

**THE NEUROIMMUNOPHARMACOLOGY
OF ALCOHOL**

Yue Wu (MSc)

Discipline of Pharmacology

School of Medical Sciences, Faculty of Health Sciences

University of Adelaide

August, 2011

A thesis submitted for the degree of Doctor of Philosophy

Table of Contents

Abstract.....	vi
Declaration.....	ix
Acknowledgements.....	xi
Statement of Authorship.....	xiii
Abbreviations.....	xxii
Human gene symbols.....	xxv
Chapter I Introduction.....	1
1.1. Behavioural effects of alcohol.....	3
1.1.1. Methods to investigate the behavioural effects of alcohol.....	3
1.1.2. Sedation (duration of LORR, sleep time test).....	4
1.1.3. Motor performance (rotarod test).....	5
1.2. Sites of effects of alcohol in the brain.....	9
1.3. Neuronal mechanisms of alcohol's effects.....	11
1.3.1. Global reward pathway: neuronal circuitry.....	11
1.3.2. Acute effects of alcohol on neuronal systems.....	13
1.3.3. Chronic effects of alcohol on neuronal systems.....	15

1.4. Alcohol exposure and glial activation.....	16
1.4.1. Glia - the immunocompetent cells in the CNS.....	16
1.4.2. Alcohol and glial activation.....	19
1.4.2.1. Alcohol and TLR4 signalling.....	20
1.4.2.2. Behavioural effects of alcohol and central immune activation...26	
1.4.3. Glial-neuronal communications.....	32
1.4.4. Potential role of immune signalling inhibitors in alcohol abuse-related diseases.....	35
1.4.4.1. Glial attenuators.....	36
1.4.4.2. TLR4 signalling inhibitors.....	37
1.4.4.3. Other anti-inflammatory therapies.....	38
1.5. Alcohol-opioid interactions.....	39
1.5.1 Animal studies.....	40
1.5.2. Neuronal mechanisms of alcohol-opioid interactions.....	41
1.5.2.1 Neuronal mechanisms of respiratory depression.....	41
1.5.2.2 Other neuronal mechanisms.....	41
1.5.3. Opioids and glial TLRs activation.....	43

1.5.4. Potential role of glial TLRs in alcohol-opioid drug interactions.....	45
1.6. Alcohol pharmacokinetics.....	46
1.6.1. ADH and ALDH.....	47
1.6.2. CYPs.....	47
1.6.3. Catalase.....	48
1.7. Alcohol and opioid pharmacogenetics.....	49
1.7.1. Opioid pharmacogenetics.....	49
1.7.2. Alcohol pharmacogenetics.....	50
1.7.3. TLR4 polymorphisms.....	52
1.8. Aims and hypotheses.....	61
Chapter II Microglial and IL-1 receptor signalling mediates acute behavioural effects of alcohol.....	66
Attenuation of microglial and IL-1 signaling protects mice from acute alcohol-induced sedation and/or motor impairment.....	68
Chapter III TLR4-MyD88 signalling mediates acute behavioural effects of alcohol.....	78
Inhibiting the TLR4-MyD88 signalling cascade by genetic or pharmacologic strategies reduces acute alcohol dose-induced sedation and motor impairment in mice.....	80

Chapter IV TLR2 and MyD88 mediate alcohol's action and alcohol-morphine interaction.....	121
TLR2 and MyD88 mediate the sedative effect of alcohol and interaction between alcohol and morphine in mice.....	123
Chapter V Investigation of the association between TLR4 genetic polymorphisms and opioid or alcohol dependence.....	153
Chapter VI Conclusions.....	159
6.1. New mechanisms for the acute effects of alcohol.....	160
6.1.1. The role of Microglia in the acute effects of alcohol.....	160
6.1.2. TLR2- and TLR4-MyD88 mediate the acute effects of alcohol.....	161
6.1.3. IL-1 receptor signalling mediates the acute effects of alcohol.....	162
6.1.4. The involvement of I κ B α in the acute effects of alcohol.....	162
6.1.5 Future directions for studies of the acute effects of alcohol.....	163
6.2. New mechanisms for the interaction between alcohol and morphine.....	166
6.2.1. TLR2-MyD88 cascade mediates the alcohol-morphine interaction.....	167
6.2.2 Future directions for studies of alcohol-morphine interactions.....	169
6.3. Alcohol and opioid pharmacogenetics.....	169
6.4. Summary.....	170

Chapter VII Bibliography.....171

Abstract

Background and purpose

Alcohol exposure induces glial toll-like receptor 4 (TLR4) signalling, while morphine administration leads to both TLR2 and TLR4 signalling in the central nervous system. However, the acute behavioural consequences of such immune activation remain unknown. This thesis aimed to examine: (a) the role of microglia, TLR2, TLR4, MyD88, and IL-1 receptor signalling in sedation and motor impairment following acute alcohol administration in mice; (b) the relationship between these observed behavioural effects and the changes in central and peripheral alcohol pharmacokinetic profiles; (c) the effect of alcohol on MAPK (ERK, JNK, and p38) and NF κ B (I κ B α) pathways and the alteration of such effects by attenuating microglial, TLR4, MyD88, and IL-1 receptor signalling *ex vivo* and *in vitro*; (d) the role of TLR2, TLR4, MyD88, IL-1 receptor, and μ opioid receptor (MOR) in the interaction between alcohol and morphine as assessed by sedation in mice; and (e) the association between the *TLR4* Asp299Gly SNP and opioid or alcohol dependence in humans.

Experimental approach

In mouse studies and mouse cellular studies, pharmacological blockade of microglial signalling, TLR4, IL-1 receptor, or both MOR and TLR4 by minocycline, (+)-naloxone (the MOR-inactive isomer), IL-1 receptor antagonist, or (-)-naloxone (the MOR-active isomer), respectively, was utilised. Mice deficient in TLR2, TLR4, both TLR2 and TLR4, or MyD88 were used. The sedative effect of alcohol and the interaction between alcohol and morphine were assessed by the sleep time (loss of righting reflex) test, and alcohol dose-induced motor impairment was

determined by the rotarod test. The activation of MAPK cascade was determined by ERK, JNK, and p38 phosphorylation using a cytometric bead array assay, and the alteration in NF κ B cascades was characterised via cellular I κ B α protein levels utilising western blotting experiments.

In the human pharmacogenetic study, *TLR4* Asp299Gly SNP genotypes were determined by a polymerase chain reaction (PCR)-restriction fragment length polymerase (RFLP) assay in 99 opioid dependent subjects, 100 alcohol dependent subjects, and 56 non-dependent healthy controls.

Key results

Pharmacological or genetic inhibition of microglial activation, TLR2, TLR4, both TLR2 and TLR4, MyD88, or IL-1 receptor signalling attenuated alcohol dose-induced sedation and motor impairment in mice. The modification of I κ B α protein levels by alcohol exposure *in vitro* was time-dependent, and the increase in such protein levels was attenuated by inhibiting proinflammatory microglial activation, TLR4, MyD88, or IL-1 receptor signalling. In contrast, blocking the activities of TLR2, both TLR2 and TLR4, and MyD88, but not TLR4 or IL-1 receptor, inhibited the enhancement of alcohol's sedative effect by morphine. The human genetic data showed a lack of association between alcohol or opioid dependence and *TLR4* Asp299Gly polymorphisms.

Conclusions and implications

Collectively, these data suggest that in mice, alcohol activates microglial and TLR2- and TLR4-MyD88-NF κ B-IL-1 receptor signalling rapidly, and this activation subsequently contributes to sedation and motor impairment induced by alcohol administration. However, TLR2-MyD88, but

not TLR4 and IL-1 receptor, cascade is involved in the interaction between alcohol and morphine. Such behavioural preclinical data provide novel insights into the immune mechanisms of the effects of alcohol and opioids.

Declaration

I, Yue Wu certify that this work contains no material which has been accepted for the award of any other degree or diploma in any university or other tertiary and, to the best of my knowledge and belief, contains no material previously published or written by another person, except where due reference has been made in the text.

I give consent to this copy of my thesis when deposited in the University Library, being made available for loan and photocopying, subject to the provisions of the Copyright Act 1968.

The author acknowledges that copyright of published works contained within this thesis (as listed below) resides with the copyright holders of those works.

Wu Y, Lousberg EL, Moldenhauer LM, Hayball JD, Robertson SA, Collier JK, Watkins LR, Somogyi AA, Hutchinson MR. (2011). Attenuation of microglial and IL-1 signaling protects mice from acute alcohol-induced sedation and/or motor impairment. *Brain Behav Immun* 25: S155-164.

Wu Y, Lousberg EL, Moldenhauer LM, Hayball JD, Collier JK, Rice KC, Watkins LR, Somogyi AA, Hutchinson MR. (2011). Inhibiting the TLR4-MyD88 signalling cascade by genetic or pharmacologic strategies reduces acute alcohol dose-induced sedation and motor impairment in mice. *Br J Pharmacol*. (accepted on 9-June-2011)

I also give permission for the digital version of my thesis to be made available on the web, via the University's digital research repository, the Library catalogue and also through web search engines, unless permission has been granted by the University to restrict access for a period of time.

Yue Wu

Date

Acknowledgements

I would like to express my sincere gratitude to my supervisors, Prof. Andrew Somogyi, Dr. Janet Coller, and Dr. Mark Hutchinson, for leading me into medical research areas. Their extensive knowledge and logical way of thinking have been of great value to me. Their understanding and encouragement have provided a great basis for the present thesis.

The research presented in this thesis would have been impossible without the financial support from a China Scholarship Council (CSC)-University of Adelaide Joint Postgraduate Scholarship and a National Health and Medical Research Council (NHMRC) of Australia Grant-funded Supplementary Scholarship. I would also like to acknowledge the funding from a NHMRC project grant, an Australian Research Council (ARC) project grant, the Faculty of Health Sciences University of Adelaide, and the National Institute of Health (NIH, the United States of America) Intramural Research Programs of National Institute on Drug Abuse (NIDA) and National Institute on Alcohol Abuse and Alcoholism (NIAAA). I sincerely thank the various authorities for their travel grants to me, including NIDA-sponsored travel award in 2010, Postgraduate Travelling Fellowships for Health Science Research funded by University of Adelaide in 2010, and the Australasian Society of Clinical and Experimental Pharmacologists and Toxicologists (ASCEPT) student travel awards in 2009 and 2010, which supported me to attend and present my results at national and international conferences.

In particular, I acknowledge the help from the following people: Ms. Erin Lousberg and Dr. John Hayball for their assistance in western blotting; Dr. Lachlan Moldenhauer and Prof. Sarah Robertson for their help in flow cytometry; Dr. Simon Phipps and Prof. Paul Foster for kindly providing the genetic deficient mice; Dr. Kenner Rice for providing (+)-naloxone; Prof. Linda

Watkins for her support of this project; Dr. Daniel Barratt, Dr. Peter Athanasos, Dr. Andrea Gordon, Dr. Justin Hay, Dr. Sophie La Vincente, Dr. Erin Morton, Mr. Mario Nguyen, Mr. Aaron Farquharson, Ms. Eloise Gelston, and Mr. Dan Magan for conducting the original clinical studies from which my pharmacogenetic study drew its participants, and all the staff and students in the Discipline of Pharmacology, University of Adelaide.

I owe my most heartfelt gratitude to Ms. Sandy McConachy and Ms. Iris Liu for their care and encouragement since I arrived in Australia. I'm also very grateful to Ms. Jocelyn Ho and Ms. Erin Lousberg who helped me improve my written English. Thanks to my general practitioners and specialists, especially Prof. Justin Beilby, who helped restore my health.

To my colleagues in N529a, thanks for making it such a warm and cozy office. To all my housemates in Adelaide, thank you for being my brothers and sisters in Australia.

To my parents, other family members, and my friends in Beijing, thanks for your support and encouragement in the past three years. I especially want to thank A.Prof. Xianglin Zhang and those friends who helped me take care of my mother while I was not there.

Statement of Authorship

Attenuation of microglial and IL-1 signaling protects mice from acute alcohol-induced sedation and/or motor impairment

Brain Behav Immun (2011) 25: S155-S164.

Impact Factor: 5.061

Wu Y (Candidate)

Contribute to the experimental design, conducted all experimental procedures, statistical analysis, graphical presentation of the data collected, and prepared the manuscript for submission

I hereby certify that the statement of contribution is accurate.

Signed

Date

29/07/2011

Lousberg EL

Assisted with western blots and manuscript evaluation

I hereby certify that the statement of contribution is accurate and I give permission for the inclusion of the paper in the thesis.

Signed

Date

18/07/2011

Moldenhauer LM

Assisted with flow cytometry and manuscript evaluation

I hereby certify that the statement of contribution is accurate and I give permission for the inclusion of the paper in the thesis.

Signed

Date

19/7/11

Hayball JD

Helped in western blots and manuscript evaluation

I hereby certify that the statement of contribution is accurate and I give permission for the inclusion of the paper in the thesis.

Signed *Date* 18/07/2011

Robertson SA

Helped in flow cytometry and manuscript evaluation

I hereby certify that the statement of contribution is accurate and I give permission for the inclusion of the paper in the thesis.

Signed *Date* 20/07/2011

Coller JK

Contributed to experimental design and manuscript evaluation

I hereby certify that the statement of contribution is accurate and I give permission for the inclusion of the paper in the thesis.

Signed *Date* 29/07/2011

Watkins LR

Provided critical evaluation of article

I hereby certify that the statement of contribution is accurate and I give permission for the inclusion of the paper in the thesis.

Signed *Date* 18/07/2011

Somogyi AA

Contributed to the experimental design, helped in data interpretation and preparation and critical evaluation of the manuscript

I hereby certify that the statement of contribution is accurate and I give permission for the inclusion of the paper in the thesis.

Signed

Date 01/08/2011

Hutchinson MR

Contributed to the experimental design, supervised the data interpretation and preparation of the manuscript, and acted as corresponding author

I hereby certify that the statement of contribution is accurate and I give permission for the inclusion of the paper in the thesis.

Signed

Date 29/07/2011

Inhibiting the TLR4-MyD88 signalling cascade by genetic or pharmacologic strategies reduces acute alcohol dose-induced sedation and motor impairment in mice

Br J Pharmacol (2011): accepted paper

Impact Factor: 4.925

Wu Y (Candidate)

Contribute to the experimental design, conducted all experimental procedures, statistical analysis, graphical presentation of the data collected, and prepared the manuscript for submission

I hereby certify that the statement of contribution is accurate.

Signed _____ *Date* 29/07/2011

Lousberg EL

Assisted with western blots and manuscript evaluation

I hereby certify that the statement of contribution is accurate and I give permission for the inclusion of the paper in the thesis.

Signed _____ *Date* 18/07/2011

Moldenhauer LM

Assisted with flow cytometry and manuscript evaluation

I hereby certify that the statement of contribution is accurate and I give permission for the inclusion of the paper in the thesis.

Signed _____ *Date* 19/7/11

Hayball JD

Helped in western blots and manuscript evaluation

I hereby certify that the statement of contribution is accurate and I give permission for the inclusion of the paper in the thesis.

Signed

Date 18/07/2011

Coller JK

Contributed to experimental design and manuscript evaluation

I hereby certify that the statement of contribution is accurate and I give permission for the inclusion of the paper in the thesis.

Signed

Date 29/07/2011

Rice KC

Contributed to manuscript evaluation

I hereby certify that the statement of contribution is accurate and I give permission for the inclusion of the paper in the thesis.

Signed

Date

Watkins LR

Provided critical evaluation of article

I hereby certify that the statement of contribution is accurate and I give permission for the inclusion of the paper in the thesis.

Signed

Date 18/07/2011

Somogyi AA

Contributed to the experimental design, helped in data interpretation, and preparation and critical evaluation of the manuscript

I hereby certify that the statement of contribution is accurate and I give permission for the inclusion of the paper in the thesis.

Signed

Date 01/08/2011

Hutchinson MR

Contributed to the experimental design, supervised the data interpretation and preparation of the manuscript, and acted as corresponding author

I hereby certify that the statement of contribution is accurate and I give permission for the inclusion of the paper in the thesis.

Signed

Date 29/07/2011

TLR2 and MyD88 mediate the sedative effect of alcohol and interaction between alcohol and morphine in mice

Br J Pharmacol (2011): submitted paper

Impact Factor: 4.925

Wu Y (Candidate)

Contribute to the experimental design, conducted all experimental procedures, statistical analysis, graphical presentation of the data collected, and prepared the manuscript for submission

I hereby certify that the statement of contribution is accurate.

Signed *Date* 29/07/2011

Coller JK

Contributed to experimental design and provided critical evaluation of article

I hereby certify that the statement of contribution is accurate and I give permission for the inclusion of the paper in the thesis.

Signed *Date* 29/07/2011

Rice KC

Contributed to manuscript evaluation

I hereby certify that the statement of contribution is accurate and I give permission for the inclusion of the paper in the thesis.

Signed *Date*

Diener KR

Provided null mutant mice and helped in manuscript evaluation

I hereby certify that the statement of contribution is accurate and I give permission for the inclusion of the paper in the thesis.

Signed

Date 18/07/2011

Hayball JD

Provided null mutant mice and helped in manuscript evaluation

I hereby certify that the statement of contribution is accurate and I give permission for the inclusion of the paper in the thesis.

Signed

Date 18/07/2011

Watkins LR

Helped in manuscript evaluation

I hereby certify that the statement of contribution is accurate and I give permission for the inclusion of the paper in the thesis.

Signed

Date 18/07/2011

Somogyi AA

Contributed to the experimental design, helped in data interpretation, and preparation and critical evaluation of the manuscript

I hereby certify that the statement of contribution is accurate and I give permission for the inclusion of the paper in the thesis.

Signed

Date 01/08/2011

Hutchinson MR

Contributed to the experimental design, supervised the data interpretation and preparation of the manuscript, and acted as corresponding author

I hereby certify that the statement of contribution is accurate and I give permission for the inclusion of the paper in the thesis.

Signed

Date 29/07/2011

Abbreviations

ADH	alcohol dehydrogenase
ALDH	aldehyde dehydrogenase
ATP	adenosine triphosphate
B2M	β -2-microglobulin
BBB	blood-brain barrier
cAMP	cyclic adenosine monophosphate
CCL	chemokine (C-C) motif ligand
CCR	chemokine (C-C) motif ligand receptor
CD	cluster of differentiation
CI	confidence interval
CNS	central nervous system
COPD	chronic obstructive pulmonary disease
COX	cyclooxygenase
CTSF	cathepsin F
CTSS	cathepsin S
CXCL	chemokine (C-X-C) motif ligand
CYP	cytochrome P450
DA	dopamine
ERK	extracellular regulated kinase
GABA	gamma-aminobutyric acid
GFAP	glial fibrillary acidic protein
HBV	hepatitis B virus
IFN	interferon
IL	interleukin
IL-1RI	interleukin-1 β receptor type I
IL-1ra	IL-1 receptor antagonist
iNOS	inducible nitric oxide synthase

IRAK4	interleukin-1 receptor-associated kinase 4
IRF3	IFN regulatory factor 3
I κ B α	NF κ B inhibitor α
JNK	c-Jun N-terminal kinase
LBP	LPS binding protein
LORR	loss of righting reflex
LPS	lipopolysaccharide
MAL	myelin and lymphocyte protein
MAPK	mitogen-activated protein kinase
MD-2	myeloid differentiation factor 2
MHC	major histocompatibility complex
MOR	μ opioid receptor
mRNA	messenger ribonucleic acid
MyD88	myeloid differentiation primary response gene 88
NAc	nucleus accumbens
NAD	nicotinamide adenine dinucleotide
NF κ B	nuclear factor kappa-light-chain-enhancer of activated B cells
NMDA	N-methyl-D-aspartate
NO	nitric oxide
OR	odds ratio
PCR	polymerase chain reaction
PI3K	phosphoinositide 3 kinase
PKA	protein kinase A
PKC	protein kinase C
SN	substantia nigra
SNP	single nucleotide polymorphism
TIR	toll/IL-1 receptor
TLR	toll-like receptor
TNF- α	tumor necrosis factor- α

TRAM	TRIF-related adaptor molecule
TRIF	toll/IL-1R domain containing adaptor inducing interferon- β
VTA	ventral tegmental area

Human gene symbols

Gene symbol	Gene name	Protein
<i>ABCB1</i>	ATP-binding cassette, sub-family B, member 1	P-glycoprotein
<i>ADH1B</i>	alcohol dehydrogenase 1B (class I), beta polypeptide	ADH1B
<i>ADH4</i>	alcohol dehydrogenase 4 (class II), pi polypeptide	ADH4
<i>ALDH1A1</i>	aldehyde dehydrogenase 1 family, member A1	ALDH1A1
<i>ALDH2</i>	aldehyde dehydrogenase 2 family (mitochondrial)	ALDH2
<i>CYP2B6</i>	cytochrome P450, family 2, subfamily B, polypeptide 6	CYP2B6
<i>CYP2E1</i>	cytochrome P450, family 2, subfamily E, polypeptide 1	CYP2E1
<i>CYP3A4</i>	cytochrome P450, family 3, subfamily A, polypeptide 4	CYP3A4
<i>GABRA6</i>	gamma-aminobutyric acid (GABA) A receptor, alpha 6	GABA _A α6
<i>IL10</i>	interleukin 10	IL-10
<i>IL1B</i>	interleukin 1, beta	IL-1β
<i>IL1RN</i>	interleukin 1 receptor antagonist	IL-1ra
<i>LY96</i>	lymphocyte antigen 96	MD-2
<i>NFKB1</i>	nuclear factor of kappa light polypeptide gene enhancer in B-cells 1	NFκB
<i>OPRM1</i>	opioid receptor, mu 1	MOR
<i>TLR2</i>	toll-like receptor 2	TLR2
<i>TLR4</i>	toll-like receptor 4	TLR4