

**Host-parasite interactions in primary and
secondary infections with *Nippostrongylus*
brasiliensis and *Heligmosomoides bakeri***

A thesis submitted for the degree of

DOCTOR OF PHILOSOPHY

as a portfolio of publications

by

Michelle Louise Knott



Discipline of Microbiology and Immunology

School of Molecular and Biomedical Science

The University of Adelaide

South Australia

Australia

August, 2010

TABLE OF CONTENTS

ABSTRACT	i
DECLARATION	iv
ACKNOWLEDGEMENT OF ANY HELP	v
STATEMENTS OF AUTHORSHIP-CHAPTER 2	vi
STATEMENTS OF AUTHORSHIP-CHAPTER 3	vii
STATEMENTS OF AUTHORSHIP-CHAPTER 4	viii
ACKNOWLEDGEMENTS	ix
PUBLICATIONS	xi
COMMONLY-USED ABBREVIATIONS	xii
CHAPTER 1: INTRODUCTION	1
1.1 HELMINTH INFECTIONS: THE GLOBAL PICTURE.....	2
1.2 IMMUNITY TO HELMINTHIC PARASITES	2
1.2.1 An overview of the immune response to helminths.....	2
1.2.2 Expulsion of gastrointestinal helminths	3
1.2.2.1 The IL-4, IL-13, STAT6 signalling pathway.....	4
1.3 IMMUNE CELLS AND KILLING OF HELMINTHS.....	7
1.3.1 Neutrophils	7
1.3.2 Macrophages	8
1.3.3 Mast cells	9
1.3.4 Basophils	10
1.3.5 Eosinophils	11
1.3.5.1 Eosinophil structure, development, differentiation and survival	11

1.3.5.2 Eosinophil migration and recruitment.....	12
1.3.5.3 Eosinophil degranulation	16
1.3.5.4 The role of complement in eosinophil recruitment and function.....	17
1.3.6 Eosinophils and disease.....	18
1.3.6.1 The roles of eosinophils in helminth infections	20
1.4 EXPERIMENTAL MODELS OF HELMINTH INFECTIONS	21
1.4.1 <i>Nippostrongylus brasiliensis</i>	21
1.4.1.1 Parasite life cycle	21
1.4.1.2 Immune responses to <i>N. brasiliensis</i>	22
1.4.1.3 Eosinophils in infection	23
1.4.2 <i>Heligmosomoides bakeri</i>	24
1.4.2.1 Parasite life cycle	24
1.4.2.2 Cytokine responses and parasite expulsion.....	24
1.5 INTRODUCTION TO THIS STUDY	25
CHAPTER 2: IMPAIRED RESISTANCE IN EARLY SECONDARY NIPPOSTRONGYLUS BRASILIENSIS INFECTIONS IN MICE WITH DEFECTIVE EOSINOPHILOPOIESIS	28
LINKAGE TO CHAPTER 2 AND ARTICLE.....	29
CHAPTER 3: THE ROLES OF EOTAXIN AND THE STAT6 SIGNALLING PATHWAY IN EOSINOPHIL RECRUITMENT AND HOST RESISTANCE TO THE NEMATODES NIPPOSTRONGYLUS BRASILIENSIS AND HELIGMOSOMOIDES BAKERI	31
LINKAGE TO CHAPTER 3 AND ARTICLE.....	32
CHAPTER 4: FVB/N MICE ARE HIGHLY RESISTANT TO PRIMARY INFECTION WITH NIPPOSTRONGYLUS BRASILIENSIS	34
LINKAGE TO CHAPTER 4 AND ARTICLE.....	35

CHAPTER 5: DISCUSSION AND CONCLUSION	37
5.1 GENERAL DISCUSSION	38
5.1.1 Summary of the main findings	38
5.1.2 Eosinophils and resistance in helminth infections	40
5.1.2.1 IL-5 and eosinophils in resistance to <i>N. brasiliensis</i>	40
5.1.2.2 Eosinophils and the intestinal phase of infection.....	40
5.1.2.3 Pre-intestinal phase of infection.....	42
5.1.3 Eosinophil recruitment in helminth infections	44
5.1.3.1 Eotaxin, IL-4, IL-13 and STAT6 in eosinophil recruitment	44
5.1.3.2 Eosinophil recruitment in the skin and intestine	45
5.1.3.3 Alternative eosinophil recruitment factors.....	47
5.2 INFECTIONS WITH <i>H. BAKERI</i>	48
5.3 RESISTANCE OF FVB/N MICE TO <i>N. BRASILIENSIS</i> INFECTIONS.....	49
5.3.1.1 Pre-lung and lung phase of infection	50
5.3.1.2 Intestinal phase of infection	51
5.4 <i>H. BAKERI</i> INFECTIONS IN FVB/N MICE.....	52
5.5 CONCLUSIONS.....	54
REFERENCES	56

ABSTRACT

Parasitic helminth infections are a significant problem worldwide. Some helminth species are becoming resistant to current therapies and new forms of treatment and/or vaccines are required. *Nippostrongylus brasiliensis* is a tissue-invasive parasitic helminth that infects rodents, and the lifecycle of this parasite is similar to that of the human hookworms. The aims of this study were to investigate primary and secondary immune responses to *N. brasiliensis*, focusing on the pre-lung phase of infection. The roles of cytokines, chemokines, signalling pathways and leukocytes such as eosinophils were investigated in the skin, lungs and small intestine. The roles of eosinophils were also investigated in primary infections with the intestinal nematode *Heligmosomoides bakeri*.

Interleukin (IL)-5 is important for eosinophil development and maturation and for protection during some helminth infections. Over-expression of IL-5 provides potent protection in primary infections with *N. brasiliensis* in the pre-lung phase of infections. Although mice deficient in IL-5 or eosinophils might therefore be predicted to be more susceptible to *N. brasiliensis*, IL-5-deficient (IL-5^{-/-}) and eosinophil-deficient (Δ dblGATA) mice showed similar infection patterns as wildtype (WT) mice during primary *N. brasiliensis* infections. Intestinal worm and/or egg numbers however were elevated in mice with defective eosinophilopoiesis compared with WT animals. In secondary infections, despite skin inflammatory responses (4 hours p.i.) being similar in WT, IL-5^{-/-} and Δ dblGATA animals, at day 2 p.i., lung larval burdens in the two latter hosts were significantly higher than in the resistant WT controls. However, parasites were expelled from intestines of all mice by day 7 of secondary infections. These data suggest that in the pre-gut phase of secondary infection, IL-5 and eosinophils play an important role in resistance to *N. brasiliensis*. Despite this, eosinophils do not appear to be essential for protection mediated within the gut during secondary infections, even though IL-5 and eosinophils do appear to confer some protection in the gut during primary infections.

Complement is required for the recruitment of eosinophils into the skin in the first 150 minutes of primary infection with *N. brasiliensis* (Giacomin *et al.*, 2008a), however other chemotactic factors appear to be involved after this time. Although eotaxin is

chemotactic for eosinophils in some tissues, the importance of eotaxin and signalling pathways involved in expression of this chemokine has not been previously characterized in *N. brasiliensis* infections. Signal transducer and activator of transcription (STAT)6 is a key transcription factor in the IL-4/IL-13 signalling pathway and these cytokines can induce expression of eotaxin in some tissues. Expulsion of *N. brasiliensis* adult worms during primary infections is profoundly impaired in STAT6-deficient (STAT6^{-/-}) mice. IL-5 Tg mice are highly resistant to primary infections with *N. brasiliensis* and in the current study, it was shown that ablation of eotaxin-1 or STAT6 in IL-5 Tg mice did not impair the strong innate resistance typically seen in the pre-lung phase of *N. brasiliensis* infections. While recruitment of eosinophils to the skin (4 hours p.i.) was reduced in these mice compared with IL-5 transgenic (Tg) mice, protective capacity was preserved in both primary and secondary infections. Further, eotaxin-1^{-/-} single mutant mice were strongly resistant to secondary *N. brasiliensis* infections, with few larvae migrating to the lungs on day 2 p.i. In contrast, STAT6^{-/-}, IL-13^{-/-}, IL-4R α ^{-/-} and IL-13^{-/-}/IL-4R α ^{-/-}-double deficient mice had significantly higher secondary lung larval burdens than WT mice and parasite egg production was prolonged in all of these strains. These data suggest a role for this signalling pathway in protection during the early stages of secondary infections with this parasite. In both primary and secondary infections, eosinophils were recruited to the skin in all gene knock out strains in comparable numbers to those seen in WT mice, and this suggests that alternative eosinophil recruitment pathways may compensate for the absence of these factors. Adding to the extensive work on the intestinal phase of *N. brasiliensis*, this work clearly indicates for the first time that early pre-lung events are crucial in determining the outcome of infection, and should be the focus of future studies with *N. brasiliensis*. In contrast, when STAT6- and eotaxin-1-deficient mice were infected with another intestinal nematode, *H. bakeri*, the mutant strains were as susceptible as WT mice, with parasite eggs present in similar numbers on all days examined. Resistance mechanisms that operate in the intestine during *N. brasiliensis* infections do not therefore appear to extend to a parasite that can infect naïve hosts for many months.

The FVB/N mouse strain was introduced into this study whilst exploring the potential roles of eosinophils in the intestinal phase of *N. brasiliensis* infections. The impact of intestine-specific expression of transgenes encoding IL-5 and eotaxin-1 were examined and although both lines of Tg mice were highly resistant to *N. brasiliensis*, naïve WT

FVB/N mice also showed very potent innate resistance. Very few worms were observed in the small intestine of WT FVB/N animals on day 7 p.i., whereas skin larval and leukocyte numbers (4 hours p.i.) and lung larval burdens (day 2 p.i.) were similar in WT FVB/N and WT CBA/Ca mice. Interestingly, lung larvae recovered from WT FVB/N animals were significantly smaller in size (days 1-2 p.i.) and less motile than lung larvae recovered from WT CBA/Ca mice. Further, there were significantly fewer eggs (day 6 p.i.) and worms (day 7 p.i.) in the former. However, WT FVB/N mice were no more resistant to infections with *H. bakeri* than WT CBA/Ca mice, with parasite eggs detected in comparable numbers until day 116 p.i. Resistance mechanisms operating against *N. brasiliensis* in WT FVB/N mice would not therefore appear to extend to *H. bakeri* infections.

Eosinophils can provide potent protection in some helminth infections and this study builds on our previous work and that of other groups. We have now shown that early pre-lung events may be critical in determining host resistance against *N. brasiliensis*. In secondary *N. brasiliensis* infections, cytokine signalling pathways that protect against adult worms in the late intestinal phase of infection also play a role in resistance during the early pre-lung phase, when the parasite is still at the larval stage. Eosinophils are also of importance for protection against this parasite, and future studies should focus on the early events to further characterize resistance mechanisms. This information may prove useful for the development of successful vaccines against hookworms and other nematodes.

DECLARATION

This work contains no material which has been accepted for the award of any other degree or diploma in any university or other tertiary institution and, to the best of my knowledge and belief, contains no material previously published or written by another person, except where due reference has been made in the text.

I give consent to this copy of my thesis when deposited in the University Library, being made available for loan and photocopying, subject to the provisions of the Copyright Act 1968.

The author acknowledges that copyright of published works contained within this thesis (as listed below) reside within the copyright holder/s of those works.

I also give permission for the digital version of my thesis to be made available on the web, via the University's digital research repository, the Library catalogue, the Australasian Digital Theses Program (ADTP) and also through web search engines, unless permission has been granted by the University to restrict access for a period of time.

Michelle Louise Knott

1. **Knott ML**, Matthaei KI, Giacomini PR, Wang H, Foster PS, Dent LA (2007). Impaired resistance in early secondary *Nippostrongylus brasiliensis* infections in mice with defective eosinophilopoiesis. *International Journal for Parasitology* **37** (12): 1367-78
2. **Knott ML**, Matthaei KI, Foster PS, Dent LA (2009). The roles of eotaxin and the STAT6 signalling pathway in eosinophil recruitment and host resistance to the nematodes *Nippostrongylus brasiliensis* and *Heligmosomoides bakeri*. *Molecular Immunology*. **46** (13): 2717-22
3. **Knott ML**, Hogan SP, Wang H, Matthaei KI, Dent LA (2009). FVB/N mice are highly resistant to primary infection with *Nippostrongylus brasiliensis*. *Parasitology* **136** (1): 93-106

ACKNOWLEDGEMENT OF ANY HELP

I acknowledge the help of:

All co-authors named on each of the published journal articles comprised in this thesis, for evaluating manuscript drafts and suggesting changes during the revision process. In particular, Dr Lindsay Dent, who acted as co-author for all manuscripts, co-wrote and revised drafts with myself before submission and critically read sections of my thesis.

Dr Paul Giacomini, for technical assistance with large-scale animal experiments.

STATEMENTS OF AUTHORSHIP-CHAPTER 2

NOTE:

Statements of authorship appear in the print copy of the thesis held in the University of Adelaide Library.

STATEMENTS OF AUTHORSHIP-CHAPTER 3

NOTE:

Statements of authorship appear in the print copy of the thesis held in the University of Adelaide Library.

STATEMENTS OF AUTHORSHIP-CHAPTER 4

NOTE:

Statements of authorship appear in the print copy of the thesis held in the University of Adelaide Library.

ACKNOWLEDGEMENTS

Firstly, I would like to thank my principal supervisor Dr Lindsay Dent for his guidance, support and encouragement during my Ph.D. I thoroughly enjoyed my time in your laboratory and am very grateful for the rewarding and enjoyable experience. I would also like to thank my co-supervisor Professor Klaus Matthaei for assistance with ideas and editing of manuscripts, as well as supplying many of the mice used in my Ph.D. studies. Without them much of my work would not have been possible.

To all the people who were Honours and Ph.D. students in the Dent laboratory, I thank you for your friendship and help over the years. In particular, I thank Paul Giacomini and Damon Tumes who were there with me from the start and who provided support and friendship on a daily basis and made the lab. an enjoyable place to work. I also thank Hui Wang who provided me with technical assistance when I first started in the lab.

I acknowledge the help of Wayne Damcevski at the John Curtin School of Medical Research in Canberra who bred and co-ordinated many of the mice shipments I received. I also thank the animal house staff at this institute who took over this role during the last part of my Ph.D. Without these mice, I would not have been able to complete my project.

I thank the staff and students of the discipline of Microbiology and Immunology and the School of Molecular and Biomedical Science for making it a great place in which to work. In particular, thanks must go to Wendy Parker and Francesca Bell for their friendship and who were always there to listen when I needed to talk. Special thanks to Georget Reaiche for your wonderful friendship and for always lending an ear. I have made a friend for life.

During my Ph.D. I received a Faculty of Science Divisional Scholarship. I also thank the School of Molecular and Biomedical Science for financial support to attend conferences and for other resources relating to my project.

Thanks must go to all my friends outside of uni for always being understanding and for your many years of friendship and support.

I am eternally grateful to my parents who have always supported me in everything I've done and have allowed me to get to where I am today. Thank you Mum and Dad for your love, friendship and for believing in me. To my extended family, thank you also for your love and support and to my parents- and brother-in-law.

Lastly, I thank my husband Stuart for always being there for me in all aspects of our lives (including helping me get through my Ph.D!) and for loving, supporting and caring for me always. I look forward to spending the rest of our lives together and to the adventures that we will have.

PUBLICATIONS

Within thesis:

1. **Knott ML**, Matthaei KI, Giacomini PR, Wang H, Foster PS, Dent LA (2007). Impaired resistance in early secondary *Nippostrongylus brasiliensis* infections in mice with defective eosinophilopoiesis. *International Journal for Parasitology* **37** (12): 1367-78
2. **Knott ML**, Matthaei KI, Foster PS, Dent LA (2009). The roles of eotaxin and the STAT6 signalling pathway in eosinophil recruitment and host resistance to the nematodes *Nippostrongylus brasiliensis* and *Heligmosomoides bakeri*. *Molecular Immunology* **46** (13): 2717-22
3. **Knott ML**, Hogan SP, Wang H, Matthaei KI, Dent LA (2009). FVB/N mice are highly resistant to primary infection with *Nippostrongylus brasiliensis*. *Parasitology* **136** (1): 93-106

Published abstracts:

1. **Knott ML**, Matthaei KI and Dent LA (2005). FVB/N mice are highly resistant to *Nippostrongylus brasiliensis*. *Tissue Antigens* **66** (5): 343-604

COMMONLY-USED ABBREVIATIONS

Abbreviation	full definition
AAM	alternatively activated macrophage
CAM	classically activated macrophage
CCR3	chemokine receptor 3
ECP	eosinophil cationic protein
EDN	eosinophil-derived neurotoxin
EPO	eosinophil peroxidase
GM-CSF	granulocyte-macrophage colony-stimulating factor
ICAM	intercellular adhesion molecule
Ig	immunoglobulin
IL	interleukin
IL-4Rα	interleukin-4 receptor α
Jak	janus kinase
KO	knock out
L3	third-stage larvae
L4	fourth-stage larvae
LT	leukotriene
mAb	monoclonal antibody
MBP	major basic protein
MCP	monocyte-chemotactic protein
MIP	macrophage inflammatory protein
mMCP-1	mouse mast cell protease-1
p.i.	post-infection
PAF	platelet activating factor
RANTES	regulated on activation, normal T cell expressed and secreted
s.c.	subcutaneously
STAT6	signal transducing and activator of transcription 6
Tg	transgenic
Th2	T-helper 2
VCAM	vascular cell adhesion molecule
WT	wildtype