determine in older men the effects of whey protein, when ingested in a lower quantity but with carbohydrate and fat as occurs frequently in a “real world” setting, on blood pressure and heart rate for 3 h. We hypothesized that the hypotensive effects of whey protein containing drinks would be dependent on the energy rather than protein content of the drink and often persist

for more than 2 h.

9.3 Materials and methods

9.3.1 Participants

Thirteen older men were recruited by advertisement. Participant characteristics are detailed in Table 9.1

Table 9.1: Participant characteristics

Age (years)	75 ± 2
Height (m)	1.75 ± 0.01
Weight (kg)	79 ± 2
BMI (kg/m ²)	25.6 ± 0.6

Mean and standard error of mean of 13 older men

Exclusion criteria were smoking; being vegetarian; alcohol intake of > 2 standard drinks on > 5 days per week; use of prescribed or non-prescribed medications which may affect appetite, body weight, gastrointestinal function or energy metabolism; intake of any illicit substance; known lactose intolerance or food allergy(s); epilepsy; gallbladder, pancreatic, cardiovascular or respiratory diseases; significant gastrointestinal symptoms including as abdominal pain, gastroesophageal reflux, diarrhea, or constipation or surgery; any other illness deemed significant by the investigator; low levels of plasma ferritin; blood donation in the previous 12 weeks; undernutrition (score < 24 on the Mini Nutritional Assessment (407); depression (score ≥ 11 on the Geriatric Depression Questionnaire (327); impaired cognitive function (score < 25 on Mini Mental State (326); or inability to comprehend the study. Anti-hypertensive medication were taken by four older men (anti-arrhythmic

n = 1; angiotensin converting enzyme inhibitor, n = 1; beta blockers, n = 1; angiotensin receptor blockers n = 1). Participants were instructed to not take medication on the morning of their study visit.

The study was conducted in accordance with the Declaration of Helsinki and Royal Adelaide Hospital Human Research Ethics Committee approved the protocol. The study was registered with the Australian New Zealand Clinical Trial Registry (www.anzctr.org.au, registration number ACTRN12614000846628). All participants provided written informed consent prior to their study inclusion.

9.3.2 Protocol

Each participant was studied on four occasions in a randomised, double-blind, placebo-controlled design (using randomly permuted blocks; www.randomization.com), separated by 3–14 days, to determine the effects of drinks (~ 450 mL) containing either: (i) 70 g whey protein (280 kcal; ‘P₂₈₀’); (ii) 14 g whey protein, 28 g carbohydrate, 12.4 g fat (280 kcal; ‘M₂₈₀’); (iii) 70 g whey protein, 28 g carbohydrate, 12.4 g fat (504 kcal; ‘M₅₀₄’); or (iv) an iso-palatable control drink (~ 2 kcal; ‘control’) on SBP, DBP and heart rate (HR). The BP and HR data are secondary outcomes of a published study which included results relating to energy intake, appetite, gastric emptying and plasma gut hormone concentrations (16). Sample size and power calculations for the original study were based on the primary outcomes of energy intake and gastric emptying (16). The drinks were prepared by homogenizing olive oil (Bertolli Australia Pty Ltd., Unilever Australasia, Sydney, NSW, Australia) and dissolving whey protein isolate (Fonterra Co-Operative Group Ltd., Palmerston North, New Zealand) and dextrose, in varying volumes of demineralized water and

diet lime cordial (Bickford's Australia Pty Ltd., Salisbury South, SA, Australia), to achieve the desired composition, on the morning of the study day, by a research officer, who was not involved in the data analysis. The drinks were stirred continuously at low speed on a stirring plate to ensure even mixing, were matched for taste and served in a covered cup. Both the investigator and the participant were blinded to the drink condition.

Participants were provided with a standardized meal [beef lasagne (McCain Foods Pty Ltd, Wendouree, Victoria VIC), Australia], ~591 kcal] to consume on the night before each study day at ~1900 h. They were instructed to fast overnight ~ 12 h from solids and liquids except water, and to refrain from strenuous physical activity until they attended the laboratory at ~0830 h.. The recruitment of participants started in March 2014, and the study was completed in December 2014.

9.3.3 Measurements

On arrival to the clinical research facility lab, level 4 at Adelaide Health and Medical Sciences, The University of Adelaide, Australia, participants were seated in an upright position where they remained throughout the study. An intravenous cannula was inserted for blood sampling. Participants sat quietly for 15 min then had 3 baseline measures of BP (Dinamap machine) and heart rate at 3-min intervals before drinking the test drink within 2 min. BP and HR were measured every 3 min until $t = 180$ min. Participants did not receive any other drink or food throughout the study.

9.4 Data and statistical analysis

Statistical analyses were performed using SPSS software (version 24; IBM, Armonk, NY). Drink-condition effects (control, P₂₈₀, M₂₈₀, M₅₀₄) were determined by using two way repeated-measures ANOVA. Significant effects were followed by Bonferroni corrected post-hoc tests to determine which specific drink conditions were different. Statistical significance was accepted at $P < 0.05$. All data are presented as means \pm SEMs. Baseline blood pressure represented as BL in the figures was calculated as an average of -9, -6 and -3 min readings. T = 0 min refers to the point immediately after drink consumption.

9.5 Results

The study protocol was well tolerated by all participants. No participant reported symptoms of dizziness, faintness or any other adverse events during the study.

9.5.1 Systolic blood pressure (SBP)

Baseline SBP values were not different on the four study days (Mean control: 128 ± 3 mmHg, P₂₈₀: 129 ± 3 mm Hg, M₂₈₀: 130 ± 5 mm Hg, M₅₀₄ 127 ± 4 mm Hg, $P = 0.95$). SBP did not change over the three hours after the control drink. SBP was lower after all three nutrient drinks compared to the control drink, particularly during the second (60-120 min $P = 0.018$) and third (120-180 min $P = 0.022$) hours, with the greatest reduction after the M₅₀₄ drink ($P = 0.008$) (Figure 9.1). SBP following M₅₀₄ ingestion was lower when compared to the P₂₈₀ drink between 0–60 min ($P = 0.044$). There was no significant difference between SBP readings after the P₂₈₀ and M₂₈₀ drinks ($P > 0.05$). Following drink ingestion, a decrease in SBP > 20 mm Hg occurred at some time in 3/13 for control, 7/13 for P₂₈₀, 6/13 for M₂₈₀, and 9/13 for M₅₀₄.

Maximal SBP decreases from baseline (control: -14 ± 2 mmHg, P₂₈₀: -22 ± 2 mmHg, M₂₈₀: -22 ± 4 mmHg, M₅₀₄: -24 ± 3 mmHg; $P = 0.11$) occurred about two hours after the drinks (time baseline to nadir SBP: control: 99 ± 16 min, P₂₈₀: 119 ± 13 min, M₂₈₀: 116 ± 12 min, M₅₀₄: 119 ± 15 min; $P = 0.86$) and was sustained thereafter following the nutrient drinks (average 120-180 min control: 128 ± 3 mmHg, P₂₈₀: 118 ± 2 mmHg, M₂₈₀: 120 ± 4 mmHg, M₅₀₄: 114 ± 3 mmHg).

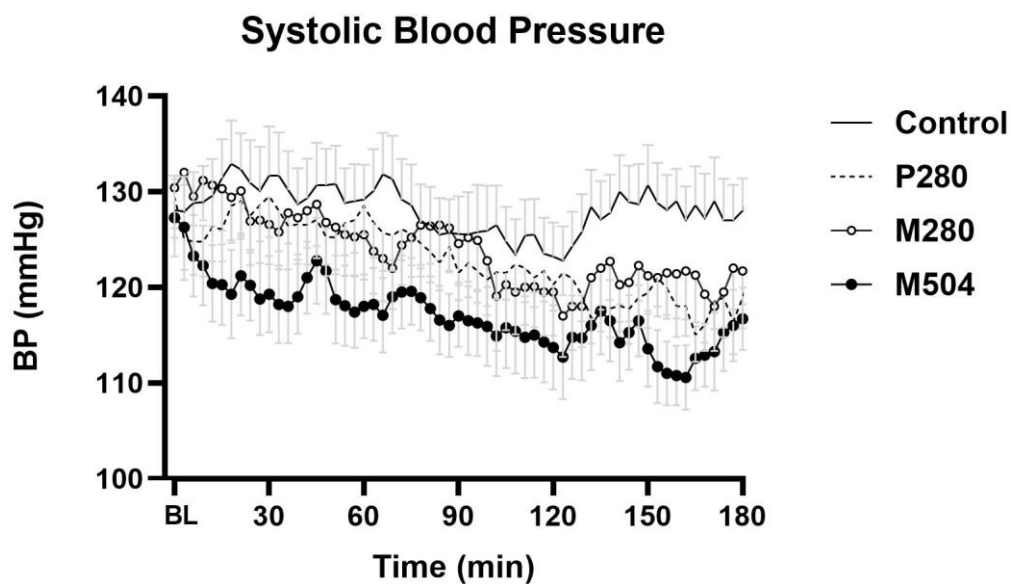


Figure 9.1: Mean (\pm SEM) systolic blood pressure (SBP; mmHg) following drink ingestion containing (i) flavored water (control, ~ 2 kcal) or (ii) 70 g whey protein (280 kcal; ‘P₂₈₀’); (iii) 14 g whey protein, 28 g carbohydrate, 12.4 g fat (280 kcal; ‘M₂₈₀’); (iv) 70 g protein, 28 g carbohydrate, 12.4 g fat (504 kcal; ‘M₅₀₄’) in older ($n = 13$) men. Drink-condition effects were determined by using repeated-measures ANOVA. Baseline blood pressure represented as BL in the figure was calculated as an average of -9, -6 and -3 min readings. SBP was lower after the M₅₀₄ drink when compared to control ($P = 0.019$) during the second (60-120 min $P = 0.035$) and third (120-180 min $P = 0.005$) hour

9.5.2 Diastolic blood pressure (DBP)

Baseline DBP values were not different on the four study days (Mean control: 74 ± 2 mmHg, P₂₈₀: 74 ± 2 mm Hg, M₂₈₀: 75 ± 3 mm Hg, M₅₀₄: 73 ± 2 mm Hg, $P = 0.94$).

DBP was lower after all three nutrient drinks compared to the control drink, with the greatest reduction after M504 (Figure 9.2).

DBP did not change over the three hours after the control drink. DBP was less after M₅₀₄ ($P < 0.001$) and P₂₈₀ ($P = 0.026$) when compared to control – the drink-condition effect was significant during all three hours following drink ingestion (0–60 min $P = 0.002$, 60–120 min $P = 0.004$, 120–180 min $P = 0.003$). There was no difference between the effects of M₂₈₀ and P₂₈₀ on DBP ($P > 0.05$). Maximal DBP decreases from baseline (control: -12 ± 2 mm Hg, P₂₈₀: -15 ± 1 mm Hg, M₂₈₀: -15 ± 2 mm Hg, M₅₀₄: -15 ± 1 mm Hg, $P = 0.17$) occurred on average between one to two hours after drink ingestion (time baseline to nadir: control: 80 ± 17 min, P₂₈₀: 108 ± 16 min, M₂₈₀: 71 ± 12 min, M₅₀₄: 99 ± 15 min, $P = 0.24$) and were sustained during the third hour for P₂₈₀ and M₅₀₄ (average control: 74 ± 1 mm Hg, P₂₈₀: 68 ± 1 mm Hg, M₂₈₀: 71 ± 1 mm Hg, M₅₀₄: 66 ± 1 mmHg)

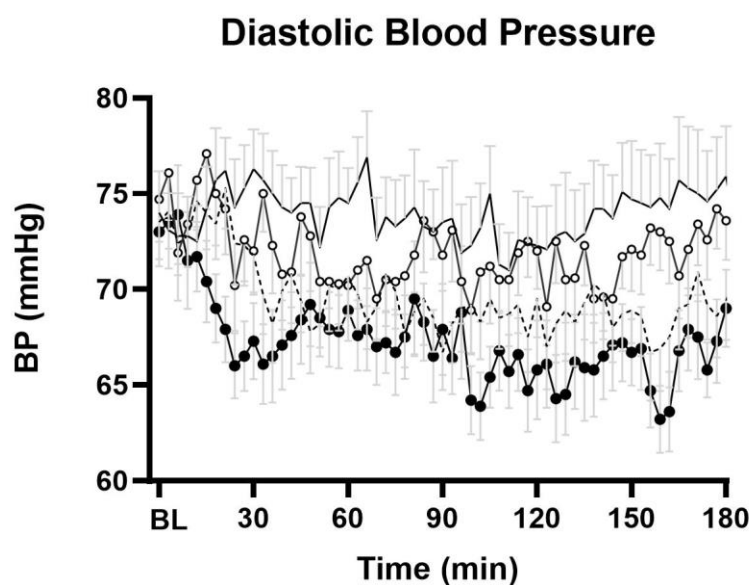


Figure 9.2: Mean (\pm SEM) systolic blood pressure (DBP; mmHg) following drink

ingestion containing (i) flavored water (control, ~ 2 kcal) or (ii) 70 g whey protein (280 kcal; 'P₂₈₀'); (iii) 14 g whey protein, 28 g carbohydrate, 12.4 g fat (280 kcal; 'M₂₈₀'); (iv) 70 g protein, 28 g carbohydrate, 12.4 g fat (504 kcal; 'M₅₀₄') in older (n = 13) men. Drink-condition effects were determined by using repeated-measures ANOVA. Baseline blood pressure represented as BL in the figure was calculated as an average of -9, -6 and -3 min readings. DBP was lower after the M₅₀₄ ($P < 0.001$) and P₂₈₀ ($P = 0.018$) drinks when compared to control during the second ($P = 0.012$) and third ($P = 0.035$) hour. There was no statistically significant difference between the effects of M₂₈₀ and P₂₈₀ on BP.

9.5.3 Heart rate (HR)

Baseline HR values were not different on the four study days (Mean control: 58 ± 2 bpm, P₂₈₀: 59 ± 3 bpm, M₂₈₀: 59 ± 2 bpm, M₅₀₄: 59 ± 3 bpm, $P = 0.95$). HR decreased over 3 h after the control drink and increased after the M₅₀₄ ($P < 0.001$) when compared to control, and did not change significantly after either the M₂₈₀ or P₂₈₀ drinks (0-180 min average control: 57 ± 1 bpm, P₂₈₀: 60 ± 1 bpm, M₂₈₀: 60 ± 2 bpm, M₅₀₄: 63 ± 1 bpm; Figure 9.3). The drink-condition effect was significant during all the three hours (0-60 min $P = 0.001$, 60-120 min $P < 0.001$, 120-180 min $P < 0.001$) hour following drink ingestion. Maximal HR increase from baseline (control: 3.8 ± 1 bpm, P₂₈₀: 9 ± 2 bpm, M₂₈₀: 12 ± 4 bpm, M₅₀₄: 13 ± 2 bpm, $P < 0.001$.) occurred on average between one to two hours after drink ingestion (time baseline to peak: control: 48 ± 14 min, P₂₈₀: 116 ± 15 min, M₂₈₀: 83 ± 14 min, M₅₀₄: 127 ± 16 min, $P = 0.003$) and were sustained during the third hour for P₂₈₀ and M₅₀₄ (average control: 55 ± 1 bpm, P₂₈₀: 61 ± 1 bpm, M₂₈₀: 60 ± 1 bpm, M₅₀₄: 64 ± 1 bpm).

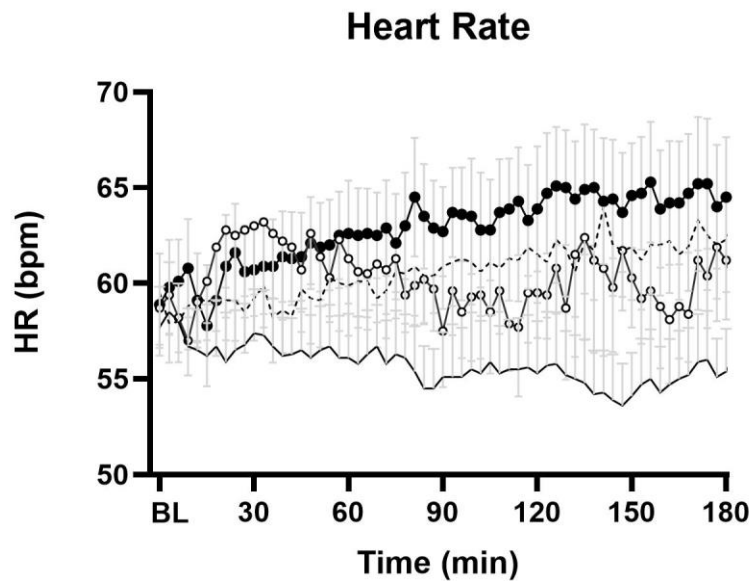


Figure 9.3: Mean (\pm SEM) Heart Rate (HR; bpm) following drink ingestion containing (i) flavored water (control, \sim 2 kcal) or (ii) 70 g whey protein (280 kcal; ‘P₂₈₀’); (iii) 14 g whey protein, 28 g carbohydrate, 12.4 g fat (280 kcal; ‘M₂₈₀’); (iv) 70 g protein, 28 g carbohydrate, 12.4 g fat (504 kcal; ‘M₅₀₄’) in older ($n = 13$) men. Drink-condition effects were determined by using repeated-measures ANOVA. Baseline Heart rate represented as BL in the figure was calculated as an average of -9, -6 and -3 min readings. HR increased after the M₅₀₄ ($P < 0.001$) and P₂₈₀ ($P = 0.017$) drinks when compared to control (0-180 min)

9.6 Discussion

The major observation in this study is that in healthy older men following ingestion of nutrient drinks containing 280 or 504 kcal energy as pure whey protein or as mixed macronutrients, the magnitude of the decrease in BP is dependent on the energy content, rather than, the protein content of the drinks. Furthermore, while the onset of the BP reduction was evident soon after nutrient drink ingestion, the hypotensive effect was sustained for at least 3 h. Both observations are of clinical relevance in a

'real world' setting. There is no advantage as far as the risk of postprandial hypotension by consuming protein with other macronutrients.

This is also consistent with the context that the rate of gastric emptying of nutrients, whether carbohydrate, fat or protein is primarily dependent on their caloric content.

Heart rate increased after ingestion of the nutrient drinks compared to the control drink, with the greatest and more sustained increase after the highest energy load drink. The resultant increase in cardiac output represents a compensatory mechanism for the potential postprandial fall in BP caused by diversion of blood to the gut. The nutrient-induced increases in HR were inadequate, however, to compensate fully for this diversion in these older men. We have reported smaller increases in HR and larger decreases in BP in healthy older than younger men after a pure 70 g whey protein drink (264) indicating that an age-related reduction in the ability to increase the heart rate after food ingestion contributes to the greater reduction in BP observed in older than younger adults after nutrient ingestion.

The nutrient drink-induced reduction in BP were substantial- a decrease of 20 mmHg SBP or more occurred at some time in 77% of the 504-kcal drink days, compared to 31% of the control drink days. None of the participants reported symptoms of dizziness or faintness, but they were seated throughout the study and not permitted to get up and walk around. A fall in systolic BP of 20 mmHg or more is clearly associated with symptoms and an increased likelihood of falls and injury, as reflected in one definition of postprandial hypotension (262). It is not known whether the risk of symptoms is influenced by the baseline systolic pressure and this should be evaluated. However, it should be recommended that current criteria for definition of

hypertension do not take into account the relationship of the blood pressure measurement to meal ingestion. Our observations suggest that caution should possibly be exercised for several hours (certainly more than two) at least in older people mobilising after nutrient containing drinks in relation to the risks of syncope and falls. Individuals at risk may potentially benefit from more frequent, lower calorie drinks or meals. Further studies should evaluate this strategy (176). The fall in BP may also be dependent on whether food is already present in the stomach and/or small intestine at the time a nutrient supplement is ingested.

Limitations of this study include that it was only conducted in men. While women tend to have a lower baseline BP that would intuitively predispose them lower BP levels after a meal, we and others have shown that the magnitude of the postprandial fall in BP is related directly to baseline BP such that the fall is greater in those who are hypertensive (229, 411). Further limitations include that the participants remained sitting to standardize study conditions and the number of participants was relatively small. Nevertheless, the observed decreases in BP and HR were clear cut. Because, the effects of the drinks on BP were still evident at 3 h, when each study ceased, it would be of relevance to determine the total duration of the postprandial hypotensive effect. Our study was designed to clarify phenomenology of 'real world' relevance rather than mechanisms. In relation to the later, evaluation of autonomic function would be of interest. We also only evaluated the acute effects of drinks, although we have no reason to believe that their effects on BP will be modified by chronic use.

Our observations suggest that if the intention is to give an older person a nutritional supplement drink containing as much whey protein as possible to preserve, or

potentially enhance, muscle mass and function, administering it in as pure a form as possible (i.e., with the minimum amount of fat and carbohydrate) may potentially reduce the BP the least and so minimize the risk of postprandial hypotension. If, on the other hand, the intention is to provide a mixed macronutrient supplement to provide a specific amount of energy, it should be appreciated the total energy content of the supplement, rather than its macronutrient composition will be the major determinant of its hypotensive effect.

9.7 Conclusions

The hypotensive effect of mixed macronutrient drinks is dependent on overall energy intake rather than macronutrient composition of a drink and may be sustained for at least 3 h. Pure whey protein drinks may, accordingly represent the best approach to maximise protein intake while minimising the potential for deleterious BP falls in older people.

CHAPTER 10: CONCLUSIONS OF THE THESIS

The results of the physiological studies presented in this thesis provide new knowledge about the short term effects in older people, obese and non-obese, and with and without type 2 diabetes mellitus, of whey protein in drink form, on feeding behaviour, glucose metabolism and post-ingestion blood pressure. Disturbances of these factors are common in older people and can have important effects on wellbeing and quality of life. The impact of whey and other increasingly used protein supplements on these factors is therefore of relevance and importance.

Obesity, T2D and postprandial blood pressure falls (leading to postprandial hypotension [PPH] when severe) are all prevalent in older people and associated with increased morbidity, mortality and health care costs. Existing and growing evidence indicates that nutrition-based strategies play an important role in the management of these conditions in older people. As outlined in chapter 2, because of an increasing awareness of weight loss, under-nutrition and sarcopaenia in older people protein supplements are gaining more attention and use in older people as a generally well tolerated and cost-effective way of preserving muscle mass and function. Whey protein appears to be a particularly effective protein for this purpose, due to its relatively high concentration of branched chain amino acids, particularly leucine.

The studies presented in this thesis have extended previous work by our group to now examine the effects on whey on feeding behaviours in older people over a longer time period than previously (chapter 3), in obese older people (chapter 4) on glucose metabolism in older people with and without T2D (chapter 5 and 5) and on blood pressure and pulse rate (chapters 7-9).

The main findings of these studies can be summarised as follows:

Chapter 3: Energy intake was suppressed by whey protein drinks in a protein load-responsive fashion at breakfast and particularly, at lunch, but not at dinner, and suppression of energy intake by protein was less in healthy older than younger men.

Chapter 4: The 30g whey protein drink did not suppress appetite or energy intake in obese younger or older men.

Chapter 5: The addition of 30g of whey protein to 30g of glucose substantially attenuated the increase in blood glucose concentrations induced by glucose alone; the magnitude was not affected by age. The stimulation of plasma insulin concentrations by whey protein was not reduced by ageing, unlike the insulin response to glucose. Whey protein suppressed hunger less in older than younger men. Glucagon concentrations were unaffected by age.

Chapter 6: The addition of 30 g of whey protein to 30 g of glucose substantially attenuated the increase in blood glucose concentrations induced by glucose alone; the magnitude was not affected by the presence of T2D and the stimulation of plasma insulin concentrations by whey protein.

Chapter 7: Older men exhibited a greater fall in SBP after whey-protein versus control than the younger men, with no BP change after the two drinks in younger men. The nadir in SBP occurred later in the older men, still declining 180 min after whey-protein ingestion. The magnitude of the rise in HR was greater in the younger men.

Chapter 8: Older men exhibited a decrease in SBP after ingestion of 30g and 70g of whey protein to a similar degree and greatest between 120 and 180 mins. HR was increased maximally after 70g particularly in the third hour and DBP decreased non significantly after protein drinks.

Chapter 9: SBP decreased after all three nutrient drinks with the greatest reduction after the M504. Maximal decreases in SBP occurred about 2 hour after drink ingestion and sustained thereafter. Maximum DBP decreases and HR increases occurred after M504, with no differences between the effects of the P280 and M280 drinks.

Collectively, the studies described in this thesis have produced clear-cut and exciting results. Overall the results of these studies suggest that whey protein has minimal suppressive effects on appetite and food intake in older obese and non-obese people, that the effects of a single dose of oral whey are relatively short-lived, that the effect of whey to lower blood glucose concentrations when co-ingested with carbohydrates persists in older people, both with and without type 2 diabetes. Whey protein, in a dose of 30gm, a dose likely to be effective in helping preserve muscle function in older people, lead to decreases in blood pressure, more in older people than young, and this could potentially cause adverse effects in predisposed older people, particularly if consumed with other macronutrients, as the BP lowering effect of the nutrient drinks studied is dependent on their total energy content rather than their macronutrient composition. These are novel findings that significantly add to the pre-existing body of literature.

Although each chapter has clearly mentioned its limitations, overall the studies presented in this thesis have several strengths and limitations. The participant numbers were relatively small, and most studies included only men, and therefore, we cannot comment on the effect of gender. We studied only men, as they appear to have the greatest ability to regulate energy intake in response to energy manipulation, and in women particularly the menstrual cycle may have a confounding effect on appetite and energy intake. We determined the effects of whey protein ingestion, a protein source which is frequently used in high energy supplements due to its high leucine content and ‘rapid’ digestion, when compared to other proteins such as casein. Whey, when compared to casein or soy, protein results in greater muscle protein synthesis in young and older men. It has to be determined whether our observations apply to other proteins. A strength of the studies was their randomised cross-over design in which the participants acted as their own control.

Further work will be required to determine if the short-term effects of whey found in the studies in this thesis persist with longer term use, with different doses and in different participant groups. Nevertheless, these results should provide guidance for further studies of whey and its use/ingestion by older people.

GLOSSARY

ACTRN; Australian New Zealand Clinical Trial Registry

ACE; Angiotension converting enzymes

ANOVA; Analysis of variance

AUC; Area under the curve

BP; Blood pressure

BMI; Body mass index

BPM; Beats per minute

CCK; Cholecystokinin

CI; Confidence interval

CT; Computerised tomography

CV; Coefficient of variation

DBP; Diastolic blood pressure

DPP-IV; Dipeptyl peptidase-4

EDTA; Ethylenediaminetetraacetic acid

ELISA, Enzyme-linked immunosorbent assay

GE; Gastric emptying

GIP;Ggastric inhibitory polypeptide/glucose-dependent insulintropic peptide

GLP-1; Glucagon-like peptide-1

HR; Heart rate

HbA1C; Glycosylated haemoglobin

IAAO; Indicator amino acid oxidation

iAUC; Incremental area under the curve

LLOQ; Lower limit of quantification

MRI; Magnetic Resonance Imaging

MPS; Muscle protein synthesis

NHANES; National health and nutrition examination survey

OC; Older men control

OP; Older men protein

PPH; Postprandial hypotension

PYY; Peptide tyrosine tyrosine

SBP; Systolic blood pressure

SD; Standard deviation

SEM; Standard error of the mean

SGLT-1; Sodium-glucose cotransporter-1

SGLT-2; Sodium-glucose cotransporter-2

2D ultrasound- Two dimensional ultrasound

3D ultrasound; Three dimensional ultrasound

T2D; Type 2 Diabetes

T₅₀; 50% gastric emptying time

VAS; Visual analogue scale

WHO; World health organisation

YC; Younger men control

YP; Younger men protein

REFERENCES

1. Chapman IM, MacIntosh CG, Morley JE, Horowitz M. The anorexia of ageing. *Biogerontology*. 2002;3(1-2):67-71.
2. Chapman IM. Nutritional disorders in the elderly. *Med Clin North Am*. 2006 Sep;90(5):887-907.
3. Deer RR, Volpi E. Protein intake and muscle function in older adults. *Curr Opin Clin Nutr Metab Care*. 2015 May;18(3):248-53.
4. Park Y, Choi J-E, Hwang H-S. Protein supplementation improves muscle mass and physical performance in undernourished prefrail and frail elderly subjects: a randomized, double-blind, placebo-controlled trial. *Am J Clin Nutr*. 2018 Nov;108(5):1026-33.
5. Katsanos CS, Chinkes DL, Paddon-Jones D, Zhang XJ, Aarsland A, Wolfe RR. Whey protein ingestion in elderly persons results in greater muscle protein accrual than ingestion of its constituent essential amino acid content. *Nutr Res*. 2008 Oct;28(10):651-8.
6. Breen L, Phillips SM. Skeletal muscle protein metabolism in the elderly: Interventions to counteract the 'anabolic resistance' of ageing. *Nutr Metab (Lond)*. 2011 Oct;8(1):68.
7. Pennings B, Boirie Y, Senden JM, Gijsen AP, Kuipers H, van Loon LJ. Whey protein stimulates postprandial muscle protein accretion more effectively than do casein and casein hydrolysate in older men. *Am J Clin Nutr*. 2011 May;93(5):997-1005.
8. Putra C, Konow N, Gage M, York CG, Mangano KM. Protein source and muscle health in older adults: a literature review. *Nutrients*. 2021 Feb;13(3):743.

9. Soenen S, Giezenaar C, Hutchison AT, Horowitz M, Chapman I, Luscombe-Marsh ND. Effects of intraduodenal protein on appetite, energy intake, and antropyloroduodenal motility in healthy older compared with young men in a randomized trial. *Am J Clin Nutr.* 2014 Oct;100(4):1108-15.
10. Giezenaar C, Chapman I, Luscombe-Marsh N, Feinle-Bisset C, Horowitz M, Soenen S. Ageing is associated with decreases in appetite and energy intake—a meta-analysis in healthy adults. *Nutrients.* 2016 Jan;8(1):28.
11. Giezenaar C, Lange K, Hausken T, Jones KL, Horowitz M, Chapman I, et al. Acute effects of substitution, and addition, of carbohydrates and fat to protein on gastric emptying, blood glucose, gut hormones, appetite, and energy intake. *Nutrients.* 2018 Oct;10(10):1451.
12. Giezenaar C, Lange K, Hausken T, Jones KL, Horowitz M, Chapman I, et al. Effects of age on acute appetite-related responses to whey-protein drinks, including energy intake, gastric emptying, blood glucose, and plasma gut hormone concentrations—a randomized controlled trial. *Nutrients.* 2020 Apr;12(4):1008.
13. Giezenaar C, Luscombe-Marsh ND, Hutchison AT, Standfield S, Feinle-Bisset C, Horowitz M, et al. Dose-dependent effects of randomized intraduodenal whey-protein loads on glucose, gut hormone, and amino acid concentrations in healthy older and younger men. *Nutrients.* 2018 Jan;10(1):78.
14. Giezenaar C, Trahair LG, Luscombe-Marsh ND, Hausken T, Standfield S, Jones KL, et al. Effects of randomized whey-protein loads on energy intake, appetite, gastric emptying, and plasma gut-hormone concentrations in older men and women. *Am J Clin Nutr.* 2017 Sep;106(3):865-77.

15. Giezenaar C, Trahair LG, Rigda R, Hutchison AT, Feinle-Bisset C, Luscombe-Marsh ND, et al. Lesser suppression of energy intake by orally ingested whey protein in healthy older men compared with young controls. *Am J Physiol Regul Integr Comp Physiol*. 2015 Oct;309(8):R845-54.
16. Giezenaar C, van der Burgh Y, Lange K, Hatzinikolas S, Hausken T, Jones KL, et al. Effects of substitution, and adding of carbohydrate and fat to whey-protein on energy intake, appetite, gastric emptying, glucose, insulin, ghrelin, cck and glp-1 in healthy older men-a randomized controlled trial. *Nutrients*. 2018 Jan;10(2):113.
17. Giezenaar C, Luscombe-Marsh ND, Hutchison AT, Lange K, Hausken T, Jones KL, et al. Effect of gender on the acute effects of whey protein ingestion on energy intake, appetite, gastric emptying and gut hormone responses in healthy young adults. *Nutr Diabetes*. 2018 Jul;8(1):40.
18. Hutchison AT, Piscitelli D, Horowitz M, Jones KL, Clifton PM, Standfield S, et al. Acute load-dependent effects of oral whey protein on gastric emptying, gut hormone release, glycemia, appetite, and energy intake in healthy men. *Am J Clin Nutr*. 2015;102(6):1574-84.
19. United Nations. The world population situation in 2014: a concise report. New York: United Nations;2014 [cited 2022 Sep 18]. Available from: https://www.un.org/en/development/desa/population/events/pdf/other/4/World%20Population%20Situation_2014_10%20key%20findings_en.pdf.
20. United Nations. World population ageing 2009. New York: United Nations; 2009 [cited 2022 Sep 18];1-46. Available from: <https://www.un.org/en/development/desa/population/publications/pdf/ageing/WorldPopulationAgeing2019-Highlights.pdf>.

21. United Nations. Population facts: population ageing and sustainable development 2014. New York: [Internet]. United Nations; 2014 [cited 2022 Sep 18]. Available from: https://www.un.org/development/desa/pd/sites/www.un.org.development.desa.pd/files/files/documents/2020/Jan/un_2017_factsheet1.pdf.
22. Australian Institute of Health and Welfare. Older Australia at a glance. Canberra: AIHW; 2018 [cited 2022 Aug 22]. 1-77p. Available from: <https://www.aihw.gov.au/reports/older-people/older-australia-at-a-glance>.
23. Harris A, Sharma A. Estimating the future health and aged care expenditure in Australia with changes in morbidity. *PLoS One*. 2018 Aug;13(8):e0201697.
24. Calvani R, Picca A, Marzetti E. Anorexia of aging. In: Gu D, Dupre ME, editors. *Encyclopedia of gerontology and population aging*. Cham: Springer International Publishing; 2019. p. 1-7.
25. Sturm K, Parker B, Wishart J, Feinle-Bisset C, Jones KL, Chapman I, et al. Energy intake and appetite are related to antral area in healthy young and older subjects. *Am J Clin Nutr*. 2004 Sep;80(3):656-67.
26. Sturm K, MacIntosh CG, Parker BA, Wishart J, Horowitz M, Chapman IM. Appetite, food intake, and plasma concentrations of cholecystokinin, ghrelin, and other gastrointestinal hormones in undernourished older women and well-nourished young and older women. *J Clin Endocrinol Metab*. 2003 Aug;88(8):3747-55.
27. Cook CG, Andrews JM, Jones KL, Wittert GA, Chapman IM, Morley JE, et al. Effects of small intestinal nutrient infusion on appetite and pyloric motility are modified by age. *Am J Physiol*. 1997 Aug;273(2 Pt 2):R755-61.

28. MacIntosh CG, Horowitz M, Verhagen MA, Smout AJ, Wishart J, Morris H, et al. Effect of small intestinal nutrient infusion on appetite, gastrointestinal hormone release, and gastric myoelectrical activity in young and older men. *Am J Gastroenterol*. 2001 Apr;96(4):997-1007.
29. MacIntosh CG, Andrews JM, Jones KL, Wishart JM, Morris HA, Jansen JB, et al. Effects of age on concentrations of plasma cholecystokinin, glucagon-like peptide 1, and peptide YY and their relation to appetite and pyloric motility. *Am J Clin Nutr*. 1999;69(5):999-1006.
30. Wurtman JJ, Lieberman H, Tsay R, Nader T, Chew B. Calorie and nutrient intakes of elderly and young subjects measured under identical conditions. *J Gerontol*. 1988 Nov;43(6):B174-80.
31. Bhasin S, Woodhouse L, Casaburi R, Singh AB, Mac RP, Lee M, et al. Older men are as responsive as young men to the anabolic effects of graded doses of testosterone on the skeletal muscle. *J Clin Endocrinol Metab*. 2005 Feb;90(2):678-88.
32. Visvanathan R, Chapman IM. Undernutrition and anorexia in the older person. *Gastroenterol Clin North Am*. 2009 Sep;38(3):393-409.
33. Norman K, Haß U, Pirlich M. Malnutrition in older adults—recent advances and remaining challenges. *Nutrients*. 2021 Aug;13(8):2764.
34. Newman AB, Yanez D, Harris T, Duxbury A, Enright PL, Fried LP. Weight change in old age and its association with mortality. *J Am Geriatr Soc*. 2001 Oct;49(10):1309-18.
35. Schoenborn CA, Adams PF, Barnes PM. Body weight status of adults: United States, 1997-98. *Adv Data*. 2002 Sep;6(330):1-15.

36. Wallace JI, Schwartz RS, LaCroix AZ, Uhlmann RF, Pearlman RA. Involuntary weight loss in older outpatients: incidence and clinical significance. *J Am Geriatr Soc.* 1995 Apr;43(4):329-37.
37. Barone L, Milosavljevic M, Gazibarich B. Assessing the older person: is the MNA a more appropriate nutritional assessment tool than the SGA? *J Nutr Health Aging.* 2003;7(1):13-7.
38. de Groot L, Beck AM, Schroll M, van Staveren WA. Evaluating the DETERMINE Your Nutritional Health Checklist and the Mini Nutritional Assessment as tools to identify nutritional problems in elderly Europeans. *Eur J Clin Nutr.* 1998 Dec;52(12):877-83.
39. Persson MD, Brismar KE, Katzarski KS, Nordenström J, Cederholm TE. Nutritional status using mini nutritional assessment and subjective global assessment predict mortality in geriatric patients. *J Am Geriatr Soc.* 2002 Dec;50(12):1996-2002.
40. Soini H, Routasalo P, Lagström H. Characteristics of the Mini-Nutritional Assessment in elderly home-care patients. *Eur J Clin Nutr.* 2004 Jan;58(1):64-70.
41. Visvanathan R, Macintosh C, Callary M, Penhall R, Horowitz M, Chapman I. The nutritional status of 250 older Australian recipients of domiciliary care services and its association with outcomes at 12 months. *J Am Geriatr Soc.* 2003 Jul;51(7):1007-11.
42. Stajkovic S, Aitken EM, Holroyd-Leduc J. Unintentional weight loss in older adults. *CMAJ.* 2011 Mar;183(4):443-9.
43. Somes GW, Kritchevsky SB, Shorr RI, Pahor M, Applegate WB. Body mass index, weight change, and death in older adults: the systolic hypertension in the elderly program. *Am J Epidemiol.* 2002 Jul;156(2):132-8.

44. Nilsson M, Stenberg M, Frid AH, Holst JJ, Björck IM. Glycemia and insulinemia in healthy subjects after lactose-equivalent meals of milk and other food proteins: the role of plasma amino acids and incretins. *Am J Clin Nutr.* 2004 Nov;80(5):1246-53.
45. Holloszy JO. The biology of aging. *Mayo Clin Proc.* 2000 Jan;75 Suppl:S3-8; discussion S-9.
46. Melton LJ, 3rd, Khosla S, Crowson CS, O'Connor MK, O'Fallon WM, Riggs BL. Epidemiology of sarcopenia. *J Am Geriatr Soc.* 2000 Jun;48(6):625-30.
47. Fry CS, Rasmussen BB. Skeletal muscle protein balance and metabolism in the elderly. *Curr Aging Sci.* 2011 Dec;4(3):260-8.
48. Volpi E, Nazemi R, Fujita S. Muscle tissue changes with aging. *Curr Opin Clin Nutr Metab Care.* 2004 Jul;7(4):405-10.
49. Santilli V, Bernetti A, Mangone M, Paoloni M. Clinical definition of sarcopenia. *Clin Cases Miner Bone Metab.* 2014 Sep;11(3):177-80.
50. Greco EA, Pietschmann P, Migliaccio S. Osteoporosis and sarcopenia increase frailty syndrome in the elderly. *Front Endocrinol (Lausanne).* 2019;10:255.
51. Visvanathan R, Chapman I. Preventing sarcopaenia in older people. *Maturitas.* 2010 Aug;66(4):383-8.
52. Hida T, Harada A, Imagama S, Ishiguro N. Managing sarcopenia and its related-fractures to improve quality of life in geriatric populations. *Aging Dis.* 2014 Aug;5(4):226-37.
53. Walston JD. Sarcopenia in older adults. *Curr Opin Rheumatol.* 2012 Nov;24(6):623-7.
54. Distefano G, Goodpaster BH. Effects of exercise and aging on skeletal muscle. *Cold Spring Harb Perspect Med.* 2018 Mar;8(3).

55. Yoshimura Y, Wakabayashi H, Yamada M, Kim H, Harada A, Arai H. Interventions for treating sarcopenia: a systematic review and meta-analysis of randomized controlled studies. *J Am Med Dir Assoc*. 2017 Jun;18(6):553.e1-.e16.
56. Ng TP, Feng L, Nyunt MSZ, Feng L, Niti M, Tan BY, et al. Nutritional, physical, cognitive, and combination interventions and frailty reversal among older adults: a randomized controlled trial. *Am J Med*. 2015 Nov;128(11):1225-36.e1.
57. Bernstein M, Munoz N. Position of the Academy of Nutrition and Dietetics: food and nutrition for older adults: promoting health and wellness. *J Acad Nutr Diet*. 2012 Aug;112(8):1255-77.
58. Roberts S, Collins P, Rattray M. Identifying and managing malnutrition, frailty and sarcopenia in the community: a narrative review. *Nutrients*. 2021 Jul;13(7):2316.
59. Leggo M, Banks M, Isenring E, Stewart L, Tweeddale M. A quality improvement nutrition screening and intervention program available to Home and Community Care eligible clients. *Nutr Diet*. 2008 Jun;65(2):162-7.
60. Hamirudin AH, Walton K, Charlton K, Carrie A, Tapsell L, Milosavljevic M, et al. Feasibility of home-based dietetic intervention to improve the nutritional status of older adults post-hospital discharge. *Nutr Diet*. 2017 Jul;74(3):217-23.
61. Charlton KE, Walton K, Moon L, Smith K, McMahon AT, Ralph F, et al. "It could probably help someone else but not me": a feasibility study of a snack programme offered to meals on wheels clients. *J Nutr Health Aging*. 2013 Apr;17(4):364-9.

62. Cameron ID, Fairhall N, Langron C, Lockwood K, Monaghan N, Aggar C, et al. A multifactorial interdisciplinary intervention reduces frailty in older people: randomized trial. *BMC Med.* 2013 Mar;11(1):65.
63. Chapman I, Oberoi A, Giezenaar C, Soenen S. Rational use of protein supplements in the elderly-relevance of gastrointestinal mechanisms. *Nutrients.* 2021 Apr;13(4):1227.
64. Apolzan JW, Carnell NS, Mattes RD, Campbell WW. Inadequate dietary protein increases hunger and desire to eat in younger and older men. *J Nutr.* 2007 Jun;137(6):1478-82.
65. Aldrich ND, Reicks MM, Sibley SD, Redmon JB, Thomas W, Raatz SK. Varying protein source and quantity do not significantly improve weight loss, fat loss, or satiety in reduced energy diets among midlife adults. *Nutr Res.* 2011 Feb;31(2):104-12.
66. Gannon MC, Nuttall FQ, Saeed A, Jordan K, Hoover H. An increase in dietary protein improves the blood glucose response in persons with type 2 diabetes. *Am J Clin Nutr.* 2003 Oct;78(4):734-41.
67. Abete I, Astrup A, Martínez JA, Thorsdottir I, Zulet MA. Obesity and the metabolic syndrome: role of different dietary macronutrient distribution patterns and specific nutritional components on weight loss and maintenance. *Nutr Rev.* 2010 Apr;68(4):214-31.
68. Nuttall FQ, Mooradian AD, Gannon MC, Billington C, Krezowski P. Effect of protein ingestion on the glucose and insulin response to a standardized oral glucose load. *Diabetes Care.* 1984 Sep-Oct;7(5):465-70.
69. Nuttall FQ, Gannon MC, Wald JL, Ahmed M. Plasma glucose and insulin profiles in normal subjects ingesting diets of varying carbohydrate, fat, and protein content. *J Am Coll Nutr.* 1985;4(4):437-50.

70. Soenen S, Rayner CK, Horowitz M, Jones KL. Gastric emptying in the elderly. *Clin Geriatr Med*. 2015 Aug;31(3):339-53.
71. Soenen S, Chapman IM. Body weight, anorexia, and undernutrition in older people. *J Am Med Dir Assoc*. 2013 Sep;14(9):642-8.
72. Rolls BJ, Dimeo KA, Shide DJ. Age-related impairments in the regulation of food intake. *Am J Clin Nutr*. 1995 Nov;62(5):923-31.
73. Westerterp-Plantenga MS, Lemmens SG, Westerterp KR. Dietary protein - its role in satiety, energetics, weight loss and health. *Br J Nutr*. 2012 Aug;108 Suppl 2:S105-12.
74. Clifton P. Effects of a high protein diet on body weight and comorbidities associated with obesity. *Br J Nutr*. 2012 Aug;108 Suppl 2:S122-9.
75. Te Morenga L, Mann J. The role of high-protein diets in body weight management and health. *Br J Nutr*. 2012 Aug;108 Suppl 2:S130-8.
76. Dong JY, Zhang ZL, Wang PY, Qin LQ. Effects of high-protein diets on body weight, glycaemic control, blood lipids and blood pressure in type 2 diabetes: meta-analysis of randomised controlled trials. *Br J Nutr*. 2013 Sep;110(5):781-9.
77. Poppitt SD, McCormack D, Buffenstein R. Short-term effects of macronutrient preloads on appetite and energy intake in lean women. *Physiol Behav*. 1998 Jun;64(3):279-85.
78. Leidy HJ, Clifton PM, Astrup A, Wycherley TP, Westerterp-Plantenga MS, Luscombe-Marsh ND, et al. The role of protein in weight loss and maintenance. *Am J Clin Nutr*. 2015 Jun;101(6):1320s-9s.
79. Weigle DS, Breen PA, Matthys CC, Callahan HS, Meeuws KE, Burden VR, et al. A high-protein diet induces sustained reductions in appetite, ad libitum caloric intake, and body weight despite compensatory changes in diurnal

- plasma leptin and ghrelin concentrations. *Am J Clin Nutr*. 2005 Jul;82(1):41-8.
80. McAuley KA, Hopkins CM, Smith KJ, McLay RT, Williams SM, Taylor RW, et al. Comparison of high-fat and high-protein diets with a high-carbohydrate diet in insulin-resistant obese women. *Diabetologia*. 2005 Jan;48(1):8-16.
81. Burd NA, Gorissen SH, van Loon LJ. Anabolic resistance of muscle protein synthesis with aging. *Exerc Sport Sci Rev*. 2013 Jul;41(3):169-73.
82. Baum JI, Kim IY, Wolfe RR. Protein consumption and the elderly: what is the optimal level of intake? *Nutrients*. 2016 Jun;8(6):359.
83. Nowson C, O'Connell S. Protein requirements and recommendations for older people: a review. *Nutrients*. 2015 Aug;7(8):6874-99.
84. Loenneke JP, Loprinzi PD, Murphy CH, Phillips SM. Per meal dose and frequency of protein consumption is associated with lean mass and muscle performance. *Clin Nutr*. 2016 Dec;35(6):1506-11.
85. Thorning TK, Raben A, Tholstrup T, Soedamah-Muthu SS, Givens I, Astrup A. Milk and dairy products: good or bad for human health? An assessment of the totality of scientific evidence. *Food Nutr Res [Internet]*. 2016 Nov 22 [cited 2022 Aug 17];60:32527. Available from: <https://pubmed.ncbi.nlm.nih.gov/27882862/>.
86. Adams RL, Broughton KS. Insulinotropic effects of whey: mechanisms of action, recent clinical trials, and clinical applications. *Ann Nutr Metab*. 2016 Aug;69(1):56-63.
87. Haraguchi FK, Pedrosa ML, Paula H, Santos RC, Silva ME. Evaluation of biological and biochemical quality of whey protein. *J Med Food*. 2010 Dec;13(6):1505-9.

88. Hoffman JR, Falvo MJ. Protein - which is best? *J Sports Sci Med*. 2004 Sep;3(3):118-30.
89. Sukkar SG, Vaccaro A, Ravera GB, Borrini C, Gradaschi R, Massa Sacchi-Nemours A, et al. Appetite control and gastrointestinal hormonal behavior (CCK, GLP-1, PYY 1-36) following low doses of a whey protein-rich nutraceutical. *Med J Nutrition Metab*. 2013 Feb;6(3):259-66.
90. Giezenaar C, Hutchison AT, Luscombe-Marsh ND, Chapman I, Horowitz M, Soenen S. Effect of age on blood glucose and plasma insulin, glucagon, ghrelin, cck, gip, and glp-1 responses to whey protein ingestion. *Nutrients*. 2017 Dec;10(1):2.
91. Rayner CK, MacIntosh CG, Chapman IM, Morley JE, Horowitz M. Effects of age on proximal gastric motor and sensory function. *Scand J Gastroenterol*. 2000 Oct;35(10):1041-7.
92. Wolfe RR. Branched-chain amino acids and muscle protein synthesis in humans: myth or reality? *J Int Soc Sports Nutr*. 2017 Aug;14(1):30.
93. Gorissen SHM, Crombag JJR, Senden JMG, Waterval WAH, Bierau J, Verdijk LB, et al. Protein content and amino acid composition of commercially available plant-based protein isolates. *Amino Acids*. 2018 Dec;50(12):1685-95.
94. Katsanos CS, Kobayashi H, Sheffield-Moore M, Aarsland A, Wolfe RR. A high proportion of leucine is required for optimal stimulation of the rate of muscle protein synthesis by essential amino acids in the elderly. *Am J Physiol Endocrinol Metab*. 2006 Aug;291(2):E381-7.
95. Mignone LE, Wu T, Horowitz M, Rayner CK. Whey protein: The "whey" forward for treatment of type 2 diabetes? *World J Diabetes*. 2015 Oct;6(14):1274-84.

96. Camargo LDR, Doneda D, Oliveira VR. Whey protein ingestion in elderly diet and the association with physical, performance and clinical outcomes. *Exp Gerontol.* 2020 Aug;137:110936.
97. Bauer J, Biolo G, Cederholm T, Cesari M, Cruz-Jentoft AJ, Morley JE, et al. Evidence-based recommendations for optimal dietary protein intake in older people: a position paper from the PROT-AGE Study Group. *J Am Med Dir Assoc.* 2013 Aug;14(8):542-59.
98. Giezenaar C, Coudert Z, Baqeri A, Jensen C, Hausken T, Horowitz M, et al. Effects of timing of whey protein intake on appetite and energy intake in healthy older men. *J Am Med Dir Assoc.* 2017 Oct;18(10):898 e9-e13.
99. Hruby A, Hu FB. The epidemiology of obesity: a big picture. *Pharmacoeconomics.* 2015 Jul;33(7):673-89.
100. Seidell JC, Halberstadt J. The global burden of obesity and the challenges of prevention. *Ann Nutr Metab.* 2015;66(Suppl. 2):7-12.
101. Okunogbe A, Nugent R, Spencer G, Ralston J, Wilding J. Economic impacts of overweight and obesity: current and future estimates for eight countries. *BMJ Global Health.* 2021 Oct;6(10):e006351.
102. Keramat SA, Alam K, Al-Hanawi MK, Gow J, Biddle SJH, Hashmi R. Trends in the prevalence of adult overweight and obesity in Australia, and its association with geographic remoteness. *Sci Rep.* 2021 May;11(1):11320.
103. Australian Institute of Health and Welfare. Overweight and obesity [Internet]. AIHW, Australian Government, 2022 Jul 07 [cited 2022 29 Aug]. Available from: <https://www.aihw.gov.au/reports/australias-health/overweight-and-obesity>.

104. Leitner DR, Frühbeck G, Yumuk V, Schindler K, Micic D, Woodward E, et al. Obesity and type 2 diabetes: two diseases with a need for combined treatment strategies - easo can lead the way. *Obes Facts*. 2017;10(5):483-92.
105. Pi-Sunyer X. The medical risks of obesity. *Postgrad Med*. 2009 Nov;121(6):21-33.
106. DeFronzo RA, Abdul-Ghani MA. Preservation of β -cell function: the key to diabetes prevention. *J Clin Endocrinol Metab*. 2011 Aug;96(8):2354-66.
107. Kalra S. Diabesity. *J Pak Med Assoc*. 2013 Apr;63(4):532-4.
108. Finkelstein EA, Khavjou OA, Thompson H, Trogdon JG, Pan L, Sherry B, et al. Obesity and severe obesity forecasts through 2030. *Am J Prev Med*. 2012 Jun;42(6):563-70.
109. Smyth S, Heron A. Diabetes and obesity: the twin epidemics. *Nat Med*. 2006 Jan;12(1):75-80.
110. Newman A. Obesity in older adults. *Online J Issues Nurs*. 2009 Jan;14(1):3.
111. Batsis JA. Obesity in the older adult: special issue. *J Nutr Gerontol Geriatr*. 2019 Jan-Mar;38(1):1-5.
112. World Health Organization. Obesity and overweight [Internet]. World Health Organization; 2021 Jun 09 [cited 2002 Aug 29]. Available from: <https://www.who.int/news-room/fact-sheets/detail/obesity-and-overweight>.
113. Stenholm S, Harris TB, Rantanen T, Visser M, Kritchevsky SB, Ferrucci L. Sarcopenic obesity: definition, cause and consequences. *Curr Opin Clin Nutr Metab Care*. 2008 Nov;11(6):693-700.
114. Baumgartner RN. Body composition in healthy aging. *Ann N Y Acad Sci*. 2000 May;904:437-48.

115. Koliaki C, Liatis S, Dalamaga M, Kokkinos A. Sarcopenic obesity: epidemiologic evidence, pathophysiology, and therapeutic perspectives. *Curr Obes Rep.* 2019 Dec;8(4):458-71.
116. Bouchonville MF, Villareal DT. Sarcopenic obesity: how do we treat it? *Curr Opin Endocrinol Diabetes Obes.* 2013 Oct;20(5):412-9.
117. Choi KM. Sarcopenia and sarcopenic obesity. *Korean J Intern Med.* 2016 Nov;31(6):1054-60.
118. Roh E, Choi KM. Health consequences of sarcopenic obesity: a narrative review. *Front Endocrinol (Lausanne).* 2020;11:332.
119. Wadden TA, Webb VL, Moran CH, Bailer BA. Lifestyle modification for obesity: new developments in diet, physical activity, and behavior therapy. *Circulation.* 2012 Mar;125(9):1157-70.
120. Ikramuddin S, Korner J, Lee WJ, Thomas AJ, Connett JE, Bantle JP, et al. Lifestyle intervention and medical management with vs without Roux-en-Y gastric bypass and control of hemoglobin A1c, LDL cholesterol, and systolic blood pressure at 5 years in the diabetes surgery study. *JAMA.* 2018 Jan;319(3):266-78.
121. Cox CE. Role of physical activity for weight loss and weight maintenance. *Diabetes Spectr.* 2017 Aug;30(3):157-60.
122. Heymsfield SB, Gonzalez MC, Shen W, Redman L, Thomas D. Weight loss composition is one-fourth fat-free mass: a critical review and critique of this widely cited rule. *Obes Rev.* 2014 Apr;15(4):310-21.
123. Paddon-Jones D, Leidy H. Dietary protein and muscle in older persons. *Curr Opin Clin Nutr Metab Care.* 2014 Jan;17(1):5-11.
124. McGregor RA, Cameron-Smith D, Poppitt SD. It is not just muscle mass: a review of muscle quality, composition and metabolism during ageing as

- determinants of muscle function and mobility in later life. *Longev Healthspan*. 2014 Dec;3(1):9.
125. Australian Bureau of Statistics. Diabetes [Internet]. Canberra: ABS; 2020-21; 2022 Mar 21 [cited 2022 May 20]. Available from: <https://www.abs.gov.au/statistics/health/health-conditions-and-risks/diabetes/latest-release>
 126. Ogurtsova K, da Rocha Fernandes JD, Huang Y, Linnenkamp U, Guariguata L, Cho NH, et al. IDF Diabetes Atlas: global estimates for the prevalence of diabetes for 2015 and 2040. *Diabetes Res Clin Pract*. 2017 Jun;128:40-50.
 127. Bradley D, Hsueh W. Type 2 diabetes in the elderly: challenges in a unique patient population. *J Geriatr Med Gerontol*. 2016;2(2):14.
 128. Sousa GTD, Lira FS, Rosa JC, de Oliveira EP, Oyama LM, Santos RV, et al. Dietary whey protein lessens several risk factors for metabolic diseases: a review. *Lipids Health Dis*. 2012 Jul;11:67.
 129. Yakaryılmaz FD, Öztürk ZA. Treatment of type 2 diabetes mellitus in the elderly. *World J Diabetes*. 2017 Jun 15;8(6):278-85.
 130. Khattab M, Khader YS, Al-Khawaldeh A, Ajlouni K. Factors associated with poor glycemic control among patients with type 2 diabetes. *J Diabetes Complications*. 2010 Mar-Apr;24(2):84-9.
 131. Marín-Peñalver JJ, Martín-Timón I, Sevillano-Collantes C, Del Cañizo-Gómez FJ. Update on the treatment of type 2 diabetes mellitus. *World J Diabetes*. 2016 Sep 15;7(17):354-95.
 132. Colagiuri S DS, Girgis S, Colagiuri R. National evidence based guideline for blood glucose control in type 2 diabetes. Canberra: Diabetes Australia and the NHMRC; 2009 [cited 2022 Jan 17]; 1-290. Available from: <https://www.diabetesaustralia.com.au/wp-content/uploads/National->

Evidence-Based-Guideline-for-Blood-Glucose-Control-in-Type-2-Diabetes.pdf.

133. Phillips SJ, Arnesen SJ, Love CB. The language of disease outbreaks, disasters, and public health emergencies: the role of the US National Library of Medicine. *Disaster Med Public Health Prep.* 2014;8(2):112-3.
134. Franz MJ. Protein: metabolism and effect on blood glucose levels. *Diabetes Educ.* 1997 Nov-Dec;23(6): 643-6, 648, 650-1.
135. Oberoi A, Giezenaar C, Rigda RS, Lange K, Horowitz M, Jones KL, et al. Comparative effects of co-ingesting whey protein and glucose alone and combined on blood glucose, plasma insulin and glucagon concentrations in younger and older men. *Nutrients.* 2022 Jul;14(15):3111.
136. Frid AH, Nilsson M, Holst JJ, Björck IM. Effect of whey on blood glucose and insulin responses to composite breakfast and lunch meals in type 2 diabetic subjects. *Am J Clin Nutr.* 2005 Jul;82(1):69-75.
137. Jakubowicz D, Froy O. Biochemical and metabolic mechanisms by which dietary whey protein may combat obesity and Type 2 diabetes. *J Nutr Biochem.* 2013 Jan;24(1):1-5.
138. Pal S, Ellis V, Dhaliwal S. Effects of whey protein isolate on body composition, lipids, insulin and glucose in overweight and obese individuals. *Br J Nutr.* 2010 Sep;104(5):716-23.
139. Baggio LL, Drucker DJ. Biology of incretins: GLP-1 and GIP. *Gastroenterology.* 2007 May;132(6):2131-57.
140. Woods SC, Lutz TA, Geary N, Langhans W. Pancreatic signals controlling food intake; insulin, glucagon and amylin. *Philos Trans R Soc Lond B Biol Sci.* 2006 Jul 29;361(1471):1219-35.

141. Smith K, Bowden Davies KA, Stevenson EJ, West DJ. The clinical application of mealtime whey protein for the treatment of postprandial hyperglycaemia for people with type 2 diabetes: a long whey to go. *Front Nutr.* 2020 Oct;7.
142. Mann E, Sunni M, Bellin, M.D. Secretion of insulin in response to diet and hormones. *The Pancreapedia: Exocrine Pancreas Knowl Base.* 2020.
143. Manders RJ, Wagenmakers AJ, Koopman R, Zorenc AH, Menheere PP, Schaper NC, et al. Co-ingestion of a protein hydrolysate and amino acid mixture with carbohydrate improves plasma glucose disposal in patients with type 2 diabetes. *Am J Clin Nutr.* 2005 Jul;82(1):76-83.
144. Manders RJ, Hansen D, Zorenc AH, Dendale P, Kloek J, Saris WH, et al. Protein co-ingestion strongly increases postprandial insulin secretion in type 2 diabetes patients. *J Med Food.* 2014 Jul;17(7):758-63.
145. Estrich D, Ravnik A, Schlierf G, Fukayama G, Kinsell L. Effects of co-ingestion of fat and protein upon carbohydrate-induced hyperglycemia. *Diabetes.* 1967 Apr;16(4):232-7.
146. Rabinowitz D, Merimee TJ, Maffezzoli R, Burgess JA. Patterns of hormonal release after glucose, protein, and glucose plus protein. *Lancet.* 1966 Aug;2(7461):454-6.
147. Ma J, Stevens JE, Cukier K, Maddox AF, Wishart JM, Jones KL, et al. Effects of a protein preload on gastric emptying, glycemia, and gut hormones after a carbohydrate meal in diet-controlled type 2 diabetes. *Diabetes Care.* 2009 Sep;32(9):1600-2.
148. Mortensen LS, Hartvigsen ML, Brader LJ, Astrup A, Schrezenmeir J, Holst JJ, et al. Differential effects of protein quality on postprandial lipemia in

- response to a fat-rich meal in type 2 diabetes: comparison of whey, casein, gluten, and cod protein. *Am J Clin Nutr.* 2009 Jul;90(1):41-8.
149. Ma J, Jesudason DR, Stevens JE, Keogh JB, Jones KL, Clifton PM, et al. Sustained effects of a protein 'preload' on glycaemia and gastric emptying over 4 weeks in patients with type 2 diabetes: a randomized clinical trial. *Diabetes Res Clin Pract.* 2015 May;108(2):e31-4.
150. Paoli A, Tinsley G, Bianco A, Moro T. The influence of meal frequency and timing on health in humans: the role of fasting. *Nutrients.* 2019 Mar 28;11(4).
151. Koffert J, Honka H, Teuho J, Kauhanen S, Hurme S, Parkkola R, et al. Effects of meal and incretins in the regulation of splanchnic blood flow. *Endocr Connect.* 2017 Apr;6(3):179-87.
152. Jenkins DJA, Sahye-Pudaruth S, Khodabandehlou K, Liang F, Kasmani M, Wanyan J, et al. Systematic review and meta-analysis examining the relationship between postprandial hypotension, cardiovascular events, and all-cause mortality. *Am J Clin Nutr.* 2022 Sep;116(3):663-71.
153. Sarafian D, Charrière N, Maufrais C, Montani JP. Cardiovascular and orthostatic responses to a festive meal associated with alcohol in young men. *Front Physiol.* 2019 Sep;10:1183.
154. Lipsitz LA, Ryan SM, Parker JA, Freeman R, Wei JY, Goldberger AL. Hemodynamic and autonomic nervous system responses to mixed meal ingestion in healthy young and old subjects and dysautonomic patients with postprandial hypotension. *Circulation.* 1993 Feb;87(2):391-400.
155. Trahair LG, Horowitz M, Jones KL. Postprandial hypotension: a systematic review. *J Am Med Dir Assoc.* 2014 Jun;15(6):394-409.

156. Jang A. Postprandial hypotension as a risk factor for the development of new cardiovascular disease: a prospective cohort study with 36 month follow-up in community-dwelling elderly people. *J Clin Med*. 2020 Jan;9(2):345.
157. Westenend M, Lenders JW, Thien T. The course of blood pressure after a meal: a difference between young and elderly subjects. *J Hypertens Suppl*. 1985 Dec;3(3):S417-9.
158. Lipsitz LA, Fullerton KJ. Postprandial blood pressure reduction in healthy elderly. *J Am Geriatr Soc*. 1986 Apr;34(4):267-70.
159. Peitzman SJ, Berger SR. Postprandial blood pressure decrease in well elderly persons. *Arch Intern Med*. 1989 Feb;149(2):286-8.
160. Aronow WS, Ahn C. Postprandial hypotension in 499 elderly persons in a long-term health care facility. *J Am Geriatr Soc*. 1994 Sep;42(9):930-2.
161. Le Couteur DG, Fisher AA, Davis MW, McLean AJ. Postprandial systolic blood pressure responses of older people in residential care: Association with risk of falling. *Gerontology*. 2003 Jul-Aug;49(4):260-4.
162. Puisieux F, Bulckaen H, Fauchais AL, Drumez S, Salomez-Granier F, Dewailly P. Ambulatory blood pressure monitoring and postprandial hypotension in elderly persons with falls or syncope. *J Gerontol A Biol Sci Med Sci*. 2000 Sep;55(9):M535-M40.
163. van Orshoven NP, Jansen PA, Oudejans I, Schoon Y, Oey PL. Postprandial hypotension in clinical geriatric patients and healthy elderly: prevalence related to patient selection and diagnostic criteria. *J Aging Res*. 2010 Sep;2010:243752.
164. Sasaki E, Kitaoka H, Ohsawa N. Postprandial hypotension in patients with non-insulin-dependent diabetes mellitus. *Diabetes Res Clin Pract*. 1992 Nov;18(2):113-21.

165. Jones KL, Doran SM, Hveem K, Bartholomeusz FD, Morley JE, Sun WM, et al. Relation between postprandial satiation and antral area in normal subjects. *Am J Clin Nutr.* 1997 Jul;66(1):127-32.
166. Trahair LG, Horowitz M, Jones KL. Postprandial hypotension is associated with more rapid gastric emptying in healthy older individuals. *J Am Med Dir Assoc.* 2015 Jun;16(6):521-3.
167. Jonsson PV, Lipsitz LA, Kelley M, Koestner J. Hypotensive responses to common daily activities in institutionalized elderly. A potential risk for recurrent falls. *Arch Intern Med.* 1990 Jul;150(7):1518-24.
168. Mathias CJ, da Costa DF, McIntosh CM, Fosbraey P, Bannister R, Wood SM, et al. Differential blood pressure and hormonal effects after glucose and xylose ingestion in chronic autonomic failure. *Clin Sci (Lond).* 1989 Jul;77(1):85-92.
169. Fukushima T, Asahina M, Fujinuma Y, Yamanaka Y, Katagiri A, Mori M, et al. Role of intestinal peptides and the autonomic nervous system in postprandial hypotension in patients with multiple system atrophy. *J Neurol.* 2013 Feb;260(2):475-83.
170. Maruta T, Komai K, Takamori M, Yamada M. Voglibose inhibits postprandial hypotension in neurologic disorders and elderly people. *Neurology.* 2006 May;66(9):1432.
171. Mathias CJ. Postprandial hypotension. Pathophysiological mechanisms and clinical implications in different disorders. *Hypertension.* 1991 Nov;18(5):694-704.
172. Kearney MT, Cowley AJ, Stubbs TA, Evans A, Macdonald IA. Depressor action of insulin on skeletal muscle vasculature: a novel mechanism for

- postprandial hypotension in the elderly. *J Am Coll Cardiol.* 1998 Jan;31(1):209-16.
173. Jansen RW, Peeters TL, van Lier HJ, Hoefnagels WH. The effect of oral glucose, protein, fat and water loading on blood pressure and the gastrointestinal peptides VIP and somatostatin in hypertensive elderly subjects. *Eur J Clin Invest.* 1990 Apr;20(2Part1):192-8.
174. Potter JF, Heseltine D, Hartley G, Matthews J, Macdonald IA, James OFW. Effects of meal composition on the postprandial blood pressure, catecholamine and insulin changes in elderly subjects. *Clin Sci (Lond).* 1989 Sep;77(3):265-72.
175. Waaler BA, Eriksen M. Post-prandial cardiovascular responses in man after ingestion of carbohydrate, protein or fat. *Acta Physiol Scand.* 1992 Nov;146(3):321-7.
176. Puvi-Rajasingham S, Mathias CJ. Effect of meal size on post-prandial blood pressure and on postural hypotension in primary autonomic failure. *Clin Auton Res.* 1996 Apr;6(2):111-4.
177. Jansen RW, Penterman BJ, van Lier HJ, Hoefnagels WH. Blood pressure reduction after oral glucose loading and its relation to age, blood pressure and insulin. *Am J Cardiol.* 1987 Nov;60(13):1087-91.
178. Teunissen-Beekman KF, Dopheide J, Geleijnse JM, Bakker SJ, Brink EJ, de Leeuw PW, et al. Blood pressure decreases more after high-carbohydrate meals than after high-protein meals in overweight adults with elevated blood pressure, but there is no difference after 4 weeks of consuming a carbohydrate-rich or protein-rich diet. *J Nutr.* 2013 Apr;143(4):424-9.

179. Sidery MB, Cowley AJ, MacDonald IA. Cardiovascular responses to a high-fat and a high-carbohydrate meal in healthy elderly subjects. *Clin Sci (Lond)*. 1993 Mar;84(3):263-70.
180. Visvanathan R, Horowitz M, Chapman I. The hypotensive response to oral fat is comparable but slower compared with carbohydrate in healthy elderly subjects. *Br J Nutr*. 2006 Feb;95(2):340-5.
181. Filho SRF, Ferreira F de CR, Oliveira PC, Nery MA. Systemic hemodynamic changes in older hypertensive patients after drinking water or eating a meal. *Hypertension*. 2007 May;49(5):e31; author reply e2.
182. Hoeldtke RD, Boden G, O'Dorisio TM. Treatment of postprandial hypotension with a somatostatin analogue (SMS 201-995). *Am J Med*. 1986 Dec;81(6b):83-7.
183. Burke V, Hodgson JM, Beilin LJ, Giangiulioi N, Rogers P, Puddey IB. Dietary protein and soluble fiber reduce ambulatory blood pressure in treated hypertensives. *Hypertension*. 2001 Oct;38(4):821-6.
184. Hodgson JM, Burke V, Beilin LJ, Puddey IB. Partial substitution of carbohydrate intake with protein intake from lean red meat lowers blood pressure in hypertensive persons. *Am J Clin Nutr*. 2006 Apr;83(4):780-7.
185. Brinkworth GD, Noakes M, Parker B, Foster P, Clifton PM. Long-term effects of advice to consume a high-protein, low-fat diet, rather than a conventional weight-loss diet, in obese adults with type 2 diabetes: one-year follow-up of a randomised trial. *Diabetologia*. 2004 Oct;47(10):1677-86.
186. Appel LJ, Sacks FM, Carey VJ, Obarzanek E, Swain JF, Miller ER, 3rd, et al. Effects of protein, monounsaturated fat, and carbohydrate intake on blood pressure and serum lipids: results of the OmniHeart randomized trial. *JAMA*. 2005 Nov;294(19):2455-64.

187. Stamler J, Elliott P, Kesteloot H, Nichols R, Claeys G, Dyer AR, et al. Inverse relation of dietary protein markers with blood pressure. Findings for 10,020 men and women in the INTERSALT Study. INTERSALT Cooperative Research Group. INTERnational study of SALT and blood pressure. *Circulation*. 1996 Oct;94(7):1629-34.
188. Stamler J, Caggiula A, Grandits GA, Kjelsberg M, Cutler JA. Relationship to blood pressure of combinations of dietary macronutrients. *Circulation*. 1996 Nov;94(10):2417-23.
189. Wang YF, Yancy WS, Jr., Yu D, Champagne C, Appel LJ, Lin PH. The relationship between dietary protein intake and blood pressure: results from the PREMIER study. *J Hum Hypertens*. 2008 Nov;22(11):745-54.
190. Elliott P, Stamler J, Dyer AR, Appel L, Dennis B, Kesteloot H, et al. Association between protein intake and blood pressure: the INTERMAP Study. *Arch Intern Med*. 2006 Jan;166(1):79-87.
191. Fekete AA, Giromini C, Chatzidiakou Y, Givens DI, Lovegrove JA. Whey protein lowers blood pressure and improves endothelial function and lipid biomarkers in adults with prehypertension and mild hypertension: results from the chronic Whey2Go randomized controlled trial. *Am J Clin Nutr*. 2016 Dec;104(6):1534-44.
192. Pal S, Ellis V. The chronic effects of whey proteins on blood pressure, vascular function, and inflammatory markers in overweight individuals. *Obesity (Silver Spring)*. 2010 Jul;18(7):1354-9.
193. Daly RM, Nowson CA. Long-term effect of calcium-vitamin D3 fortified milk on blood pressure and serum lipid concentrations in healthy older men. *Eur J Clin Nutr*. 2009 Aug;63(8):993-1000.

194. Petrogianni M, Grammatikaki E, Kalogeropoulos N, Peristeraki A, Moschonis G, Pitsavos C, et al. Additional benefit in CVD risk indices derived from the consumption of fortified milk when combined with a lifestyle intervention. *Public Health Nutr.* 2014 Feb;17(2):440-9.
195. Petyaev IM, Dovgalevsky PY, Klochkov VA, Chalyk NE, Kyle N. Whey protein lysosome formulation improves vascular functions and plasma lipids with reduction of markers of inflammation and oxidative stress in prehypertension. *Sci World J.* 2012 12;2012:269476.
196. Hidayat K, Du H-Z, Yang J, Chen G-C, Zhang Z, Li Z-N, et al. Effects of milk proteins on blood pressure: a meta-analysis of randomized control trials. *Hypertens Res.* 2017 Mar;40(3):264-70.
197. Pal S, Ellis V. Acute effects of whey protein isolate on blood pressure, vascular function and inflammatory markers in overweight postmenopausal women. *Br J Nutr.* 2011 May;105(10):1512-9.
198. Pihlanto-Leppälä A, Koskinen P, Piilola K, Tupasela T, Korhonen H. Angiotensin I-converting enzyme inhibitory properties of whey protein digests: concentration and characterization of active peptides. *J Dairy Res.* 2000 Feb;67(1):53-64.
199. Sturrock ED, Natesh R, van Rooyen JM, Acharya KR. Structure of angiotensin I-converting enzyme. *Cell Mol Life Sci.* 2004 Nov;61(21):2677-86.
200. Pal S, Radavelli-Bagatini S. The effects of whey protein on cardiometabolic risk factors. *Obes Rev.* 2013 Apr;14(4):324-43.
201. Fluegel SM, Shultz TD, Powers JR, Clark S, Barbosa-Leiker C, Wright BR, et al. Whey beverages decrease blood pressure in prehypertensive and hypertensive young men and women. *Int Dairy J.* 2010 Nov;20(11):753-60.

202. Lee Y-M, Skurk T, Hennig M, Hauner H. Effect of a milk drink supplemented with whey peptides on blood pressure in patients with mild hypertension. *Eur J Nutr.* 2007;46(1):21-7.
203. Gentilcore D, Jones KL, O'Donovan DG, Horowitz M. Postprandial hypotension - novel insights into pathophysiology and therapeutic implications. *Curr Vasc Pharmacol.* 2006 Apr;4(2):161-71.
204. Kuipers HM, Jansen RW, Peeters TL, Hoefnagels WH. The influence of food temperature on postprandial blood pressure reduction and its relation to substance-P in healthy elderly subjects. *J Am Geriatr Soc.* 1991 Feb;39(2):181-4.
205. Deguchi K, Ikeda K, Sasaki I, Shimamura M, Urai Y, Tsukaguchi M, et al. Effects of daily water drinking on orthostatic and postprandial hypotension in patients with multiple system atrophy. *J Neurol.* 2007 Jun;254(6):735-40.
206. Jones KL, MacIntosh C, Su Y-C, Wells F, Chapman IM, Tonkin A, et al. Guar gum reduces postprandial hypotension in older people. *J Am Geriatr Soc.* 2001 Feb;49(2):162-7.
207. Oberman AS, Harada RK, Gagnon MM, Kiely DK, Lipsitz LA. Effects of postprandial walking exercise on meal-related hypotension in frail elderly patients. *Am J Cardiol.* 1999 Nov;84(9):1130-2, a11.
208. Puvi-Rajasingham S, Smith GD, Akinola A, Mathias CJ. Hypotensive and regional haemodynamic effects of exercise, fasted and after food, in human sympathetic denervation. *Clin Sci (Lond).* 1998 Jan;94(1):49-55.
209. Rees WDW, Go VLW, Malagelada J-R. Antroduodenal motor response to solid-liquid and homogenized meals. *Gastroenterology.* 1979 Jun;76(6):1438-42.

210. Goyal RK, Guo Y, Mashimo H. Advances in the physiology of gastric emptying. *Neurogastroenterol Motil.* 2019 Apr;31(4):e13546.
211. Deane AM, Besanko LK, Burgstad CM, Chapman MJ, Horowitz M, Fraser RJ. Modulation of individual components of gastric motor response to duodenal glucose. *World J Gastroenterol.* 2013 Sep;19(35):5863-9.
212. Ryan AT, Luscombe-Marsh ND, Saies AA, Little TJ, Standfield S, Horowitz M, et al. Effects of intraduodenal lipid and protein on gut motility and hormone release, glycemia, appetite, and energy intake in lean men. *Am J Clin Nutr.* 2013 Aug;98(2):300-11.
213. Pilichiewicz AN, Chaikomin R, Brennan IM, Wishart JM, Rayner CK, Jones KL, et al. Load-dependent effects of duodenal glucose on glycemia, gastrointestinal hormones, antropyloroduodenal motility, and energy intake in healthy men. *Am J Physiol Endocrinol Metab.* 2007 Sep;293(3):E743-53.
214. Clarkston WK, Pantano MM, Morley JE, Horowitz M, Littlefield JM, Burton FR. Evidence for the anorexia of aging: gastrointestinal transit and hunger in healthy elderly vs. young adults. *Am J Physiol.* 1997 Jan;272(1 Pt 2):R243-8.
215. Horowitz M, Maddern GJ, Chatterton BE, Collins PJ, Harding PE, Shearman DJ. Changes in gastric emptying rates with age. *Clin Sci (Lond).* 1984 Aug;67(2):213-8.
216. Horowitz M, Edelbroek MA, Wishart JM, Straathof JW. Relationship between oral glucose tolerance and gastric emptying in normal healthy subjects. *Diabetologia.* 1993 Sep;36(9):857-62.
217. Phillips LK, Rayner CK, Jones KL, Horowitz M. Measurement of gastric emptying in diabetes. *J Diabetes Complications.* 2014 Nov-Dec;28(6):894-903.

218. Goyal RK, Cristofaro V, Sullivan MP. Rapid gastric emptying in diabetes mellitus: Pathophysiology and clinical importance. *J Diabetes Complications*. 2019 Nov;33(11):107414.
219. Horowitz M, Harding PE, Maddox A, Maddern GJ, Collins PJ, Chatterton BE, et al. Gastric and oesophageal emptying in insulin-dependent diabetes mellitus. *J Gastroenterol Hepatol*. 1986 Apr;1(2):97-113.
220. Jones KL, Horowitz M, Wishart MJ, Maddox AF, Harding PE, Chatterton BE. Relationships between gastric emptying, intragastric meal distribution and blood glucose concentrations in diabetes mellitus. *J Nucl Med*. 1995 Dec;36(12):2220-8.
221. Chaikomin R, Rayner CK, Jones K-L, Horowitz M. Upper gastrointestinal function and glycemic control in diabetes mellitus. *World J Gastroenterol*. 2006 Sep;12(35):5611-21.
222. Watson LE, Xie C, Wang X, Li Z, Phillips LK, Sun Z, et al. Gastric emptying in patients with well-controlled type 2 diabetes compared with young and older control subjects without diabetes. *J Clin Endocrinol Metab*. 2019 Aug;104(8):3311-9.
223. Phillips LK, Deane AM, Jones KL, Rayner CK, Horowitz M. Gastric emptying and glycaemia in health and diabetes mellitus. *Nat Rev Endocrinol*. 2015 Feb;11(2):112-28.
224. Bharucha AE, Batey-Schaefer B, Cleary PA, Murray JA, Cowie C, Lorenzi G, et al. Delayed gastric emptying is associated with early and long-term hyperglycemia in type 1 diabetes mellitus. *Gastroenterology*. 2015 Aug;149(2):330-9.
225. Bharucha AE, Kudva Y, Basu A, Camilleri M, Low PA, Vella A, et al. Relationship between glycemic control and gastric emptying in poorly

- controlled type 2 diabetes. *Clin Gastroenterol Hepatol*. 2015 Mar;13(3):466-76.e1.
226. Matsumoto M, Yoshimura R, Akiho H, Higuchi N, Kobayashi K, Matsui N, et al. Gastric emptying in diabetic patients by the (13)C-octanoic acid breath test: role of insulin in gastric motility. *J Gastroenterol*. 2007 Jun;42(6):469-74.
227. Jones KL, Tonkin A, Horowitz M, Wishart JM, Carney BI, Guha S, et al. Rate of gastric emptying is a determinant of postprandial hypotension in non-insulin-dependent diabetes mellitus. *Clin Sci (Lond)*. 1998 Jan;94(1):65-70.
228. Vanis L, Gentilcore D, Hausken T, Pilichiewicz AN, Lange K, Rayner CK, et al. Effects of gastric distension on blood pressure and superior mesenteric artery blood flow responses to intraduodenal glucose in healthy older subjects. *Am J Physiol Regul Integr Comp Physiol*. 2010 Sep;299(3):R960-R7.
229. Pham H, Marathe CS, Phillips LK, Trahair LG, Hatzinikolas S, Huynh L, et al. Longitudinal changes in fasting and glucose-stimulated GLP-1 and GIP in healthy older subjects. *J Clin Endocrinol Metab*. 2019 Dec;104(12):6201-6.
230. Gentilcore D, Hausken T, Horowitz M, Jones KL. Measurements of gastric emptying of low- and high-nutrient liquids using 3D ultrasonography and scintigraphy in healthy subjects. *Neurogastroenterol Motil*. 2006 Dec;18(12):1062-8.
231. MacIntosh CG, Morley JE, Wishart J, Morris H, Jansen JB, Horowitz M, et al. Effect of exogenous cholecystokinin (CCK)-8 on food intake and plasma CCK, leptin, and insulin concentrations in older and young adults: evidence for increased CCK activity as a cause of the anorexia of aging. *J Clin Endocrinol Metab*. 2001 Dec;86(12):5830-7.

232. Gutzwiller J-P, Göke B, Drewe J, Hildebrand P, Ketterer S, Handschin D, et al. Glucagon-like peptide-1: a potent regulator of food intake in humans. *Gut*. 1999 Jan;44(1):81-6.
233. Bolondi L, Bortolotti M, Santi V, Calletti T, Gaiani S, Labò G. Measurement of gastric emptying time by real-time ultrasonography. *Gastroenterology*. 1985 Oct;89(4):752-9.
234. Ghos YF, Maes BD, Geypens BJ, Mys G, Hiele MI, Rutgeerts PJ, et al. Measurement of gastric emptying rate of solids by means of a carbon-labeled octanoic acid breath test. *Gastroenterology*. 1993 Jun;104(6):1640-7.
235. Ziegler D, Schadewaldt P, Pour Mirza A, Piolot R, Schommartz B, Reinhardt M, et al. [¹³C]octanoic acid breath test for non-invasive assessment of gastric emptying in diabetic patients: validation and relationship to gastric symptoms and cardiovascular autonomic function. *Diabetologia*. 1996 Jul;39(7):823-30.
236. Delbende B, Perri F, Couturier O, Leodolter A, Mauger P, Bridgi B, et al. ¹³C-octanoic acid breath test for gastric emptying measurement. *Eur J Gastroenterol Hepatol*. 2000 Jan;12(1):85-91.
237. Dickman R, Steinmetz A, Bernnstine H, Groshar D, Niv Y. A novel continuous breath test versus scintigraphy for gastric emptying rate measurement. *J Clin Gastroenterol*. 2011 Jan;45(1):22-5.
238. Zahn A, Langhans CD, Hoffner S, Haberkorn U, Rating D, Haass M, et al. Measurement of gastric emptying by ¹³C-octanoic acid breath test versus scintigraphy in diabetics. *Z Gastroenterol*. 2003 May;41(5):383-90.
239. Stevens JE, Jones KL, Rayner CK, Horowitz M. Pathophysiology and pharmacotherapy of gastroparesis: current and future perspectives. *Expert Opin Pharmacother*. 2013 Jun;14(9):1171-86.

240. Camilleri M, Shin A. Novel and validated approaches for gastric emptying scintigraphy in patients with suspected gastroparesis. *Dig Dis Sci*. 2013 Jul;58(7):1813-5.
241. Galmiche JP, Delbende B, Perri F, Andriulli A. 13C octanoic acid breath test. *Gut*. 1998 Nov;43(suppl 3):S28-30.
242. Sanaka M, Nakada K, Nosaka C, Kuyama Y. The Wagner-Nelson method makes the [13C]-breath test comparable to radioscintigraphy in measuring gastric emptying of a solid/liquid mixed meal in humans. *Clin Exp Pharmacol Physiol*. 2007 Jul;34(7):641-4.
243. Morley JE, Silver AJ. Anorexia in the elderly. *Neurobiol Aging*. 1988 Jan-Feb;9(1):9-16.
244. Castillo EM, Goodman-Gruen D, Kritz-Silverstein D, Morton DJ, Wingard DL, Barrett-Connor E. Sarcopenia in elderly men and women: the Rancho Bernardo study. *Am J Prev Med*. 2003 Oct;25(3):226-31.
245. Janssen I. Evolution of sarcopenia research. *Appl Physiol Nutr Metab*. 2010 Oct;35(5):707-12.
246. Luo D, Lin Z, Li S, Liu SJ. Effect of nutritional supplement combined with exercise intervention on sarcopenia in the elderly: A meta-analysis. *Int J Nurs Sci*. 2017 Oct;4(4):389-401.
247. Bell KE, Snijders T, Zulyniak M, Kumbhare D, Parise G, Chabowski A, et al. A whey protein-based multi-ingredient nutritional supplement stimulates gains in lean body mass and strength in healthy older men: A randomized controlled trial. *PLoS One*. 2017 Jul;12(7):e0181387.
248. Roberts HC, Lim SER, Cox NJ, Ibrahim K. The challenge of managing undernutrition in older people with frailty. *Nutrients*. 2019 Apr;11(4):808.

249. Phillips SM. Current concepts and unresolved questions in dietary protein requirements and supplements in adults. *Front Nutr.* 2017 May;4:13.
250. Traylor DA, Gorissen SHM, Phillips SM. Perspective: protein requirements and optimal intakes in aging: are we ready to recommend more than the recommended daily allowance? *Adv Nutr.* 2018 May;9(3):171-82.
251. Li M, Sun F, Piao JH, Yang XG. Protein requirements in healthy adults: a meta-analysis of nitrogen balance studies. *Biomed Environ Sci.* 2014 Aug;27(8):606-13.
252. Rand WM, Pellett PL, Young VR. Meta-analysis of nitrogen balance studies for estimating protein requirements in healthy adults. *Am J Clin Nutr.* 2003 Jan;77(1):109-27.
253. Rafii M, Chapman K, Elango R, Campbell WW, Ball RO, Pencharz PB, et al. Dietary protein requirement of men >65 years old determined by the indicator amino acid oxidation technique is higher than the current estimated average requirement. *J Nutr.* 2015 Apr;146(4):681-7.
254. Rafii M, Chapman K, Owens J, Elango R, Campbell WW, Ball RO, et al. Dietary protein requirement of female adults >65 years determined by the indicator amino acid oxidation technique is higher than current recommendations. *J Nutr.* 2015 Jan;145(1):18-24.
255. Fulgoni VL, 3rd. Current protein intake in America: analysis of the National Health and Nutrition Examination Survey, 2003-2004. *Am J Clin Nutr.* 2008 May;87(5):1554S-7S.
256. Farsijani S, Payette H, Morais JA, Shatenstein B, Gaudreau P, Chevalier S. Even mealtime distribution of protein intake is associated with greater muscle strength, but not with 3-y physical function decline, in free-living older

- adults: the Quebec longitudinal study on Nutrition as a Determinant of Successful Aging (NuAge study). *Am J Clin Nutr.* 2017 Jul;106(1):113-24.
257. Weinstein JR, Anderson S. The aging kidney: physiological changes. *Adv Chronic Kidney Dis.* 2010 Jul;17(4):302-7.
258. Trumbo P, Schlicker S, Yates AA, Poos M. Dietary reference intakes for energy, carbohydrate, fiber, fat, fatty acids, cholesterol, protein and amino acids. *J Am Diet Assoc.* 2002 Nov;102(11):1621-30.
259. Friedman AN, Ogden LG, Foster GD, Klein S, Stein R, Miller B, et al. Comparative effects of low-carbohydrate high-protein versus low-fat diets on the kidney. *Clin J Am Soc Nephrol.* 2012 Jul;7(7):1103-11.
260. Rizzoli R, Biver E, Bonjour JP, Coxam V, Goltzman D, Kanis JA, et al. Benefits and safety of dietary protein for bone health-an expert consensus paper endorsed by the European Society for Clinical and Economical Aspects of Osteoporosis, Osteoarthritis, and Musculoskeletal Diseases and by the International Osteoporosis Foundation. *Osteoporos Int.* 2018 Sep;29(9):1933-48.
261. Aronow WS, Ahn C. Association of postprandial hypotension with incidence of falls, syncope, coronary events, stroke, and total mortality at 29-month follow-up in 499 older nursing home residents. *J Am Geriatr Soc.* 1997 Sep;45(9):1051-3.
262. Jansen RW, Lipsitz LA. Postprandial hypotension: epidemiology, pathophysiology, and clinical management. *Ann Intern Med.* 1995 Feb;122(4):286-95.
263. Gentilcore D, Hausken T, Meyer JH, Chapman IM, Horowitz M, Jones KL. Effects of intraduodenal glucose, fat, and protein on blood pressure, heart

- rate, and splanchnic blood flow in healthy older subjects. *Am J Clin Nutr.* 2008 Jan;87(1):156-61.
264. Giezenaar C, Oberoi A, Jones KL, Horowitz M, Chapman I, Soenen S. Effects of age on blood pressure and heart rate responses to whey protein in younger and older men. *J Am Geriatr Soc.* 2021 May;69(5):1291-99.
265. Oberoi A, Giezenaar C, Tippet R, Jones KL, Chapman, I, Soenen S. Effects of whey protein and glucose intake on energy intake, gastric emptying and glycaemia in healthy older subjects. Paper presented at European Association for the Study of Diabetes; 2020 Sep 22; Virtual.
266. Milne AC, Potter J, Vivanti A, Avenell A. Protein and energy supplementation in elderly people at risk from malnutrition. *Cochrane Database Syst Rev.* 2009 Apr;2009(2):Cd003288.
267. McDonald CK, Ankarfeldt MZ, Capra S, Bauer J, Raymond K, Heitmann BL. Lean body mass change over 6 years is associated with dietary leucine intake in an older Danish population. *Br J Nutr.* 2016 May;115(9):1556-62.
268. Norton LE, Layman DK, Bunpo P, Anthony TG, Brana DV, Garlick PJ. The leucine content of a complete meal directs peak activation but not duration of skeletal muscle protein synthesis and mammalian target of rapamycin signaling in rats. *J Nutr.* 2009 Jun;139(6):1103-9.
269. Mitchell CJ, McGregor RA, D'Souza RF, Thorstensen EB, Markworth JF, Fanning AC, et al. Consumption of milk protein or whey protein results in a similar increase in muscle protein synthesis in middle aged men. *Nutrients.* 2015 Oct;7(10):8685-99.
270. Pennings B, Groen B, de Lange A, Gijsen AP, Zorenc AH, Senden JM, et al. Amino acid absorption and subsequent muscle protein accretion following

- graded intakes of whey protein in elderly men. *Am J Physiol Endocrinol Metab.* 2012 Apr;302(8):E992-9.
271. Komar B, Schwingshackl L, Hoffmann G. Effects of leucine-rich protein supplements on anthropometric parameter and muscle strength in the elderly: a systematic review and meta-analysis. *J Nutr Health Aging.* 2015 Apr;19(4):437-46.
272. Churchward-Venne TA, Holwerda AM, Phillips SM, van Loon LJ. What is the optimal amount of protein to support post-exercise skeletal muscle reconditioning in the older adult? *Sports Med.* 2016 Sep;46(9):1205-12.
273. Yang Y, Breen L, Burd NA, Hector AJ, Churchward-Venne TA, Josse AR, et al. Resistance exercise enhances myofibrillar protein synthesis with graded intakes of whey protein in older men. *Br J Nutr.* 2012 Nov;108(10):1780-8.
274. Esmarck B, Andersen JL, Olsen S, Richter EA, Mizuno M, Kjaer M. Timing of postexercise protein intake is important for muscle hypertrophy with resistance training in elderly humans. *J Physiol.* 2001 Aug;535(Pt 1):301-11.
275. Grech A, Rangan A, Allman-Farinelli M. Macronutrient composition of the Australian population's Diet; trends from three national nutrition surveys 1983, 1995 and 2012. *Nutrients.* 2018 Aug 8;10(8).
276. Soenen S, Westerterp-Plantenga MS. Proteins and satiety: implications for weight management. *Curr Opin Clin Nutr Metab Care.* 2008 Nov;11(6):747-51.
277. Oberoi A, Giezenaar C, Clames A, Bøhler K, Lange K, Horowitz M, et al. Whey protein drink ingestion before breakfast suppressed energy intake at breakfast and lunch, but not during dinner, and was less suppressed in healthy older than younger men. *Nutrients.* 2020 Oct;12(11):3318.

278. Oberoi A, Giezenaar C, Jensen C, Lange K, Hausken T, Jones KL, et al. Acute effects of whey protein on energy intake, appetite and gastric emptying in younger and older, obese men. *Nutr Diabetes*. 2020 Oct 2;10(1):37.
279. Yasawy MI, Al-Quorain AA, Hussameddin AM, Yasawy ZM, Al-Sulaiman RM. Obesity and gastric balloon. *J Family Community Med*. 2014 Sep;21(3):196-9.
280. Miyawaki K, Yamada Y, Ban N, Ihara Y, Tsukiyama K, Zhou H, et al. Inhibition of gastric inhibitory polypeptide signaling prevents obesity. *Nat Med*. 2002 Jul;8(7):738-42.
281. Nass R, Farhy LS, Liu J, Pezzoli SS, Johnson ML, Gaylinn BD, et al. Age-dependent decline in acyl-ghrelin concentrations and reduced association of acyl-ghrelin and growth hormone in healthy older adults. *J Clin Endocrinol Metab*. 2014 Feb;99(2):602-8.
282. Rigamonti AE, Pincelli AI, Corrà B, Viarengo R, Bonomo SM, Galimberti D, et al. Plasma ghrelin concentrations in elderly subjects: comparison with anorexic and obese patients. *J Endocrinol*. 2002 Oct;175(1):R1-5.
283. Leslie W, Hankey C. Aging, nutritional status and health. *Healthcare (Basel)*. 2015 Jul;3(3):648-58.
284. Siparsky PN, Kirkendall DT, Garrett WE, Jr. Muscle changes in aging: understanding sarcopenia. *Sports Health*. 2014 Jan;6(1):36-40.
285. Soenen S, Rayner CK, Jones KL, Horowitz M. The ageing gastrointestinal tract. *Curr Opin Clin Nutr Metab Care*. 2016 Jan;19(1):12-8.
286. Cawood AL, Elia M, Stratton RJ. Systematic review and meta-analysis of the effects of high protein oral nutritional supplements. *Ageing Res Rev*. 2012 Apr;11(2):278-96.

287. Malafarina V, Uriz-Otano F, Iniesta R, Gil-Guerrero L. Effectiveness of nutritional supplementation on muscle mass in treatment of sarcopenia in old age: a systematic review. *J Am Med Dir Assoc*. 2013 Jan;14(1):10-7.
288. Marshall K. Therapeutic applications of whey protein. *Altern Med Rev*. 2004 Jun;9(2):136-56.
289. Smithers GW. Whey and whey proteins—From ‘gutter-to-gold’. *Int Dairy J*. 2008 Jul;18(7):695-704.
290. Hveem K, Jones KL, Chatterton BE, Horowitz M. Scintigraphic measurement of gastric emptying and ultrasonographic assessment of antral area: relation to appetite. *Gut*. 1996;38(6):816-21.
291. Parker BA, Sturm K, MacIntosh CG, Feinle C, Horowitz M, Chapman IM. Relation between food intake and visual analogue scale ratings of appetite and other sensations in healthy older and young subjects. *Eur J Clin Nutr*. 2004 Feb;58(2):212-8.
292. Janssen P, Vanden Berghe P, Verschueren S, Lehmann A, Depoortere I, Tack J. Review article: the role of gastric motility in the control of food intake. *Aliment Pharmacol Ther*. 2011 Apr;33(8):880-94.
293. Rigamonti AE, Leoncini R, Casnici C, Marelli O, Col A, Tamini S, et al. Whey proteins reduce appetite, stimulate anorexigenic gastrointestinal peptides and improve glucometabolic homeostasis in young obese women. *Nutrients*. 2019 Jan;11(2):247.
294. Roberts SB, Fuss P, Heyman MB, Evans WJ, Tsay R, Rasmussen H, et al. Control of food intake in older men. *JAMA*. 1994 Nov;272(20):1601-6.
295. Griffin HJ, Cheng HL, O'Connor HT, Rooney KB, Petocz P, Steinbeck KS. Higher protein diet for weight management in young overweight women: a

- 12-month randomized controlled trial. *Diabetes Obes Metab.* 2013 Jun;15(6):572-5.
296. Leidy HJ, Carnell NS, Mattes RD, Campbell WW. Higher protein intake preserves lean mass and satiety with weight loss in pre-obese and obese women. *Obesity (Silver Spring)*. 2007 Feb;15(2):421-9.
297. Mokdad AH, Ford ES, Bowman BA, Dietz WH, Vinicor F, Bales VS, et al. Prevalence of obesity, diabetes, and obesity-related health risk factors, 2001. *JAMA*. 2003 Jan;289(1):76-9.
298. Flegal KM, Carroll MD, Ogden CL, Curtin LR. Prevalence and trends in obesity among US adults, 1999-2008. *JAMA*. 2010 Jan;303(3):235-41.
299. Huse O, Hettiarachchi J, Gearon E, Nichols M, Allender S, Peeters A. Obesity in Australia. *Obes Res Clin Pract*. 2018 Jan-Feb;12(1):29-39.
300. Petermann-Rocha F, Chen M, Gray SR, Ho FK, Pell JP, Celis-Morales C. Factors associated with sarcopenia: a cross-sectional analysis using UK Biobank. *Maturitas*. 2020 Mar;133:60-7.
301. Yu C-Y, Woo A, Emrich CT, Wang B. Social Vulnerability Index and obesity: an empirical study in the US. *Cities*. 2020 Feb;97:102531.
302. Janssen I. Morbidity and mortality risk associated with an overweight BMI in older men and women. *Obesity (Silver Spring)*. 2007 Jul;15(7):1827-40.
303. Arterburn DE, Crane PK, Sullivan SD. The coming epidemic of obesity in elderly Americans. *J Am Geriatr Soc*. 2004 Nov;52(11):1907-12.
304. Roubenoff R. Sarcopenic obesity: the confluence of two epidemics. *Obes Res*. 2004 Jun;12(6):887-8.
305. Villareal DT, Apovian CM, Kushner RF, Klein S. Obesity in older adults: technical review and position statement of the American Society for Nutrition and NAASO, The Obesity Society. *Am J Clin Nutr*. 2005 Nov;82(5):923-34.

306. Villareal DT, Chode S, Parimi N, Sinacore DR, Hilton T, Armamento-Villareal R, et al. Weight loss, exercise, or both and physical function in obese older adults. *N Engl J Med*. 2011 Mar;364(13):1218-29.
307. Kyrou I, Tsigos C. Obesity in the elderly diabetic patient: is weight loss beneficial? No. *Diabetes Care*. 2009 Nov;32 Suppl 2(Suppl 2):S403-9.
308. Evans WJ, Campbell WW. Sarcopenia and age-related changes in body composition and functional capacity. *J Nutr*. 1993 Feb;123(2 Suppl):465-8.
309. Beals JW, Burd NA, Moore DR, van Vliet S. Obesity alters the muscle protein synthetic response to nutrition and exercise. *Front Nutr*. 2019 Jun;6:87.
310. Li Z, Heber D. Sarcopenic obesity in the elderly and strategies for weight management. *Nutr Rev*. 2012 Jan;70(1):57-64.
311. Blaum CS, Xue QL, Michelon E, Semba RD, Fried LP. The association between obesity and the frailty syndrome in older women: the Women's Health and Aging Studies. *J Am Geriatr Soc*. 2005 Jun;53(6):927-34.
312. Lapane KL, Resnik L. Obesity in nursing homes: an escalating problem. *J Am Geriatr Soc*. 2005 Aug;53(8):1386-91.
313. Villareal DT, Banks M, Siener C, Sinacore DR, Klein S. Physical frailty and body composition in obese elderly men and women. *Obes Res*. 2004 Jun;12(6):913-20.
314. Zizza CA, Herring A, Stevens J, Popkin BM. Obesity affects nursing-care facility admission among whites but not blacks. *Obes Res*. 2002 Aug;10(8):816-23.
315. Elkins JS, Whitmer RA, Sidney S, Sorel M, Yaffe K, Johnston SC. Midlife obesity and long-term risk of nursing home admission. *Obesity (Silver Spring)*. 2006 Aug;14(8):1472-8.

316. Soenen S, Hochstenbach-Waelen A, Westerterp-Plantenga MS. Efficacy of α -lactalbumin and milk protein on weight loss and body composition during energy restriction. *Obesity (Silver Spring)*. 2011 Feb;19(2):370-9.
317. Bray GA, Siri-Tarino PW. The role of macronutrient content in the diet for weight management. *Endocrinol Metab Clin North Am*. 2016 Sep;45(3):581-604.
318. Blatt AD, Roe LS, Rolls BJ. Increasing the protein content of meals and its effect on daily energy intake. *J Am Diet Assoc*. 2011 Feb;111(2):290-4.
319. Devries MC, Phillips SM. Supplemental protein in support of muscle mass and health: advantage whey. *J Food Sci*. 2015 Mar;80 Suppl 1:A8-A15.
320. Ryan AT, Feinle-Bisset C, Kallas A, Wishart JM, Clifton PM, Horowitz M, et al. Intraduodenal protein modulates antropyloroduodenal motility, hormone release, glycemia, appetite, and energy intake in lean men. *Am J Clin Nutr*. 2012 Sep;96(3):474-82.
321. Marathe CS, Rayner CK, Jones KL, Horowitz M. Relationships between gastric emptying, postprandial glycemia, and incretin hormones. *Diabetes Care*. 2013 May;36(5):1396-405.
322. Hunt JN. A possible relation between the regulation of gastric emptying and food intake. *Am J Physiol*. 1980 Jul;239(1):G1-4.
323. Hunt JN, Stubbs DF. The volume and energy content of meals as determinants of gastric emptying. *J Physiol*. 1975 Feb;245(1):209-25.
324. Wright RA, Krinsky S, Fleeman C, Trujillo J, Teague E. Gastric emptying and obesity. *Gastroenterology*. 1983 Apr;84(4):747-51.
325. Moore JG, Tweedy C, Christian PE, Datz FL. Effect of age on gastric emptying of liquid—solid meals in man. *Dig Dis Sci*. 1983 Apr;28(4):340-4.

326. Folstein MF, Folstein SE, McHugh PR. "Mini-mental state". A practical method for grading the cognitive state of patients for the clinician. *J Psychiatr Res.* 1975 Nov;12(3):189-98.
327. Yesavage JA, Brink TL, Rose TL, Lum O, Huang V, Adey M, et al. Development and validation of a geriatric depression screening scale: a preliminary report. *J Psychiatr Res.* 1982;17(1):37-49.
328. Atkins RC. *Dr Atkins' new diet revolution.* 1st Avon pbk. ed. New York: Avon Books;2002. 540 p.
329. Sears B Lawren B. *The zone: a dietary road map.* New York: Regan Books; 1995.286 p.
330. Julia C, Péneau S, Andreeva VA, Méjean C, Fezeu L, Galan P, et al. Weight-loss strategies used by the general population: how are they perceived? *PLoS One.* 2014 May;9(5):e97834.
331. Brennan IM, Luscombe-Marsh ND, Seimon RV, Otto B, Horowitz M, Wishart JM, et al. Effects of fat, protein, and carbohydrate and protein load on appetite, plasma cholecystokinin, peptide YY, and ghrelin, and energy intake in lean and obese men. *Am J Physiol Gastrointest Liver Physiol.* 2012 Jul;303(1):G129-40.
332. Mourao DM, Bressan J, Campbell WW, Mattes RD. Effects of food form on appetite and energy intake in lean and obese young adults. *Int J Obes (Lond).* 2007 Nov;31(11):1688-95.
333. St-Onge MP, Gallagher D. Body composition changes with aging: the cause or the result of alterations in metabolic rate and macronutrient oxidation? *Nutrition.* 2010 Feb;26(2):152-5.

334. Lancha AH, Jr., Zanella R, Jr., Tanabe SG, Andriamihaja M, Blachier F. Dietary protein supplementation in the elderly for limiting muscle mass loss. *Amino Acids*. 2017 Jan;49(1):33-47.
335. Stevens JE, Gilja OH, Gentilcore D, Hausken T, Horowitz M, Jones KL. Measurement of gastric emptying of a high-nutrient liquid by 3D ultrasonography in diabetic gastroparesis. *Neurogastroenterol Motil*. 2011 Mar;23(3):220-5, e113-4.
336. Khan MAB, Hashim MJ, King JK, Govender RD, Mustafa H, Al Kaabi J. Epidemiology of type 2 diabetes - global burden of disease and forecasted trends. *J Epidemiol Glob Health*. 2020 Mar;10(1):107-11.
337. Lin X, Xu Y, Pan X, Xu J, Ding Y, Sun X, et al. Global, regional, and national burden and trend of diabetes in 195 countries and territories: an analysis from 1990 to 2025. *Sci Rep*. 2020 Sep;10(1):14790.
338. Dunstan DW, Zimmet PZ, Welborn TA, de Courten MP, Cameron AJ, Sicree RA, et al. The rising prevalence of diabetes and impaired glucose tolerance: the Australian diabetes, obesity and lifestyle study. *Diabetes Care*. 2002 May;25(5):829-34.
339. Australian Institute of Health and Welfare. Diabetes prevalence in Australia: detailed estimates for 2007–08. Canberra: AIHW; 2011 [cited 2022 Feb 10]. 32 p. Available from: <https://www.aihw.gov.au/reports/diabetes/diabetes-prevalence-detailed-estimates-2007-08/contents/table-of-contents>
340. Monnier L, Colette C. Contributions of fasting and postprandial glucose to hemoglobin A1c. *Endocr Pract*. 2006 Jan-Feb;12 Suppl 1:42-6.
341. Stevenson EJ, Allerton DM. The role of whey protein in postprandial glycaemic control. *Proc Nutr Soc*. 2018 Feb;77(1):42-51.

342. Kirkman MS, Briscoe VJ, Clark N, Florez H, Haas LB, Halter JB, et al. Diabetes in older adults. *Diabetes Care*. 2012 Dec;35(12):2650-64.
343. Leung E, Wongrakpanich S, Munshi MN. Diabetes management in the elderly. *Diabetes Spectr*. 2018 Aug;31(3):245-53.
344. Node K, Inoue T. Postprandial hyperglycemia as an etiological factor in vascular failure. *Cardiovasc Diabetol*. 2009 Apr;8:23.
345. Blaak EE, Antoine JM, Benton D, Björck I, Bozzetto L, Brouns F, et al. Impact of postprandial glycaemia on health and prevention of disease. *Obes Rev*. 2012 Oct;13(10):923-84.
346. Gannon MC, Nuttall FQ. Amino acid ingestion and glucose metabolism—a review. *IUBMB Life*. 2010 Sep;62(9):660-8.
347. Pallotta JA, Kennedy PJ. Response of plasma insulin and growth hormone to carbohydrate and protein feeding. *Metabolism*. 1968 Oct;17(10):901-8.
348. Gannon MC, Nuttall FQ, Lane JT, Burmeister LA. Metabolic response to cottage cheese or egg white protein, with or without glucose, in type II diabetic subjects. *Metabolism*. 1992 Oct;41(10):1137-45.
349. Gannon MC, Nuttall FQ, Grant CT, Ercan-Fang S, Ercan-Fang N. Stimulation of insulin secretion by fructose ingested with protein in people with untreated type 2 diabetes. *Diabetes Care*. 1998 Jan;21(1):16-22.
350. Gunnerud U, Östman E, Björck I. Effects of whey proteins on glycaemia and insulinaemia to an oral glucose load in healthy adults; a dose–response study. *Eur J Clin Nutr*. 2013 Jul;67(7):749-53.
351. Kung B, Anderson GH, Paré S, Tucker AJ, Vien S, Wright AJ, et al. Effect of milk protein intake and casein-to-whey ratio in breakfast meals on postprandial glucose, satiety ratings, and subsequent meal intake. *J Dairy Sci*. 2018 Oct;101(10):8688-701.

352. Floyd JC, Jr., Fajans SS, Conn JW, Knopf RF, Rull J. Insulin secretion in response to protein ingestion. *J Clin Invest.* 1966 Sep;45(9):1479-86.
353. McGregor RA, Poppitt SD. Milk protein for improved metabolic health: a review of the evidence. *Nutr Metab (Lond).* 2013 Jul;10(1):46.
354. Akhavan T, Luhovyy BL, Brown PH, Cho CE, Anderson GH. Effect of premeal consumption of whey protein and its hydrolysate on food intake and postmeal glycemia and insulin responses in young adults. *Am J Clin Nutr.* 2010 Apr;91 4:966-75.
355. Hermans MP, Pepersack TM, Godeaux LH, Beyer I, Turc AP. Prevalence and determinants of impaired glucose metabolism in frail elderly patients: the Belgian Elderly Diabetes Survey (BEDS). *J Gerontol A Biol Sci Med Sci.* 2005 Feb;60(2):241-7.
356. Chia CW, Egan JM, Ferrucci L. Age-related changes in glucose metabolism, hyperglycemia, and cardiovascular risk. *Circ Res.* 2018 Sep;123(7):886-904.
357. Kalyani RR, Egan JM. Diabetes and altered glucose metabolism with aging. *Endocrinol Metab Clin North Am.* 2013 Jun;42(2):333-47.
358. Lindström T, Hedman CA, Arnqvist HJ. Use of a novel double-antibody technique to describe the pharmacokinetics of rapid-acting insulin analogs. *Diabetes Care.* 2002 Jun;25(6):1049-54.
359. Braden B, Adams S, Duan LP, Orth KH, Maul FD, Lembcke B, et al. The [13C]acetate breath test accurately reflects gastric emptying of liquids in both liquid and semisolid test meals. *Gastroenterology.* 1995 Apr;108(4):1048-55.
360. Bjorkman DJ, Moore JG, Klein PD, Graham DY. 13C-bicarbonate breath test as a measure of gastric emptying. *Am J Gastroenterol.* 1991 Jul;86(7):821-3.

361. Sanaka M, Yamamoto T, Ishii T, Kuyama Y. The Wagner-Nelson method can generate an accurate gastric emptying flow curve from CO₂ data obtained by a ¹³C-labeled substrate breath test. *Digestion*. 2004;69(2):71-8.
362. Trahair LG, Nauck MA, Wu T, Stevens JE, Butfield MD, Hatzinikolas S, et al. Measurement of gastric emptying using a ¹³C-octanoic acid breath test with Wagner-Nelson analysis and scintigraphy in type 2 diabetes. *Exp Clin Endocrinol Diabetes* [Internet]. 2022 Mar 1 [cited 2022 Jun 15]. Available from: https://eref.thieme.de/ejournals/1439-3646_efirst#/10.1055-a-1784-6185
363. Meneilly GS, Ryan AS, Minaker KL, Elahi D. The effect of age and glycemic level on the response of the β -cell to glucose-dependent insulinotropic polypeptide and peripheral tissue sensitivity to endogenously released insulin. *J Clin Endocrinol Metab*. 1998 Aug;83(8):2925-32.
364. Weber P, Kolácný I. [Glucose tolerance in the elderly--changes in insulin, C-peptide and glucagon secretion]. *Wien Med Wochenschr*. 1992;142(4):73-8.
365. Stevic R, Zivkovic TB, Erceg P, Milosevic D, Despotovic N, Davidovic M. Oral glucose tolerance test in the assessment of glucose-tolerance in the elderly people. *Age ageing*. 2007 Jul;36(4):459-62.
366. Watson LE, Phillips LK, Wu T, Bound MJ, Checklin HL, Grivell J, et al. A whey/guar "preload" improves postprandial glycaemia and glycated haemoglobin levels in type 2 diabetes: a 12-week, single-blind, randomized, placebo-controlled trial. *Diabetes Obes Metab*. 2019 Apr;21(4):930-8.
367. Tinajero MG, Malik VS. An update on the epidemiology of type 2 diabetes: a global perspective. *Endocrinol Metab Clin North Am*. 2021 Sep;50(3):337-55.

368. Galicia-Garcia U, Benito-Vicente A, Jebari S, Larrea-Sebal A, Siddiqi H, Uribe KB, et al. Pathophysiology of type 2 diabetes mellitus. *Int J Mol Sci*. 2020 Aug;21(17).
369. American Diabetes Association. Diagnosis and classification of diabetes mellitus. *Diabetes Care*. 2009 Jan;32 Suppl 1(Suppl 1):S62-7.
370. Tahrani AA, Bailey CJ, Del Prato S, Barnett AH. Management of type 2 diabetes: new and future developments in treatment. *Lancet*. 2011 Jul;378(9786):182-97.
371. Salas-Salvadó J, Martínez-González M, Bulló M, Ros E. The role of diet in the prevention of type 2 diabetes. *Nutr Metab Cardiovasc Dis*. 2011 Sep;21(Suppl 2):B32-48.
372. Knowler WC, Barrett-Connor E, Fowler SE, Hamman RF, Lachin JM, Walker EA, et al. Reduction in the incidence of type 2 diabetes with lifestyle intervention or metformin. *N Engl J Med*. 2002 Feb;346(6):393-403.
373. Tuomilehto J, Lindström J, Eriksson JG, Valle TT, Hämäläinen H, Ilanne-Parikka P, et al. Prevention of type 2 diabetes mellitus by changes in lifestyle among subjects with impaired glucose tolerance. *N Engl J Med*. 2001 May;344(18):1343-50.
374. Valencia WM, Florez H. Pharmacological treatment of diabetes in older people. *Diabetes Obes Metab*. 2014 Dec;16(12):1192-203.
375. Nathan DM, Buse JB, Davidson MB, Ferrannini E, Holman RR, Sherwin R, et al. Medical management of hyperglycemia in type 2 diabetes: a consensus algorithm for the initiation and adjustment of therapy: a consensus statement of the American Diabetes Association and the European Association for the Study of Diabetes. *Diabetes Care*. 2009 Jan;32(1):193-203.

376. Manders RJ, Koopman R, Sluijsmans WE, van den Berg R, Verbeek K, Saris WH, et al. Co-ingestion of a protein hydrolysate with or without additional leucine effectively reduces postprandial blood glucose excursions in Type 2 diabetic men. *J Nutr.* 2006 May;136(5):1294-9.
377. van Loon LJ, Kruijshoop M, Menheere PP, Wagenmakers AJ, Saris WH, Keizer HA. Amino acid ingestion strongly enhances insulin secretion in patients with long-term type 2 diabetes. *Diabetes Care.* 2003 Mar;26(3):625-30.
378. Hruby A, Ma J, Rogers G, Meigs JB, Jacques PF. Associations of dairy intake with incident prediabetes or diabetes in middle-aged adults vary by both dairy type and glycemic status. *J Nutr.* 2017 Sep;147(9):1764-75.
379. Díaz-López A, Bulló M, Martínez-González MA, Corella D, Estruch R, Fitó M, et al. Dairy product consumption and risk of type 2 diabetes in an elderly Spanish Mediterranean population at high cardiovascular risk. *Eur J Nutr.* 2016 Feb;55(1):349-60.
380. Lovegrove JA, Givens DI. Dairy food products: good or bad for cardiometabolic disease? *Nutr Res Rev.* 2016 Dec;29(2):249-67.
381. Da Silva MS, Julien P, Couture P, Lemieux S, Vohl MC, Rudkowska I. Associations between dairy intake and metabolic risk parameters in a healthy French-Canadian population. *Appl Physiol Nutr Metab.* 2014 Dec;39(12):1323-31.
382. Akhavan T, Luhovyy BL, Panahi S, Kubant R, Brown PH, Anderson GH. Mechanism of action of pre-meal consumption of whey protein on glycemic control in young adults. *J Nutr Biochem.* 2014 Jan;25(1):36-43.
383. Jakubowicz D, Froy O, Ahrén B, Boaz M, Landau Z, Bar-Dayán Y, et al. Incretin, insulinotropic and glucose-lowering effects of whey protein pre-

- load in type 2 diabetes: a randomised clinical trial. *Diabetologia*. 2014 Sep;57(9):1807-11.
384. Aune D, Norat T, Romundstad P, Vatten LJ. Dairy products and the risk of type 2 diabetes: a systematic review and dose-response meta-analysis of cohort studies. *Am J Clin Nutr*. 2013 Oct;98(4):1066-83.
385. Salehi A, Gunnerud U, Muhammed SJ, Östman E, Holst JJ, Björck I, et al. The insulinogenic effect of whey protein is partially mediated by a direct effect of amino acids and GIP on β -cells. *Nutr Metab (Lond)*. 2012 May;9(1):48.
386. Pallotta JA, Kennedy PJ. Response of plasma insulin and growth hormone to carbohydrate and protein feeding. *Metabolism*. 1968 Oct;17(10):901-8.
387. Porte D, Jr., Kahn SE. Beta-cell dysfunction and failure in type 2 diabetes: potential mechanisms. *Diabetes*. 2001 Feb;50 Suppl 1:S160-3.
388. Stanstrup J, Schou SS, Holmer-Jensen J, Hermansen K, Dragsted LO. Whey protein delays gastric emptying and suppresses plasma fatty acids and their metabolites compared to casein, gluten, and fish protein. *J Proteome Res*. 2014 May;13(5):2396-408.
389. Krishnasamy S, Abell TL. Diabetic gastroparesis: principles and current trends in management. *Diabetes Ther*. 2018 Jul;9(Suppl 1):1-42.
390. Horowitz M, Harding PE, Maddox AF, Wishart JM, Akkermans LM, Chatterton BE, et al. Gastric and oesophageal emptying in patients with type 2 (non-insulin-dependent) diabetes mellitus. *Diabetologia*. 1989 Mar;32(3):151-9.
391. King DG, Walker M, Campbell MD, Breen L, Stevenson EJ, West DJ. A small dose of whey protein co-ingested with mixed-macronutrient breakfast and lunch meals improves postprandial glycemia and suppresses appetite in

- men with type 2 diabetes: a randomized controlled trial. *Am J Clin Nutr.* 2018 Apr;107(4):550-7.
392. Fagius J, Ellerfelt K, Lithell H, Berne C. Increase in muscle nerve sympathetic activity after glucose intake is blunted in the elderly. *Clin Auton Res.* 1996 Aug;6(4):195-203.
393. Fisher AA, Davis MW, Srikusalanukul W, Budge MM. Postprandial hypotension predicts all-cause mortality in older, low-level care residents. *J Am Geriatr Soc.* 2005 Aug;53(8):1313-20.
394. Vaitkevicius PV, Esserwein DM, Maynard AK, O'Connor FC, Fleg JL. Frequency and importance of postprandial blood pressure reduction in elderly nursing-home patients. *Ann Intern Med.* 1991 Dec;115(11):865-70.
395. Lubart E, Segal R, Baumoehl Y, Matron M, Leibovitz A. Postprandial hypotension in long-term care elderly patients on enteral feeding. *J Am Geriatr Soc.* 2006;54(9):1377-81.
396. Trahair LG, Vanis L, Gentilcore D, Lange K, Rayner CK, Horowitz M, et al. Effects of variations in duodenal glucose load on blood pressure, heart rate, superior mesenteric artery blood flow and plasma noradrenaline in healthy young and older subjects. *Clin Sci (Lond).* 2012 Mar;122(6):271-9.
397. Trahair LG, Horowitz M, Stevens JE, Feinle-Bisset C, Standfield S, Piscitelli D, et al. Effects of exogenous glucagon-like peptide-1 on blood pressure, heart rate, gastric emptying, mesenteric blood flow and glycaemic responses to oral glucose in older individuals with normal glucose tolerance or type 2 diabetes. *Diabetologia.* 2015 Aug;58(8):1769-78.
398. Jones KL, Rigda RS, Buttfield MDM, Hatzinikolas S, Pham HT, Marathe CS, et al. Effects of lixisenatide on postprandial blood pressure, gastric

- emptying and glycaemia in healthy people and people with type 2 diabetes. *Diabetes Obes Metab.* 2019 May;21(5):1158-67.
399. Zhang X, Jones KL, Horowitz M, Rayner CK, Wu T. Effects of proximal and distal enteral glucose infusion on cardiovascular response in health and type 2 diabetes. *J Clin Endocrinol Metab.* 2020 Aug;105(8):e2877-e84.
400. Goodman BE. Insights into digestion and absorption of major nutrients in humans. *Adv Physiol Educ.* 2010 Jun;34(2):44-53.
401. Qamar MI, Read AE. Effects of ingestion of carbohydrate, fat, protein, and water on the mesenteric blood flow in man. *Scand J Gastroenterol.* 1988 Jan;23(1):26-30.
402. Maurer MS, Karmally W, Rivadeneira H, Parides MK, Bloomfield DM. Upright posture and postprandial hypotension in elderly persons. *Ann Intern Med.* 2000 Oct;133(7):533-6.
403. Bannister R, Da Costa DF, Forster S, Fosbraey P, Mathias CJ. Cardiovascular effects of lipid and protein meals in autonomic failure. *J Physiol.* 1986;377(suppl):19-78.
404. Uijtdehaage SHJ, Shapiro D, Jaquet F. Effects of carbohydrate and protein meals on cardiovascular levels and reactivity. *Biol Psychol.* 1994 Sep;38(1):53-72.
405. Vloet LCM, Smits R, Jansen RWMM. The effect of meals at different mealtimes on blood pressure and symptoms in geriatric patients with postprandial hypotension. *J Gerontol A Biol Sci Med Sci.* 2003 Nov;58(11):M1031-M5.
406. Pham H, Holen IS, Phillips LK, Hatzinikolas S, Huynh LQ, Wu T, et al. The effects of a whey protein and guar gum-containing preload on gastric

- emptying, glycaemia, small intestinal absorption and blood pressure in healthy older subjects. *Nutrients*. 2019 Nov;11(11).
407. Guigoz Y, Vellas B, Garry PJ. Assessing the nutritional status of the elderly: The Mini Nutritional Assessment as part of the geriatric evaluation. *Nutr Rev*. 1996 Jan;54(1 Pt 2):S59-65.
408. Cicero AFG, Fogacci F, Veronesi M, Grandi E, Dinelli G, Hrelia S, et al. Short-term hemodynamic effects of modern wheat products substitution in diet with ancient wheat products: a cross-over, randomized clinical trial. *Nutrients*. 2018 Nov;10(11):1666.
409. National Institute on Aging, National Institutes of Health. Global health and aging. Washington: World Health Organization; 2011 [cited 2022 Sep 27]. 32 p. Available from: https://www.nia.nih.gov/sites/default/files/2017-06/global_health_aging.pdf.
410. Vasan RS, Beiser A, Seshadri S, Larson MG, Kannel WB, D'Agostino RB, et al. Residual lifetime risk for developing hypertension in middle-aged women and men: the Framingham heart study. *JAMA*. 2002 Feb;287(8):1003-10.
411. Madden KM, Feldman B, Meneilly GS. Characteristics Associated with the Postprandial Hypotensive Response in Falling Older Adults. *CJA/RCV*. 2019;38:434-40.

