

**Investigation of immune-suppressive genes expressed**

**by the *Cotesia rubecula* bracovirus (CrBV)**



**Richard V. Glatz, B. Ag. Sc. (Hons.)**

**Department of Crop Protection**

**The University of Adelaide**

**A Thesis Submitted for The Degree of**

**Doctor of Philosophy**

**Discipline of Plant and Pest Science**

**The University of Adelaide**

**January 2004**

# Contents

<b><u>Declaration</u></b>	<b>vii</b>
<b><u>Acknowledgements</u></b>	<b>viii</b>
<b><u>List of figures and tables</u></b>	<b>x</b>
<b><u>Abstract</u></b>	<b>xv</b>
<b><u>Chapter 1: Literature Review</u></b>	<b>1</b>
1.1 Introduction to polydnaviruses	1
1.1.1 Genus <i>Ichnovirus</i>	2
1.1.2 Genus <i>Bracovirus</i>	5
1.1.3 Relationship between <i>Ichnovirus</i> and <i>Bracovirus</i>	11
1.2 The polydnavirus infection-cycle	12
1.2.1 Polydnavirus replication and morphogenesis	14
1.2.2 Transmission of polydnavirus to the parasitised host	18
1.3 Polydnavirus genes and gene expression	22
1.3.1 Polydnavirus gene families	23
1.3.2 Expression of polydnavirus genes in parasitoid wasps	26
1.3.3 Expression of polydnavirus genes in lepidopteran hosts	27
1.3.3.1 Insect immune system components	27
1.3.3.2 Parasitism-induced changes to host cellular immunity	31

1.3.3.3	Parasitism-induced changes to host humoral immunity	35
1.3.3.4	Parasitism-induced changes to host development	38
1.4	<i>Cotesia rubecula</i> and its associated bracovirus (CrBV)	42
1.4.1	CrV1	45
1.4.2	Crp32	49
1.5	Utilisation of baculoviruses for <i>in vitro</i> protein production	52
1.6	Utilisation of baculoviruses as bioinsecticides	53
1.6.1	The pros and cons of wild-type baculovirus bioinsecticides	55
1.6.2	Genetically modified baculoviruses as bioinsecticides	56
1.6.3	Genetically enhanced baculoviruses expressing insect genes	58
1.6.4	Genetically enhanced baculoviruses expressing insect-specific toxins	59
1.7	Summary and project aims	62
<b><u>Chapter 2: Experimental Techniques</u></b>		<b>66</b>
2.1	Isolation and characterisation of bracovirus genes and production of antibodies against gene products	66
2.1.1	Construction and screening of a 6 hour parasitised larval <i>P. rapae</i> cDNA library	66
2.1.2	Southern and Northern hybridization	66
2.1.3	5' amplification of CrV2 cDNA (5' RACE)	67
2.1.4	Cloning and sequencing of 5' RACE CrV2 fragment	67

2.1.5	CrV2 and CrV3 nucleotide sequence analysis	68
2.1.6	CrV2 primer design	68
2.1.7	CrV3 primer design	68
2.1.8	PCR amplification of CrV2 and CrV3	69
2.1.9	Expression of CrV2 and CrV3 in bacteria	69
2.1.10	Purification of recombinant CrV2 and CrV3 proteins	70
2.1.11	Production of antibodies against recombinant CrV2 and CrV3	70
2.2	Characterisation of isolated bracovirus-expressed proteins	71
2.2.1	Computer analyses of CrV2 and CrV3 deduced amino acid sequences	71
2.2.2	Reverse transcription-PCR (RT-PCR) of CrV2 and CrV3	72
2.2.3	Collection of protein samples and Western blotting	72
2.2.4	Fluorescent labelling of CrV2 and CrV3 associated with infected <i>P. rapae</i> haemocytes	73
2.2.5	N-glycosidase digestion of CrV3	73
2.2.6	Characterisation of CrV3-mediated hemagglutination	74
2.2.7	Cloning of <i>P. rapae</i> 18S ribosomal RNA gene fragment	74
2.3	Construction and bioassays of recombinant baculoviruses containing immune-suppressive genes	75
2.3.1	Construction of recombinant baculovirus transfer vector containing CrV1	75

2.3.2	Construction of recombinant baculovirus transfer vector containing Crp32	76
2.3.3	Production of recombinant baculoviruses via transfection of insect cell cultures	76
2.3.4	Isolation of polyhedra and budded virus from infected cell cultures	76
2.3.5	Confirmation of production of desired transcript and protein by recombinant baculoviruses	77
2.3.6	Bioassay of recombinant baculovirus	77
2.4	Generally applied techniques	78
2.4.1	Insect cultures	78
2.4.2	Polydnavirus and genomic DNA isolation	78
2.4.3	Determination of nucleic acid concentrations	78
2.4.4	Removal of hybridised probe from Southern or Northern membrane to allow reprobing	79
2.4.5	Calf Intestinal Phosphatase (CIP) reaction	79
2.4.6	Preparation of competent bacterial cells for heat-shock transformation	79
2.4.7	Heat-shock transformation of competent bacterial cells	80
<b><u>Chapter 3: Results</u></b>		<b>81</b>
3.1	Isolation and characterisation of bracovirus genes and production of antibodies against gene products	81

3.2	Characterisation of isolated bracovirus-expressed proteins	96
3.2.1	Characterisation of CrV2	96
3.2.2	Characterisation of CrV3	106
3.2.3	Comparison of CrV2 and CrV3 expression	121
3.3	Construction and bioassays of recombinant baculoviruses containing immune-suppressive genes	124
	<b><u>Chapter 4: Discussion</u></b>	<b>133</b>
4.1	Isolation and characterisation of CrV2	133
4.2	Isolation and characterisation of CrV3	138
4.3	Comparison of CrV2 and CrV3 expression	147
4.4	Construction and bioassays of recombinant baculoviruses	148
4.5	General discussion: the <i>C. rubecula</i> /CrBV/ <i>P. rapae</i> system	154
	<b><u>Appendix 1: Abbreviations</u></b>	<b>164</b>

<b><u>Appendix 2:</u></b> Published data: Characterisation of a novel protein with homology to C-type lectins expressed by the <i>Cotesia rubecula</i> bracovirus in larvae of the lepidopteran host, <i>Pieris rapae</i>	<b>172</b>
<b><u>References</u></b>	<b>180</b>

# Abstract

The hymenopteran endoparasitoid, *Cotesia rubecula*, employs integrated forms of active and passive immune-suppression in overcoming the defences of its host, *Pieris rapae*, a cosmopolitan pest of cruciferous crops. The immune-suppressive activity arises from a complex of maternally secreted proteins and polydnavirus (PDV) particles, which are injected in a host larva with the parasitoid egg at oviposition. The PDV associated with *C. rubecula* (CrBV) is unusual in that only four main viral genes (CrV1-CrV4) are expressed in *P. rapae* tissues and that expression is transient, remaining at high levels only in the period between four and eight hours post-parasitisation (hpp). Previously, CrV1 was characterised and found to inactivate host haemocytes by causing disruption of their cytoskeleton, leading to abrogation of immune-associated processes such as spreading. In this study, a cDNA library was constructed from parasitised *P. rapae* larvae and screened with total CrBV DNA, leading to isolation of CrV2 and CrV3. The open reading frame of each gene was cloned in a bacterial expression vector and the resultant recombinant proteins were used to produce antibodies against CrV2 and CrV3.

CrV2 has an open reading frame of 960 bp (with no introns) and encodes a glycoprotein of  $\approx 40$  kDa, which is secreted from infected haemocytes and fat body. Comparison of CrV2 deduced amino acid sequence with other known sequences revealed no significant homologies. CrV2 protein was detected in host larvae at 6 hpp, remaining in large amounts for at least a day and was declining by 48 hpp. A putative coiled-coil region at the C-terminus of CrV2 is suspected of involvement in formation of CrV2 trimers that were detected under non-denaturing conditions. CrV2 was visualised within haemocytes in large endosomes at 24 hpp. Although the function of CrV2 remains unclear, it appears to interact with host haemocytes presumably to suppress their immune function.

The CrV3 gene contained an intron and was found to encode a C-type lectin (CTL) homologue, which is secreted from infected host haemocytes and fat body into haemolymph. Two CrV3 monomers (of  $\approx 14$  and  $17$  kDa) were detected in parasitised larvae with the larger monomer being an N-glycosylated form of the smaller monomer. CrV3 dimers and tetramers were also detected *in vivo*. Recombinant CrV3 forms larger complexes and was shown to agglutinate ovine red blood cells, an activity that was  $Mn^{2+}$ - and  $Mg^{2+}$ -dependent but was independent of  $Ca^{2+}$ . CrV3-mediated hemagglutination was inhibited by EDTA but not by biological concentrations of 29 potential ligands tested. Interestingly, CrV3 is similar to invertebrate CTLs associated with humoral defence but not with previously isolated viral lectins. Further, CrV3 homologues were recently detected in bracoviruses from *C. ruficornis* and *C. karyai*, indicating that a novel CTL family is expressed by some *Cotesia*-associated PDVs. CrV3 probably interacts with a soluble host haemolymph component associated with host humoral immune defences.

CrV1 and Crp32 (an immune-suppressive *C. rubecula* calyx protein) were used to produce recombinant *Autographa californica* multiple nucleopolyhedrosis viruses (AcMNPVs), pathogens with putatively enhanced virulence in *P. rapae*. Bioassays were undertaken to investigate the pathogenicity of wild-type AcMNPV in *P. rapae* (previously unreported) and the effect of insertion of Crp32. Although the proportion of larval deaths due to wild-type AcMNPV was significant, the slow rate of mortality indicated that *P. rapae* is only semi-permissive to AcMNPV. Crp32 insertion proved insignificant in terms of the proportion and rate of larval mortality. Given the semi-permissive nature of *P. rapae*, recombinant AcMNPVs expressing immune-suppressive and appropriate reporter genes may be useful for elucidating mechanisms of insect immunity and more specifically, how CrBV acts to subvert these mechanisms in *P. rapae*.