

09 MS.M
T 366



**TARGETING PKC AND NF κ B BY
POLYUNSATURATED FATTY ACID MIMETICS
IN DIABETIC RETINOPATHY**

ELAINE BING-AI THAM

M.B.,B.S., DCH

Thesis submitted for the degree of Master of Medical Science (M Med Sc)

Department of Immunopathology

Women's and Children's Hospital

Faculty of Health Sciences

Department of Paediatrics

The University of Adelaide

May 2004

SUMMARY

Hyperglycaemia-induced vascular complications of diabetes mellitus continues to cause significant morbidity despite recent advances in therapy. Protein kinase C (PKC) and nuclear factor κ B (NF κ B) are two key signaling molecules which contribute to the development of diabetic complications, nephropathy, retinopathy and cardiovascular disease. While the omega-3 fatty acids have some protective value in diabetes their use has remained limited. To improve this type of application, our department has undertaken studies to identify the relationship between specific structural elements of polyunsaturated fatty acids and their biological activities. The findings led to the synthesis of a group of novel polyunsaturated fatty acids (PUFAs), one of which (β -oxa 21:3n-3/MP5) inhibits PKC β activation and another (β -oxa 23:4n-6/MP3) which inhibits NF κ B activation. It was therefore of interest to determine the relevance of this technology to the treatment of retinopathy.

This research characterizes the effect of hyperglycaemic conditions on PKC and NF κ B activation in bovine retinal endothelial cells (BREC) in culture, and assesses the ability of the novel PUFAs to inhibit PKC and NF κ B activation.

The research first established conditions for optimal isolation procedures for preparation of highly pure BREC and culture conditions under which the cells retained the BREC characteristics. The cell preparations were Von Willebrand Factor positive and devoid of pericyte contamination.

A major finding was that BREC expressed PKC α , β I, δ and ϵ but not β II. This contrasts with previous findings which have reported the activation of the β II isozyme in BREC. Since we were unable to see the expression of PKC β II, the results suggest that previous work had been conducted with BREC contaminated with pericytes.

The data demonstrated that BREC exposed to hyperglycaemic conditions (25mM glucose) showed preferential activation of PKC β I and δ . Hyperglycaemia-induced PKC activation is therefore not generalized to all isozymes, with PKC β I being the most significant.

The results showed that while high glucose alone failed to activate NF κ B (measured as degradation of I κ B α), it caused a more persistent activation of NF κ B in response to tumour necrosis factor α (TNF) suggesting that the pathogenic effects of TNF are amplified by hyperglycaemic conditions. The results are consistent with the recent evidence that TNF is a cytokine involved in the pathogenesis of diabetes.

The findings from the research showed that β -oxa 21:3n-3 preferentially inhibited the activation of PKC β I in BREC cultured under hyperglycaemic conditions. Treatment of BREC with β -oxa 23:4n-6 significantly inhibited the activation of NF κ B induced by TNF under hyperglycaemic conditions.

This research not only contributes to a better understanding of diabetic retinopathy but also demonstrates novel ways of targeting these signaling molecules (PKC and NF κ B) with the PUFA mimetics, MP3 and MP5.

TABLE OF CONTENTS

	Page number
Summary	ii
Declaration	iv
Acknowledgements	v
Table of Contents	vi
Abbreviations	x
Index of Figures	xii
Index of Tables	xiv
Chapter One: Introduction	1
1.1 General Introduction	2
1.2 Type 1 diabetes	4
1.3 Type 2 diabetes	9
1.4 Diabetes associated complications	10
<i>1.4.1 Cardiovascular disease</i>	11
<i>1.4.2 Nephropathy</i>	12
<i>1.4.3 Neuropathy</i>	13
<i>1.4.4 Retinopathy</i>	14
1.5 New approaches to treat diabetes associated complications	15
<i>1.5.1 Protein kinase C</i>	15
<i>1.5.2 Nuclear factor kappa B</i>	17

1.5.3	<i>Targeting PKC and NFκB in diabetes</i>	23
1.6	Omega-3 polyunsaturated fatty acids	25
1.7	Structure and synthesis of fatty acids	26
1.8	Transport of fatty acids	27
1.9	Metabolism of fatty acids	28
1.10	Polyunsaturated fatty acid mimetics	34
1.11	Significance	37
1.12	Hypotheses	37
1.13	Aims	37
 Chapter Two: Materials and Methods		38
2.1	Materials	39
2.1.1	<i>General biochemicals</i>	39
2.1.2	<i>Serum, albumin, culture media and buffers</i>	40
2.1.3	<i>Protease inhibitors</i>	40
2.1.4	<i>Antibodies and conjugates</i>	41
2.1.5	<i>Materials</i>	41
2.2	Preparation of plasma	41
2.3	Preparation of culture media for BREC	42
2.4	Preparation of endothelial cells	43
2.4.1	<i>Primary cell culture of BREC</i>	43
2.4.2	<i>Primary cell culture of HUVEC</i>	44
2.4.3	<i>Determination of BREC and HUVEC purity</i>	44
2.4.4	<i>Trypsinisation of cells</i>	45

2.4.5	<i>Cryopreservation and thawing of cells</i>	45
2.5	Culture of T lymphocytes	45
2.6	Culture of HL60 cells	46
2.7	PKC isozyme expression in different cells types	47
2.8	PKC isozyme translocation in BREC	48
2.9	I κ B α degradation	49
2.10	Lowry's Protein determination	49
2.11	Western Blotting	50
2.12	Western Blot recycling	52
2.13	Synthesis of Engineered Polyunsaturated Fatty Acids	52
2.14	Presentation of fatty acids to cells	54
2.15	Statistics	54
Chapter Three: PKC expression in BREC		55
3.1	Introduction	56
3.2	Isolation of BREC	57
3.3	PKC isozyme expression in endothelial cells	57
3.4	PKC isozyme expression in human T lymphocytes	60
3.5	PKC isozyme expression in myeloid HL60 cells	60
3.6	Summary	65
Chapter Four: Inhibition of PKC activation in BREC by β-oxa 21:3n-3		66
4.1	Introduction	67
4.2	Activation of PKC by PMA	67

4.3	Activation of PKC by hyperglycaemic conditions	68
4.4	Inhibition of PKC β I by β -oxa 21:3n-3 (MP5)	73
4.5	Summary	79
Chapter Five: Inhibition of NFκB activation in BREC by β-oxa 23:4n-6		80
5.1	Introduction	81
5.2	Activation of NF κ B by hyperglycaemic conditions	82
5.3	Activation of NF κ B by TNF in the presence of high ambient glucose levels	82
5.4	Inhibition of NF κ B activation by β -oxa 23:4n-6 (MP3)	88
5.5	Summary	92
Chapter Six: Discussion		93
6.1	PKC activation in BREC by exposure to hyperglycaemic conditions	94
6.2	NF κ B activation in BREC by exposure to hyperglycaemic conditions	97
6.3	Inhibition of PKC and NF κ B activation by β -oxa polyunsaturated fatty acids	99
6.4	Concluding remarks	106
6.5	Future research	107
Chapter Seven: Bibliography		109