

Novel Multiparameter Flow Cytometry Techniques for the Detection of Leukaemia Associated Phenotypes and Minimal Residual Disease Monitoring In Acute Myeloid Leukaemia

# Adhra Al-Mawali

Research conducted at Institute of Medical and Veterinary Science

Divisions of Haematology and Human Immunology

Hanson Institute

Adelaide, South Australia

A thesis submitted in fulfilment of the requirement for the degree of Doctor of Philosophy in the School of Medicine at the University of Adelaide

March 2008

# **TABLE OF CONTENTS**

DECL	_ARATION	
ACKI	NOWLEDGMENTS	
PUBL	LICATIONS ARISING FROM THIS THESIS	VII
ABBF	REVIATIONS	IX
CONF	FERENCE PRESENTATIONS	XII
AWA	RD ARISING FROM THIS THESIS	XIV
DEDI	CATION	XV
ABST	TRACT	XVI
CHAF	PTER 1: GENERAL INTRODUCTION	1
1.1	SIGNIFICANCE OF THE RESEARCH	1
1.2	INTRODUCTION	2
1.3	LEUKAEMIA	2
1.4	WHAT IS AML?	2
1.5	EPIDEMIOLOGY	
1.5.1	1 Genetic Predisposition	3
1.5.2	1	
1.5.3 1.5.4	1 2	
1.5.5		
1.6	PATHOPHYSIOLOGY	4
1.7	DIAGNOSIS	
1.7.1 1.7.2	$\varepsilon$	
1.8	CHEMOTHERAPEUTIC TREATMENT	7
1.9	OUTCOMES	10
1.10	PROGNOSTIC FACTORS IN AML AT DIAGNOSIS	11
1 10		

1.10.2	Cytogenetics and molecular genetics	
1.10.3	Molecular markers	
1.10.3 1.10.3		
1.10		
1.10.		
1.10.3	· · · · · · · · · · · · · · · · · · ·	
1.10.	6	
1.10.4	Secondary leukaemia and advanced disease	
1.10.5	White blood cell count	
1.10.6 1.10.7	ImmunophenotypingFAB Classification	
1.10.7	Gene expression profiles.	
1.11 PI	ROGNOSTIC FACTORS IN AML AFTER INDUCTION THERAPY	18
1.11.1	Response to first induction	
1.11.2	Minimal residual disease as a predictive factor for relapse	19
1.12 W	HAT IS MINIMAL RESIDUAL DISEASE?	20
	ETHODS AVAILABLE FOR THE DETECTION OF MRD	
1.13.1	Morphology	
1.13.2 1.13.3	Cell culture assaysCytogenetic analysis	
1.13.3	Fluorescence in situ hybridization techniques (FISH)	
1.13.5	Polymerase Chain Reaction (PCR)	
1.13.6	Flow cytometry and immunophenotyping	
	ACTORS THAT INFLUENCE FLOW CYTOMETRY FOR DETECTING M	
1.14.1	Frequency of LAPs	
1.14. 1.14.	1 1 7	
1.14.2	1.2 Points for consideration for LAPs detection	
1.15 Al	DVANTAGES AND DRAWBACKS OF MFC FOR DETECTING MRD	29
1.16 CI	LINICAL STUDIES IN MRD DETECTION	31
1.17 NI	EWER APPROACHES: LEUKAEMIC STEM CELLS AND MARKERS	32
1.17.1	Understanding AML stem cells will lead to more effective therapies	32
1.18 FU	UNCTIONAL AND PHENOTYPIC CHARACTERIZATION OF LEUKAEN	MIC
	LLSLLS	
1.18.1	Functional characterization of LSC	33
1.18.2	Phenotypic characterization of LSC	34
	ARGETS OF AML THERAPY DETECTABLE IN LEUKAEMIC STEM CH	
1.19.1	Targets expressed on the surface of AML cells	
1.19.2	Intracellular targets of AML therapy	
1.20 H	YPOTHESES AND AIMS OF THE RESEARCH	41
1.20.1	Hypotheses of the research:  Aims of the research	
1.20.2		
1.21 H	OW THE THESIS AIMS CONTRIBUTE TO THE CURRENT KNOWLED	GE? .41
CHAPTE	R 2: MATERIALS & METHODS	44
2.1 M	ATERIALS	44

2.1.1 PATIENT POPULATION	
2.1.1.1 Sample size calculation	
2.1.1.1.1 Understanding the Sample Size Table	45
2.1.1.1.2 Choosing the sample size	46
2.1.1.2 Patient samples	47
2.1.1.2.1 Selection criteria	
2.1.1.2.2 Study conduct	
2.1.1.2.3 Study parameters	
2.1.1.3 Normal bone marrow samples	
2.1.1.4 Regenerating bone marrow samples	
2.1.1.5 Chemotherapeutic treatment	48
2.1.2 SUPPLIERS OF COMMONLY USED REAGENTS	
2.1.3 IMMUNOFLUORESCENCE REAGENTS	51
2.1.3.1 FACS Fix	
2.1.3.2 FCS - Foetal Calf Serum	
2.1.3.3 Lysis Solution	
2.1.3.4 Reagent A - Erythrocyte Lysis Reagent for Q-Prep System	
2.1.3.5 Reagent B - Leukocyte Stabiliser Reagent for Q-Prep System.	52
2.1.3.6 Reagent C - Leukocyte Fixative Reagent for Q-Prep System	
2.1.3.7 1% Sodium Hypochlorite (W/V)	
2.1.3.8 White Cell Fluid (WCF)	
2.1.3.9 MACS CD34 <sup>+</sup> Buffer (PBS/0.5% BSA/2mM EDTA)	53
2.1.3.10 Thawing solution	53
2.1.3.11 Monoclonal Antibodies (MoAbs)	
2.1.4 DNA MANIPULATIONS REAGENTS	60
2.1.4.1 TBE Buffer	
2.1.4.1.1 5 x Stock Tris-Borate (TBE) Buffer	
2.1.4.1.2 TBE Buffer - Working Solution	
2.1.4.2 Agarose gel mix	60
2.1.4.2.1 2% Agarose Gel	
2.1.4.3 Loading Buffer	
2.1.4.4 Molecular Weight Marker pUC19	
2.1.4.4.1 Working Solution for pUC19	
2.1.4.5 Ethidium Bromide	
2.1.4.5.1 Ethidium Bromide Stock Solution	
2.1.4.5.2 Ethidium Bromide Working Solution	
2.1.4.6 dNTP Reagent	
2.1.4.6.1 dNTP Stock Reagent 100 mM	
2.1.4.6.2 dNTP Working Solution 2 mM of each dNTP	
2.1.4.7 Taq DNA polymerase kits	63
2.2 METHODS	
2.2.1 MORPHOLOGY AND CYTOCHEMISTRY	
2.2.2 CYTOGENETICS	
2.2.3 IMMUNOPHENOTYPING	64
2.2.3.1 Cryopreservation of cells	64
2.2.3.2 Thawing cryopreserved cells	
2.2.3.3 Specimen processing	64
2.2.3.3.1 Cell count	
2.2.3.3.1.1 Cell count using Haematocytometer	65
2.2.3.3.1.2 Cell count using Epics® - XL-MCL and Cytomics® F	C500 flow cytometer
65	
2.2.3.3.1.3 Cell count data interpretation	65
2.2.3.3.2 Specimen dilution	
2.2.3.4 Staining technique	
2.2.3.4.1 Manual staining	
2.2.3.4.2 Lysis Method	
2.2.3.4.3 Surface and cytoplasmic staining	
2.2.3.4.4 Blocking techniques	
2.2.3.4.5 Viability staining	

2.2.3.5	Method of Antibody evaluation	
2.2.3.5.		
2.2.3.5.2	1 · · · · · · · · · · · · · · · · · · ·	
2.2.3.5.		
2.2.3.5.4	r r	
2.2.3.5.		
2.2.3.5.0		
2.2.3.6	Antibody dilution list	
2.2.3.7	Antibody panels	
2.2.3.8	Compensation	
2.2.3.8.		
2.2.3.8.2	1	
2.2.3.8.3 2.2.3.9	3 BD CompBeads	
2.2.3.9.		
2.2.3.9.		
2.2.3.9.		
2.2.3.9.4	4 After a 2nd run: Establishment of a LAP	84
2.2.3.9.		
2.2.3.10	Gating strategy for MRD	
2.2.3.10		
2.2.3.10	0.2 Analyses of MRD	85
2.2.3.10	Data reporting and interpretation	85
2.2.3.11	Fluorescence activated cell sorting.	
	IUNO-FLUORESCENCE CONFOCAL MICROSCOPY ASSAYS	
	LECULAR TECHNIQUES	
2.2.5.1	Purification of DNA	
2.2.5.1.		
2.2.5.1.2	T -	
2.2.5.2	Calculation of DNA Concentration and Purity by Spectrophotometer	
2.2.5.3	Electrophoresis of DNA	
2.2.5.4	Size determination and quantitation of DNA fragments	
2.2.5.5 2.2.5.6	Primers used for PCR and sequencing	
2.2.5.7	Sequencing the DNA	
	IUNOMAGNETIC ASSAYS	
2.2.6.1	Isolation of CD34 <sup>+</sup> Cells: MACS Separation	
	TISTICAL ANALYSIS	
PHENOTYP	: THE PRESENCE OF LEUKAEMIA-ASSOCIATED ES IS AN INDEPENDENT PREDICTOR OF INDUCTION ACUTE MYELOID LEUKAEMIA	
3.1 ABSTR	RACT	102
3.2 INTRO	DUCTION	104
	TS	
	_ samples	
	taemia-associated phenotypes	
3.3.2.1	Lineage infidelity	
3.3.2.2	Asynchronous antigen expression	
3.3.2.3	Absence of lineage specific antigens	108
	presence of leukaemia-associated phenotypes is associated with a poor response	
	rapy	
3.3.4 Leuk	aemia-associated phenotypes may predict relapse of AML	114
3.4 DISCU	SSION	119

PHEN FOR T	OTYPE HE DE	INCIDENCE OF LEUKAEMIA ASSOCIATED S IN ACUTE MYELOID LEUKAEMIA PATIENTS: A BASIS SIGN OF SPECIFIC FIVE-COLOUR STAINING TO BE US AL RESIDUAL DISEASE INVESTIGATION	ED
4.1	ABSTR	ACT	.124
4.2	INTRO	DUCTION	.126
4.3	RESUL	ΓS	.128
4.3.1	Cluste	ering of bone marrow lineages using CD45 gating strategy	. 128
4.3.2		y of CD45/SSC gating in frozen samples	
4.3.3		ession of different MoAbs	
	3.3.1 3.3.2	CD45HLA-DR	
	3.3.3	CD34/CD38	
	3.3.4	CD117	
	3.3.5	CD33	
4.3	3.3.6	CD13	. 143
4.3	3.3.7	CD14	
	3.3.8	CD15	
	3.3.9	CD56	
	3.3.10	CD2	
	3.3.11	CD7	
	3.3.12 3.3.13	CD19	
	3.3.14	CD11b	
	3.3.15	CD64	
	3.3.16	CD65	
	3.3.17	CD123	
4.3	3.3.18	Cytoplasmic MPO	
4.3	3.3.19	Nuclear TdT	. 146
		fication of leukaemia-associated phenotypes	
		Lineage infidelity	
	3.4.2	Asynchronous antigen expression	
	3.4.3	Absence of lineage specific antigens	
4.3.5	Aberr	rant markers expression in normal and regenerating bone marrow cells	. 155
		tification of differences in cells carrying aberrant immunophenotypes between	160
4.3.7	Samples,	and normal and regenerating bone marrowstigation of the 'in vitro' sensitivity of flow cytometry immunophenotyping for	. 100
		RD	163
	3.7.1	Dilution of AML samples in normal and regenerating bone marrow	
	3.7.2	Assessment of linearity	
4.3	3.7.3	Assessment of reproducibility	
4.3.8	Appe	arance of LAPs using Immuno-Fluorescence Confocal Microscopy	
4.4	DISCUS	SSION	175
CHAP ACUT	TER 5: E MYE ELAPSI ABSTR	DETECTION OF MINIMAL RESIDUAL DISEASE IN LOID LEUKAEMIA IDENTIFIES PATIENTS WITH HIGH RIEST CONTROLLED TO THE CONTROLLED	SK 179
5.3	RESUL	ΓS	. 182
5.3.1	Clinic	al Characteristics	. 182

5.3.2		
	nemic cells in terms of relapse status	
5.3.3		
5.3.4		
5.3.5	1	
5.3.6		
5.3.7	e e e e e e e e e e e e e e e e e e e	
5.3.8		
5.3.9	Prognostic impact of log difference in OS	206
5.4	DISCUSSION	210
ACUT	TER 6: CHARACTERISTICS AND PROGNOSIS OF ADULT EMYELOID LEUKAEMIA WITH INTERNAL TANDEM ICATIONS IN THE <i>FLT3</i> GENE	216
6.1	ABSTRACT	216
6.2	INTRODUCTION	217
6.3	RESULTS	220
6.3.1		
6.3.2		
6.3.3	1	
6.3.4		
	3.4.1 Correlation to cytogenetics	
	3.4.2 Correlation with leukocyte count in FAB subgroups	
	3.4.3 Correlation with percentage of blasts at diagnosis	
	3.4.4 Correlation with FAB subtype	
	3.4.5 Correlation with Age and sex	
- 4		•••
6.4	DISCUSSION	
6.4.1	rr	
	4.1.1 Incidence of FLT3/ITD	
	4.1.2 The impact of FLT3/ITDs on patient prognosis	230
TAND	PTER 7: THE ONCOGENIC EVENTS OF FLT3 INTERNAL DEM DUPLICATION HAPPEN AT A STAGE OF STEM CELLS TO	
POSS	SESS IL-3 ALPHA RECEPTOR (CD123)	237
7.1	ABSTRACT	237
7.2	INTRODUCTION	239
7.3	RESULTS	242
7.3.1		
7.3.2		
7.3.3		tion
7.3.4	Characterization and phenotype of leukaemic stem cells	
7.3.5		
7.3.6	Optimisation of the PCR technique for low DNA concentrations	249
7.3.7		
7.3.8		
7.3.9	Detection of FLT3/ITD in the sorted population of LAP positive AML patient	254
7.4	DISCUSSION	258

		8: SUMMARY, GENERAL DISCUSSION, AND	263
8.1	SUM	MARY	263
8.2	GENI	ERAL DISCUSSION	267
8	.2.1 Cli	nical applications of MRD in AML	267
	8.2.1.1		
	8.2.1.2	Future directions for MRD detection	271
	8.2.1.3	MRD detection using five-colour MFC	271
8	.2.2 Pot	ential limitations for MRD detection using immunophenotyping	
8		nts for consideration	
8	.2.4 CD	123 as a marker for LSC and a single standardised marker for MRD studies	276
8	.2.5 Pro	gnostic value of FLT3/ITD in AML	277
8	.2.6 Do	es FLT3/ITD mutation occur at the level of the leukaemic stem cells?	278
8.3	CON	CLUDING REMARKS	279
СН	APTER	9: BIBLIOGRAPHY	283
PU	BLICAT	IONS	324

# **DECLARATION**

This thesis contains no material which has been accepted for the award of any other degree or diploma in any university or other tertiary institution and, to the best of my knowledge and belief, contains no material previously published or written by another person, except where due reference has been made in the text.

I give consent to this copy of my thesis, when deposited in the University Library, being available for loan and photocopying.

Adhra Al-Mawali			
Signed:	Date:		

## **ACKNOWLEDGMENTS**

In the name of Allah, the most gracious, the most merciful. Looking at the whole PhD thesis, it is no longer merely about me, but it is about those who gave their effort, time and made this work possible.

Thanking every one for all their help during my PhD isn't an easy task, as there have been many people over the course of my studies that have gone out of their way to help. And I know that without their encouragement and support, I wouldn't have got this far.

First of all, I wish to express my sincere appreciation to my supervisors, Dr Ian Lewis and Dr David Gillis for their help, ideas and patience throughout this research. I am thankful to them for the opportunity to carry out my PhD studies under their supervision, their continuous guidance, support and the hours they spent with me every week questioning and challenging my arguments, asking questions, editing the (many!) drafts that came their way and guiding me through this PhD thesis. I am grateful to Dr David Gillis who taught me every thing I need to know in flow cytometry. Particular thanks also go to Dr Ian Lewis for supporting me financially in my last year of PhD with University Tuition fees. In addition, I would also like to thank the following:

## From Division of Human Immunology, IMVS, Australia

Mr John Bahnisch for being always kind to answer my questions and Ms Joy Mundy for her enormous amount of advice, support and continuous assistance with flow cytometry, given answers to my endless questions, and regularly dropped by to check that things were going ok throughout my PhD.

Ros Foster and Anne-Marie Kennedy for their excellent technical assistance and friendship.

From Molecular Lab, Heather James, Pina Morelli, Shree Brozyna, and Cassie Hill for their encouragement, help with PCR and sequencing DNA.

From Autoimmune and chemistry lab, Zack Browning for his jokes and sense of humour, John Cassidy, Grahm Brierley, Hazel Willis, Di Gillham, Jeniffer Nicholas, Benno Bruin, and Leeanne Koziren for their friendship and help.

There were many times I was so engrossed in my experiments or writing that I was oblivious to the daily dramas of a busy diagnostic laboratory, I thank all of them for always giving me the time to do this work.

Dr Pravin Hissaria for his continuous support by giving his valuable comments, discussing my research and reading my papers and giving his advices.

## From Division of Haematology, IMVS, Australia

Professor L Bik To for his valuable comments on my papers, discussing my research and his continuous encouragement.

Dr Marion Roberts for providing me with Morphology Cytogenetics Immunophenotyping (MCI) data for AML patients.

From Haemopoietic Expansion Laboratory, Pam Dyson, Sonia Young, Rick Tochetti, and Smita Hiwase who willingly shared their knowledge and skills to help me with this study.

Alan Bishop, Kate Pilkington, Andrew Macintyre for their help with flow cytometry and sorting. Ghafar Sarvestani for his help with Confocal Microscopy. Dr David Ross and Dr Devendra Hiwase for discussing my results and analyses especially in the MRD part. Tony Cambareri for his advice, happily answering all my questions.

## From Department of Public health, University of Adelaide, Australia

Big thank you also goes to Dr. Nancy Briggs, Lisa Yelland and Dr. John Field for their enormous amount of help with statistical analysis and for answering and discussing my endless questions in statistics.

## From Flinders Medical Centre, Australia

Dr Peter Macardle for giving me the opportunity to do training in his lab. Shree Bailey for helping me with compensation especially when I was just started.

## From ITS, University of Adelaide, Australia

Ms Helen Foster for helping me with many technical issues in the final formatting of the thesis.

## From Division of Molecular Pathology, IMVS, Australia

Glenice Cheetham for her help with sequencing the DNA.

#### From Sultanate of Oman

I wish to thank Directorate General of Training and Scholarship for funding this research and Dr Muhanna Al-Muslahi (Head of Haematology Department, Royal Hospital, Muscat, (Oman) for his guidance and support.

## From St. Jude Children's Research Hospital, Memphis, USA

Prof. Dario Campana for his valuable advices on MRD studies when I was just started. Prof Dario Campana, MD PhD is a Member, Haematology-Oncology and Pathology, St. Jude Children's Research Hospital, Professor of Paediatrics, University of Tennessee College of Medicine.

## From Purdue Cytometry Mailing list, USA

All the members of Purdue cytometry mailing list for answering my questions about MRD and immunophenotyping. From this list, I have learnt a lot about flow cytometry and I am still learning.

## For the anonymous examiners who reviewed this thesis

I would like to thank you for your effort and time in bringing this work to life.

## Very special "Thank You"

To my family, friends in Oman and friends who I met in Adelaide during the PhD program for their continuous support and encouragement.

I am grateful for all who have helped me to get to this point, directly or indirectly, thank you very much.

## Lastly but most importantly

However, I owe the most to my husband Essa. This research succeeded largely due to his tolerance, love, support, understanding and harassment to get this done! Essa: words cannot express my appreciation for your support, companionship

and love throughout this study. Thank you Essa for being my inspiration and motivation in life. Thank you for being always there when I needed you most.

"Obstacles cannot crush me; every obstacle yields to stern resolve"

Leonardo da Vinci

"I don't know what I may appear to the world, but to myself I seem to have been only like a boy playing on the sea-shore, and diverting myself in now and then finding a smoother pebble or a prettier shell than ordinary, whilst the great ocean of truth lay all undiscovered before me."

**Sir Isaac Newton (1642- 1727)** 

## PUBLICATIONS ARISING FROM THIS THESIS

- The Presence Of Leukaemia-Associated Phenotypes Is An Independent Predictor Of Induction Failure In Acute Myeloid Leukaemia. Al-Mawali A, L Bik T, Gillis D, Hissaria P, Mundy J, Lewis I. International Journal of Laboratory Hematology (2008). In Press. Early online published article (accepted 17/9/07)
- Incidence, Sensitivity and Specificity of Leukaemia Associated Phenotypes
  in Acute Myeloid Leukaemia Patients using Specific Five-Colour
  Multiparameter Flow Cytometry. Al-Mawali A, Hissaria P, Gillis D, Lewis
  I. American Journal of Clinical Pathology (2008). In Press. (accepted
  21/12/07).
- 3. The Oncogenic Events Of FLT3 Internal Tandem Duplication Happen At A Stage Of Stem Cells That Possess IL-3 Alpha Receptor. Al-Mawali A, Gillis D, Thomas D, Ramshaw H, Lopez A, Lewis I. (2008). Manuscript in preparation.
- Detection Of Minimal Residual Disease Post Induction In Acute Myeloid Leukaemia Identifies Patients With High Risk Of Relapse. Al-Mawali A, Lewis I, Gillis D. Cytometry Part B: Clinical Cytomtry (2008). Manuscript submitted (3/3/2008).

5. The Role of Multiparameter Flow Cytometry for Detection of Minimal Residual Disease in Acute Myeloid Leukaemia. **Al-Mawali A**, Gillis D, Lewis I. (2008). *Manuscript submitted*.

## **ABBREVIATIONS**

Ab Antibody

ALL Acute lymphoid leukaemia

AlloSCT Allogeneic stem cell transplants

AML Acute myeloid leukaemia

APL Acute promyelocytic leukaemia

AutoSCT Autologous stem cell transplants

BM bone marrow

BSA bovine serum albumin

CBF core binding factor

CD Cluster of differentiation

CD34 (sialomucin = haemopoietic progenitor cell antigen-1, HPCA-1)

CLL-1 C-type lectin–like molecule 1

CML Chronic myeloid leukaemia

CR Complete remission

DMSO Dimethyl sulphoxide

DNA Deoxyribonucleic acid

DNA Deoxyribonucleic acid

ECD Phycoerythrin-Texas Red (PE-TR)

EDTA Ethylenediaminetetra acetic acid

FAB French- American- British

FACS Flouresence activating cell sorter

FC Flow cytometry

FCS Foetal calf serum

FITC Fluorescein isothiocyanate

FLT3 Fms like tyrosine kinase3

FLT3/ITD Fms like tyrosine kinase3 – internal tandem duplication

HSC Haemopoietic stem cell

IL-3α Interleukin 3 alpha receptor

IMDM Iscove's modification of dulbecco's medium

LAP leukaemia associated phenotype

Lin– lineage specific marker negative

LSC Leukaemic stem cells

MACS Magnetic activated cell sorting

MCF Mean cell fluorescence

MDR1 Multidrug resistance protein

MFC Multiparametric flow cytometry

MFI Mean fluorescence Intensity

MNC Mononuclear cells

MoAb Monoclonal antibody

MRD Minimal residual disease

MUD Matched unrelated donor

ND Not done

NK Natural killer

NOD/SCID Non obese diabetic –severe combined immunodeficient

OS Overall survival

PB Peripheral blood

PBS Phosphate-buffered saline

PC5 Cy5 coupled to R-Phycoerythrin.

PC7 Cy7 coupled to R-Phycoerythrin.

PCR Polymerase chain reaction

PE Phycoerythrin

PFA Paraformaldehyde

RFS Relapse free survival

RNA Ribonucleic acid

ROC Receiver operating characteristics

RT Room temperature

SCID Severe combined immunodeficiency

WBC White blood cell

WHO World health organisation

WT Wildtype

## **CONFERENCE PRESENTATIONS**

#### New Direction in Leukaemia Research NDLR 2006 Conference

Sunshine Coast, Brisbane from 2 – 5 April 2006

"Leukaemia-Associated Phenotypes in adult Acute Myeloid Leukaemia at diagnosis: their characteristics, clinical significance and usefulness as prognostic markers".

A. Al-Mawali, J. Mundy, D. Gillis, L. B. To, I. Lewis

# The Australian Society for Medical Research, ASMR , SA Scientific Meeting 2006

Adelaide, South Australia 2-June 2006

"The presence of Leukaemia-Associated Phenotypes in adult Acute Myeloid Leukaemia at diagnosis is of prognostic significance"

A. Al-Mawali, J. Mundy, D. Gillis, L. B. To, I. Lewis

## Eleventh Adelaide Blood Club Meeting with HSA and TIGSA

Sunset Cove, Yankalilla, SA from 8-10 September 2006

"The Presence of Leukaemia-Associated Phenotypes is an Independent Predictor of Poor Prognosis in Acute Myeloid Leukaemia"

A. Al-Mawali, J. Mundy, D. Gillis, L. B. To, I. Lewis

# The Australian Society for Medical Research, ASMR, SA Scientific Meeting 2007

Adelaide, South Australia 6-June 2007

"The Oncogenic Events Of FLT3 Internal Tandem Duplication Happen At A Stage Of Stem Cells That Possess IL-3 Alpha Receptor"

## A. Al-Mawali, D. Gillis, I. Lewis

## 30th Annual Meeting of the Australasian Flow Cytometry Group AFCG 2007

Telstra Dome, Melbourne, Victoria September 16<sup>th</sup>-19<sup>th</sup>, 2007.

"Incidence Of Leukaemia Associated Phenotypes In Acute Myeloid Leukaemia Patients: A Basis For The Design Of Specific Five-Colour Staining To Be Used For Minimal Residual Disease Investigation" AND

"The Oncogenic Events Of FLT3 Internal Tandem Duplication Happen At A Stage Of Stem Cells That Possess IL-3 Alpha Receptor"

A. Al-Mawali, D. Gillis, I. Lewis

**Research Expo 2007.** University of Adelaide, Adelaide, October 23<sup>rd</sup>, 2007.

"Incidence Of Leukaemia Associated Phenotypes In Acute Myeloid Leukaemia Patients: A Basis For The Design Of Specific Five-Colour Staining To Be Used For Minimal Residual Disease Investigation",

"The Oncogenic Events of FLT3 Internal Tandem Duplication Happen At A Stage
Of Stem Cells That Possess IL-3 Alpha Receptor" AND

"Detection of Minimal Residual Disease in Acute Myeloid Leukaemia Identifies Patients with High Risk of Relapse and May Play A Role in Post Induction Treatment Stratification".

A. Al-Mawali, D. Gillis, I. Lewis

# **AWARD ARISING FROM THIS THESIS**

The Australian Society for Medical Research: "The 2007 Clinical Research Poster Presentation Award" for the most outstanding presentation of clinical research at the South Australian ASMR Scientific Meeting on June 6 2007 on "The Oncogenic Events Of FLT3 Internal Tandem Duplication Happen At A Stage Of Stem Cells That Possess IL-3 Alpha Receptor".

# To my grandmother: Thuraya

Who passed away when I was in the second year of my PhD program. She was always very kind, encouraging, supportive and praying for me all the time.

# To my father: Hilal

You have taught me so much throughout my life. Your examples of determination and ambition have encouraged me to always strive to be the very best I can be.

Father: the best is yet to come!

# To my mother: Zayana

You never stopped believing in me and encouraging me to always give of my best.

Mum: This is a small gift.

# To my son: Ezzaldeen

You have grown so much during the time in which I have been studying for this PhD, and I feel that I have missed so many special moments of your childhood while doing my PhD. I promise to be there for you always from now on.

# Primarily to my husband: Essa

For your endless support, love and encouragement through the hardest days of my PhD, especially for your patience during the most difficult times when I had to work on weekends and remain at work for very long

hours. Essa: I dedicate this PhD to you

## **ABSTRACT**

Despite high remission rate in acute myeloid leukaemia (AML) after chemotherapy, relapse of the underlying disease remains a major challenge and one of the most frequent causes of treatment failure. In this study, the presence of leukaemia-associated phenotypes (LAPs) was first studied retrospectively using our standard diagnostic protocol with 3-colour flow cytometry. LAPs were present in 54 (64%) of 84 AML patients analysed between 2002 to 2004. The presence of LAPs was correlated with failure to respond to induction chemotherapy (p <0.05) in univariate analysis. Presence of LAPs was shown to be an independent predictor for failure to respond to induction chemotherapy with a relative risk ratio of 1.6 (p < 0.05, 95% CI, 1.0-2.6) in multivariate analysis.

Subsequently, in a prospective study, we used 5-colour multiparametric flow cytometry (MFC) for detection of LAPs to determine if LAPs could be detected in a greater proportion of leukaemic patients and minimal residual disease (MRD) detection could therefore be applied in more patients. In 54 consecutive, newly diagnosed AML patients from 2005 to 2007, LAPs were identified in 51 (94%). Thus, MRD studies were potentially applicable to virtually all patients. The sensitivity and specificity of MFC technique was improved by analysing 10 normal and 5 regenerating bone marrows (BM) for the presence of these LAPs and by determining maximum log difference (LD). CD7, CD19, CD2, CD11b and CD56 were the most sensitive and reliable markers for MRD studies. LAPs were rarely detected in either normal or regenerating BMs. Through dilutional experiments from 50% LAPs to 0.001%, it was determined that 1 leukaemic in 10<sup>4</sup> and 10<sup>5</sup> normal cells could be detected using the improved techniques.

Of the 54 patients, 31 received chemotherapy, with 27 achieving complete remission (CR). Two were LAP negative and thus 25 were evaluable for MRD post induction and 22-post consolidation chemotherapy. Detection of MRD >0.15% was able to distinguish between two groups of patients according to relapse status. Although, the number of patients was small, detection of MRD post induction > 0.15% was shown to be an independent predictor of adverse prognosis for both relapse free survival (RFS) and overall survival (OS) in a multivariate analysis [p = 0.037 and 0.026, 95% CI (1.1-20.5 and 1.2-22.2), hazard ratio 4.7 and 5.2 respectively]. Post consolidation, there was a trend for patients with higher MRD values to show shorter RFS (p = 0.06).

MFC using 5-colour allows us to detect LAPs in virtually all AML patients and our preliminary results suggest the technique is a suitable approach for MRD analysis. However, 5-colour MFC is technically challenging, resource intensive, and may not be feasible in a routine diagnostic laboratory. This led us to assess whether we could identify other potential markers for LAPs.

Interleukin-3 alpha receptor- chain IL-3α (CD123) has been suggested to be a marker of leukaemic stem cells (LSC). These cells are thought to be responsible for initiating and maintaining leukaemic cell growth post chemotherapy and hence to give rise to relapse of the disease. Therefore, we analysed 34 AML patients for expression of CD123 in the blast population and defined a population containing leukaemic stem cells using the immunophenotypic markers CD123<sup>+</sup>/CD34<sup>+</sup>/CD38<sup>-</sup>. Thirty-two (94%) of AML patients expressed CD123. We then used a molecular marker to determine whether CD123 expression was confined to the LSC. Thirty-nine patients were screened for the presence of FMS-like tyrosine kinase 3 - internal tandem duplication (FLT3/ITD) as the most common molecular abnormality in

AML patients. Of those, 12 (31%) were FLT3/ITD positive. In seven of them, CD34<sup>+</sup>/CD38<sup>-</sup>/CD123<sup>+</sup> and CD34<sup>+</sup>/CD38<sup>-</sup>/CD123<sup>-</sup> populations were sorted to homogeneity by Fluorescence Activated Cell Sorting (BD FACSAria<sup>TM</sup> Cell Sorter) and tested for FLT3/ITD. In six of seven patients with FLT3/ITD positive AML, we could not detect the mutation in the CD34<sup>+</sup>/CD38<sup>-</sup>/CD123<sup>-</sup> fraction, but the mutation was detected in the CD34<sup>+</sup>/CD38<sup>-</sup>/CD123<sup>+</sup> fraction in all seven patients. This novel finding demonstrates that, the oncogenic event occurs in CD123 positive cells, thus supporting the concept that CD123 is a marker of the LSC in CD123 positive AML. This observation suggests novel treatment approaches employing surface marker CD123-targeting antibodies may be of use in the treatment of AML. In conclusion, we demonstrate that using five-colour MFC improves LAP detection in AML and enables MRD studies using immunophenotyping to be applied to virtually all AML patients. Additionally, it increases the sensitivity of the technique for detecting LAP populations. Moreover, evaluation of MRD post induction chemotherapy is the most sensitive time point for detection of MRD, with MRD levels >0.15% predicting relapse and worse prognosis. As an alternative to using individualised LAPs specific to each patient, CD34+/CD38-/CD123+ cells may in the future serve as a better marker for MRD studies. This marker identifies the putative LSC, which is responsible for regrowth of leukaemia and relapse of the disease. Thus, instead of looking at whole "blast" population which results in huge data analysis and interpretation for the different LAPs which may have different underlying biology, it may be more informative to look at the frequency of LSC after achieving CR using CD34<sup>+</sup>/CD38<sup>-</sup>/CD123<sup>+</sup> as the single LAP for MRD studies.