

AGROCIN 84 SELECTIVITY AND

TOXICITY IN AGROBACTERIUM

by

PETER JEFFREY MURPHY

Departments of Plant Pathology
and Agricultural Biochemistry
Waite Agricultural Research Institute,
University of Adelaide
South Australia.

A thesis submitted to The University of Adelaide
in fulfilment of the requirements for the
degree of Doctor of Philosophy
January 1981

2.13 In vitro DNA polymerase activity - E. coli polymerase I

2.14 Eucaryotic DNA polymerases from Ehrlich ascites cells

agrobacteria

14

15

16

CHAPTER 3. PREPARATION OF COMPOUNDS

3.1 General	18
3.2 Preparation of agrocin 84	18
3.2.1 Production of agrocin 84	18
3.2.2 Purification of agrocin 84	19
3.2.3 Preparation of [³² P]agrocin 84	20
3.3 Preparation of breakdown products of agrocin 84	21
3.3.1 Preparation of non-labelled compounds	21
3.3.2 Preparation of [³² P]fragment 3	22
3.3.3 Preparation of [³² P]fragment 9	22
3.4 Compound X (CX)	23
3.4.1 Isolation of Compound X (CX)	23
3.4.2 Characterization of CX	25
3.5 Preparation of 5'(3'-d-ara-)ATP	26
CHAPTER 4. THE BASIS OF SELECTIVITY OF AGROCIN 84	
4.1 Introduction	28
4.1.1 Aim of the study	28
4.1.2 Gram negative bacterial cell walls	29
4.1.3 Bacterial transport	29
4.1.3.1 Transport of low molecular weight compounds	30
4.1.3.2 Transport of high molecular weight compounds	33
4.1.4 Bacteriocin specificity of colicins	33
4.2 Results	36
4.2.1 Plasmids of <i>A. radiobacter</i> strains K57, K57A and K57A Agr ^r	36
4.2.2 Uptake of [32P]agrocin 84	37
4.2.3 An agrocin 84 binding protein	40
4.2.3.1 Cellular localization of the agrocin 84 binding protein in A. radiobacter	41

4.2.3.1.1 Preparation of periplasmic proteins from A. radiobacter	41
4.2.3.1.2 Equilibrium dialysis binding of agrocin 84 to periplasmic and cytoplasmic protein fractions	43
4.2.3.2 Reversibility of agrocin 84 binding to the binding protein	43
4.2.3.3 Characterization of the agrocin 84 binding protein	44
4.2.4 The role of the glucofuranosyl moiety of agrocin 84 in selectivity	45
4.2.4.1 Biological activity of a nucleotide fragment of agrocin 84 which does not contain the gluco- furanosyl moiety:- fragment 3	45
4.2.4.2 Reduced uptake of [32P]fragment 3	45
4.2.4.3 Inhibition of [P]agrocin 84 uptake by a nucleotide fragment of agrocin 84 which contains the glucofuranosyl moiety:- fragment 9	46
4.2.4.4 Inhibition by fragment 9 of [32] Plagrocin 84 binding to the agrocin 84 binding protein	47
4.2.4.5 Uptake of [32P]fragment 9	48
4.2.5 In vivo breakdown of agrocin 84	48
4.2.6 Effect of agrocinopine A on [32 P]agrocin 84 uptake	49
4.3 Discussion	49
4.3.1 The plasmid coded periplasmic agrocin 84 binding protein transport system	49
4.3.2 Recognition by the glucofuranosyl moiety of agrocin 84	52
4.3.3 In vivo breakdown of agrocin 84	54
4.3.4 The natural substrate for the agrocin 84 transport system	55
CHAPTER 5. THE BASIS OF TOXICITY OF AGROCIN 84	
5.1 Introduction	57
5.1.1 Aim of the study	57
5.1.2 The toxic action of agrocin 84	57
5.1.3 Inhibition by nucleoside analogues	59

5.1.4 Is agrocin 84 a potential chain terminator?	61
5.1.5 Nucleoside analogues as chemotherapeutic agents	62
5.2 Results	64
5.2.1 Effect of agrocin 84 on A. radiobacter DNA	64
5.2.1.1 Incorporation of 5'(3'-d-ara-)AMP into A. radiobacter DNA	64
5.2.1.2 Effect of 5'(3'-d-ara-)ATP on A. radiobacter polymerase activity	66
5.2.1.3 Sucrose density gradient investigations on the effect of agrocin 84 on A. radiobacter DNA synthesis in vivo	67
5.2.2 Biological activity of agrocin 84 fragments and Compound X	68
5.2.3 Effect of 5'(3'-d-ara-)ATP on other DNA polymerases	69
5.2.3.1 E. coli DNA polymerase I	69
5.2.3.2 Eucaryote DNA polymerases	70
5.3 Discussion	71
5.3.1 Mode of inhibition by agrocin 84	71
5.3.2 DNA repair	74
5.3.3 The role of agrocin 84 and breakdown products as chemotherapeutic agents	75
CHAPTER 6. GENERAL DISCUSSION	78
APPENDICES	
A Culture media and buffers	82
B Calculation of Ki	8
C Publications	86
BIBLIOGRAPHY	8

SUMMARY

This study encompasses a biochemical investigation of the unique mode of toxicity and selectivity of the first known nucleotide bacteriocin, agrocin 84.

Sensitivity to agrocin 84 has been shown to be a consequence of its uptake into the bacterial cytoplasm of a susceptible strain of Agrobacterium radiobacter. Only the pathogenic strain which harbours a tumour-inducing (Ti)-plasmid transports agrocin 84 into the cytoplasm and is thus sensitive. The non-pathogenic parent recipient strain devoid of the Ti-plasmid does not transport agrocin 84 and is accordingly not sensitive to its toxic action.

This uptake process is by a high affinity active transport system. A Ti-plasmid coded periplasmic agrocin 84 binding protein is an important component of this transport system. Only bacteria sensitive to agrocin 84 have this binding protein.

The N ⁶-glucofuranosyl moiety of agrocin 84 determines the strain specificity of agrocin 84 for the pathogenic bacteria. When this moiety is absent the fragment formed is toxic, at low levels, to both pathogenic and non-pathogenic bacteria.

It is the N^6 -glucofuranosyl moiety which is recognized by the agrocin 84 binding protein. Recognition by this moiety results in transport into pathogenic bacteria by the high affinity transport system. Without this moiety uptake is non-specific and is much slower, presumably by diffusion.

A compound from crown gall tissue, agrocinopine A (Ellis J.G., Ph.D. student, Adelaide University) inhibited the uptake of agrocin 84

and is implicated as a possible natural substrate for the agrocin 84 transport system.

Studies to determine the basis of toxicity of agrocin 84 were also instigated and a key finding was that the 5'(3'-d-ara-)AMP core of agrocin 84 could be isolated from bacterial DNA. The incorporation of this component would by necessity result in DNA chain termination. The *in vivo* metabolism of agrocin 84 to a toxic component and a role at the DNA polymerase level is discussed.