



THE UNIVERSITY
of ADELAIDE

**ELUCIDATION AND
ISOLATION OF SPECIFIC
BIOACTIVE COMPOUND IN
CYANOBACTERIA ISOLATES**

by

Chee Kuan Kwei

A Thesis submitted for the degree of
Doctor of Philosophy

2012

School of Chemical Engineering
The University of Adelaide
Australia

Learn from yesterday, live for today, hope for tomorrow. The important thing is not to stop questioning.

Albert Einstein

ABSTRACT

Sulfoquinovosyldiacylglyceride (SQDG), one of the bioactive compounds isolated from *Spirulina*, has been proven to act as an inhibitor of reverse-transcriptase, and has potential for the use in the Combination Anti-Retroviral Therapy (cART). A number of researchers report that the many potential uses of *Spirulina*, with the most widely used strains being *Spirulina platensis* and *S. maxima*, require further investigation to determine their actual usefulness in medical applications.

In this work, four commercial isolates marketed as *Spirulina* and an Australian isolate, *Spirulina* sp. CS-785/01 (CSIRO), were selected to determine their bio-composition and applicability as a source of bioactive compounds. During the course of the investigation, it was determined that the four commercial isolates were in fact *Arthrospira* and the Australian isolate is *Halospirulina*. However, this insight occurred towards the end of the study and the '*Spirulina*' has been adopted as common name when referring to the isolates during the experimental programme that has been reported in the thesis. The commercial isolates used in this research were *Spirulina* (J), *Spirulina* (M), *Spirulina* (P) and *Spirulina* (S), while the Australian isolate *Spirulina* sp. was also used throughout the investigation.

The identification of these isolates was examined based on their molecular and the chemotaxonomic classification. Furthermore, the genes responsible for the production of SQDG were isolated to assess the potential therapeutic value of these isolates in treating disease, such as HIV (Human immunodeficiency virus). Since SQDG is a compound of current interest, a suitable extraction technique was developed to optimise its production. Different extraction techniques, such as microwave-assisted sonication and homogenisation, coupled with various forms of organic solvents, were examined in this study. Preliminary phytochemical analysis was also undertaken to reveal the potential use of the investigated isolates for the further development of pharmaceuticals due to the presence of specific phyto-constituents. Finally, sequential extracts and the isolated compound of SQDG were used to determine their bioactivity against a range of microbes and HIV.

The results revealed that the Australian isolate was not highly productive in respect to growth. This is due to its limited adaptability to changes in temperature and culture media type. The Australian isolate is limited to a specific temperature range and culture media type. Additionally, it has substantial biochemical variation compared to the commercial isolates. The diversity of these isolates could be explained by their molecular and chemotaxonomic classification, which revealed that the Australian isolate belongs to the genus of *Halospirulina*, while the commercial isolates belong to the genus *Arthrospira*. The carbohydrate, protein, lipid and fatty acid content of the commercial isolates also indicated that they have higher nutritional value when compared to the *Spirulina* sp. This strongly suggested that the commercial isolates belonged to a different genus to the *Spirulina* sp.

In spite of the diversity of the classification, all investigated isolates showed the presence of SQDG. Overall, the results showed that *Spirulina* (S) has a high potential for synthesis of SQDG, due to its high content of C16:0 and C18:2, while the SQDG content of Australian isolate was lowest among the investigated isolates. By applying this data, the Australia isolate is unlikely to be suitable for large-scale production. It also showed lower appreciable amounts of SQDG and gamma linolenic acid (GLA). The highest yield of SQDG from *Spirulina* (M) was from using a chloroform:methanol (2:1, v/v) extraction solvent system. However, a methanol extraction solvent system is suggested, due to its high recovery of SQDG and low toxicity. Because SQDG is a potent inhibitor of HIV-reverse transcriptase, it can be concluded that commercial isolates are good sources for drug production because of their high content of SQDG and rapid biomass production.

STATEMENT OF ORIGINALITY

This work contains no material that has been accepted for the award of any other degree or diploma in any university or other tertiary institution from Chee Kuan Kwei and, to the best of my knowledge and belief, contains no material previously published or written by another person, except where due reference has been made in the text.

I give consent for a copy of my thesis, when deposited in the University library, to be made available for loan and photocopying, subject to the provisions of the Copyright Act 1968.

I also give permission for a digital version of my thesis to be made available on the web, via the University's digital research repository, the library catalogue, the Australasian Digital Theses Program (ADTP), and also through web search engines, unless permission has been granted by the University to restrict access for a period of time.

Signed: Date:

Chee Kuan Kwei

ACKNOWLEDGEMENTS

I would like to take this opportunity to thank a number of people who have supported me throughout my Ph.D. research. Firstly, I would like to thank my supervisors, Associate Professor David Lewis, Professor Keith King, Dr William Donohue and Professor Brett Neilan. Cheers to Associate Professor David for giving me guidance, patience, support and encouragement for commencing the thesis. Cheers to Professor Keith for being a fantastic mentor in helping me to achieve this goal. Cheers to Dr William for taking part in my thesis and assisting me in the completion of a daunting piece of work. Cheers to Professor Brett for providing me endless support in the molecular genetics investigation.

I really appreciate the help from other outstanding academics: Dr Tuck Weng Kok (IMVS), Dr Gordon Wilkinson, A/Prof Wei Zhang (Flinders University), and Dr Eric Capelle (SARDI) for allowing me to access to their apparatuses. Thanks are also due to Margaret Stopa (SARDI), Su Peng (Flinders University) and David Apps (University of Adelaide, Waite Campus) for their willingness to share their ideas. I would like to seize this opportunity to acknowledge Dr. Michelle Yvette Picard and Miss Patricia Zoltan, who guided me during the write up the research and the consultation for my research proposal during my first six months of my Ph.D.

Acknowledgement is also given to all my colleagues and friends within the School of Chemical Engineering, and the people in the microalgae lab who have been in the lab in the past four years. Special mention must be made to Dr Troco Mihali for the help I got at the University of New South Wales, and the fun from everyone else in the BAN lab. The financial support from the School of Chemical Engineering at the University of Adelaide is also acknowledged.

Last but not the least; I am indebted to my parents and brother for being so supportive during my studies. And to Clayton, my lovely partner, who has been a constant source of encouragement throughout this long journey.

Thank You!

LIST OF PUBLICATIONS

1. C. K. Kwei, D. M. Lewis, K. D. King, W. Donohue, B. A. Neilan (2007), 'Therapeutic potential of *Spirulina* for the treatment of HIV', International Society for Pharmaceutical Engineering, Gold Coast, Australia, 2-4 September 2007 (Poster Presentation).
2. C. K. Kwei, D. M. Lewis, K. D. King, W. Donohue, B. A. Neilan (2008), 'Therapeutic potential of *Spirulina* for the treatment of HIV', International Biotechnology Symposium and Exhibition, Dalian, China, 12-17 October 2008 (Poster Presentation).
3. C. K. Kwei, D. M. Lewis, K. D. King, W. Donohue, B. A. Neilan (2010), 'The extraction of sulfoquinovosyldiacylglyceride from *Spirulina*', CHEMECA, Adelaide, Australia, 27-30 September 2010 (Oral Presentation).
4. C. K. Kwei, D. M. Lewis, K. D. King, W. Donohue, B. A. Neilan (2011), 'Molecular classification of commercial *Spirulina* strains and identification of sulfolipid biosynthesis genes', *Journal of Microbiology and Biotechnology*, 21(4), pp. 365-371.
5. C. K. Kwei, D. M. Lewis, K. D. King, W. Donohue, B. A. Neilan (n.d), 'Preliminary phytochemical analysis of *Arthrospira maxima* and *Arthrospira seawater*', *Indian Journal of Pharmaceutical Sciences* (Submitted).

TABLE OF CONTENTS

| | |
|---|------------|
| ABSTRACT | ii |
| STATEMENT OF ORIGINALITY | iv |
| ACKNOWLEDGEMENTS | v |
| LIST OF PUBLICATIONS | vi |
| TABLE OF CONTENTS | vii |
| LIST OF FIGURES | xi |
| LIST OF TABLES | xiv |
| ABBREVIATION | xvi |
| CHAPTER 1 INTRODUCTION | 1 |
| 1.1 Aims | 1 |
| 1.2 Natural products..... | 2 |
| 1.3 Current HIV treatment used..... | 6 |
| 1.4 Human immunodeficiency virus..... | 7 |
| 1.4.1 Epidemiology of HIV | 8 |
| 1.4.2 Signs and symptoms of HIV/AIDS infection | 9 |
| 1.5 Gap statement | 11 |
| 1.6 Thesis overview | 12 |
| CHAPTER 2 LITERATURE REVIEW | 15 |
| 2.1 <i>Spirulina</i> | 15 |
| 2.1.1 History of <i>Spirulina</i> | 15 |
| 2.1.2 Classification of <i>Spirulina</i> | 16 |
| 2.1.3 Taxonomy | 16 |
| 2.1.4 Chemical composition of <i>Spirulina</i> | 18 |
| 2.2 Maintain an axenic culture..... | 22 |
| 2.3 Cultivation of <i>Spirulina</i> | 22 |
| 2.3.1 Effect of the environment on <i>Spirulina</i> | 24 |
| 2.3.2 Effect of growth parameters on bioactive compound..... | 26 |
| 2.4 Application of <i>Spirulina</i> | 28 |
| 2.4.1 Use of <i>Spirulina</i> in waste-water treatment | 28 |
| 2.4.2 Use of <i>Spirulina</i> in aquaculture | 29 |
| 2.5 Therapeutic potential of <i>Spirulina</i> | 29 |
| 2.5.1 Cyanovirin-N | 30 |
| 2.5.2 Sulphated polysaccharides | 30 |
| 2.5.3 Sulfolipids..... | 31 |
| 2.6 Target natural products—Sulfoquinovosyldiacylglyceride | 31 |
| 2.6.1 Pathways for the biosynthesis of Sulfolipids..... | 32 |
| 2.7 Metabolism within <i>Spirulina</i> | 37 |
| 2.7.1 Primary and secondary metabolism..... | 37 |
| 2.7.2 Metabolic pathways of the production of metabolites encoded with NRPS and PKS..... | 38 |
| 2.8 Drying techniques | 41 |
| 2.9 Extraction..... | 42 |
| 2.9.1 Traditional solvent extraction | 44 |

| | |
|---|-----------|
| 2.9.2 Extraction techniques..... | 45 |
| 2.9.3 Summary of studies on extraction methods applied to <i>Spirulina</i> | 46 |
| 2.10 Summary | 47 |
| CHAPTER 3 MATERIALS AND METHOD..... | 49 |
| 3.1 Cyanobacterial isolates | 49 |
| 3.1.1 <i>Spirulina</i> sp..... | 50 |
| 3.1.2 <i>Spirulina</i> (P)..... | 52 |
| 3.1.3 <i>Spirulina</i> (M) | 53 |
| 3.1.4 <i>Spirulina</i> (S)..... | 53 |
| 3.1.5 <i>Spirulina</i> (J) | 53 |
| 3.2 Ultraviolet radiation | 54 |
| 3.3 Growth | 54 |
| 3.4 Cell preparation for extraction from culture medium | 55 |
| 3.5 Isolation of genomic DNA (DNA extraction) | 55 |
| 3.5.1 XS DNA extraction..... | 55 |
| 3.5.2 Improved XS DNA extraction | 56 |
| 3.6 Identification of cyanobacterial isolates | 56 |
| 3.6.1 16S rDNA, PKS and NRPS genes PCR amplification | 56 |
| 3.6.2 Identification of gene responsible for sulfolipids biosynthesis—Primer design..... | 58 |
| 3.7 Agarose gel electrophoresis | 59 |
| 3.8 Purification of DNA..... | 60 |
| 3.8.1 Purification of DNA from PCR products | 60 |
| 3.8.2 Purification of DNA from agarose gels | 60 |
| 3.9 Sequence clean up..... | 62 |
| 3.10 DNA sequence analysis | 62 |
| 3.10.1 GenBank accession numbers | 62 |
| 3.11 Ash free dry weight..... | 63 |
| 3.12 Carbohydrate extraction..... | 63 |
| 3.13 Protein extraction | 63 |
| 3.14 Sequential extraction..... | 63 |
| 3.15 Lipid extraction | 64 |
| 3.15.1 Pre-treatment for lipid extraction..... | 64 |
| 3.15.2 Lipid extraction techniques..... | 65 |
| 3.16 Chromatography | 69 |
| 3.16.1 Thin layer chromatography..... | 69 |
| 3.16.2 Gas chromatography | 71 |
| 3.16.3 High performance liquid chromatography..... | 72 |
| 3.16.4 Column chromatography | 74 |
| 3.17 Phytochemical analysis | 75 |
| 3.17.1 Test for tannins | 76 |
| 3.17.2 Test for saponins | 76 |
| 3.17.3 Test for phlobatannins | 76 |
| 3.17.4 Test for cardiac glycosides (Keller-Kiliani test)..... | 76 |
| 3.17.5 Test for steroids | 76 |
| 3.17.6 Test for terpenoids | 76 |
| 3.17.7 Test for flavonoids | 77 |
| 3.17.8 Test for alkaloids | 77 |
| 3.18 Anti-bacterial activity | 77 |
| 3.19 Statistical Analysis..... | 78 |

| | |
|---|------------|
| CHAPTER 4 MOLECULAR CLASSIFICATION, GROWTH CHARACTERISTICS AND CELL COMPOSITION OF <i>SPIRULINA</i> ISOLATES | 79 |
| 4.1 Introduction..... | 79 |
| 4.2 Screening of 16s rDNA..... | 80 |
| 4.3 Exposure under ultraviolet radiation..... | 84 |
| 4.4 Growth characteristics | 86 |
| 4.5 Growth condition of <i>Spirulina</i> sp. | 87 |
| 4.5.1 Effect of nutrient uptake | 87 |
| 4.5.2 Effect of temperature | 88 |
| 4.6 Chemical composition of <i>Spirulina</i> isolates | 88 |
| 4.7 Fatty acid profile of <i>Spirulina</i> isolates..... | 89 |
| 4.8 Discussion..... | 92 |
| 4.9 Summary..... | 98 |
| CHAPTER 5 SQDG SYNTHESIS | 100 |
| 5.1 Introduction..... | 100 |
| 5.2 Screening of <i>Spirulina</i> isolates with NRPS and PKS | 101 |
| 5.3 Screening of <i>Spirulina</i> isolates with sqdB | 102 |
| 5.4 Screening of <i>Spirulina</i> isolates with sqdX..... | 106 |
| 5.5 Biosynthesis pathway of SQDG for <i>Spirulina</i> | 108 |
| 5.6 Phylogenetic studies on sqdB and sqdX | 110 |
| 5.7 Discussion..... | 113 |
| 5.8 Summary..... | 116 |
| CHAPTER 6 EXTRACTION OF SQDG FROM <i>SPIRULINA</i> ISOLATES | 118 |
| 6.1 Introduction..... | 118 |
| 6.2 Analysis of sequential extracts | 119 |
| 6.3 The drying condition of <i>Spirulina</i> | 122 |
| 6.4 Lipid extraction..... | 125 |
| 6.4.1 Effect of cell disruption | 125 |
| 6.4.2 Effect of solvent systems | 125 |
| 6.4.3 The effect of different ratio of solvent systems (Chloroform:methanol)..... | 128 |
| 6.4.4 Soxhlet and supercritical carbon dioxide extraction..... | 129 |
| 6.5 Identification of sulfolipids..... | 132 |
| 6.6 Discussion..... | 134 |
| 6.7 Summary..... | 141 |
| CHAPTER 7 QUANTIFICATION OF SQDG | 143 |
| 7.1 Introduction..... | 143 |
| 7.2 Quantification of SQDG | 144 |
| 7.2.1 HPLC-UV detector | 146 |
| 7.2.2 HPLC-ELSD detector | 146 |
| 7.3 Purification of SQDG | 149 |
| 7.4 Preliminary antibacterial assay | 151 |
| 7.5 Anti-HIV assay | 151 |
| 7.6 Discussion..... | 153 |
| 7.7 Summary..... | 158 |
| CHAPTER 8 CONCLUSION..... | 160 |
| 8.1 Further direction | 164 |
| CHAPTER 9 REFERENCES | 165 |

| | |
|-------------------------|------------|
| Appendix A | 193 |
| Appendix B | 196 |
| Appendix C | 200 |
| Appendix D | 207 |
| Appendix E | 214 |
| Appendix F | 215 |

LIST OF FIGURES

| | |
|--|----|
| Figure 1.1: Global prevalence of HIV in 2009 | 9 |
| Figure 1.2: Targets within the different phases of HIV-1 viral replication cycle and infection of a T-cell, as used for anti-HIV cell-based assays..... | 10 |
| Figure 1.3: The expression of HIV | 11 |
| Figure 2.1: View of <i>Spirulina</i> under a microscope..... | 17 |
| Figure 2.2: Photomicrographs of cyanobacteria isolates | 18 |
| Figure 2.3: Global production figures of <i>Spirulina</i> by country based on literature, company and trade information | 23 |
| Figure 2.4: An existing site of production of Australian <i>Spirulina</i> at Darwin, Northern Territory..... | 24 |
| Figure 2.5: Structure of the sulfolipids sulfoquinovosyl diacylglycerol (SQDG)..... | 33 |
| Figure 2.6: Proposed pathway for the biosynthesis of the plant sulfolipids (sulfoquinovosyldiacylglycerol) | 34 |
| Figure 2.7: The final step of elucidation of sulfolipids biosynthesis..... | 35 |
| Figure 2.8: Biosynthesis of SL-1 | 36 |
| Figure 2.9: Main pathways of some secondary and primary metabolites biosynthesis | 38 |
| Figure 2.10: Appearance of oven dried <i>Spirulina</i> in different air temperatures..... | 41 |
| Figure 2.11: Relationship between the cohesion values with the yield of lipid based on solvent extractions of EtOH (Ethanol), Meth (Methanol), IAA (Isoamylalcohol), But (Butanol), Chlor (Chloroform), B/C (Butanol/Chloroform, 1:1, v/v), E/C (Ethanol/ Chloroform, 1:1, v/v), M/C (Methanol/Chloroform, 1:1, v/v) | 44 |
| Figure 3.1: The locations of the five different isolates of <i>Spirulina</i> | 49 |
| Figure 3.2: Location of Pearse Lake, Rottnest Island, Western Australia..... | 50 |
| Figure 3.3: Subculture of <i>Spirulina</i> sp. from tissue culture flasks (small scale) to 2L flasks (large-scale) | 52 |
| Figure 3.4: Gene ruler (ladder mix 10 kb)..... | 60 |
| Figure 3.5: Soxhlet extraction apparatus | 67 |
| Figure 3.6: Schematic of the supercritical carbon dioxide extraction unit | 68 |
| Figure 3.7: Supercritical carbon dioxide extractor used in present study..... | 69 |
| Figure 3.8: The operation of ELSD (Daniel, 2007)..... | 73 |
| Figure 3.9: Luer-lock column used for the purification of SQDG | 74 |
| Figure 3.10: Diagram of the fractionation of SQDG from the crude extract..... | 75 |
| Figure 3.11: Potato dextrose agar plate with 6 mm diameter discs for anti-fungal testing | 78 |
| Figure 4.1: PCR amplification of 16S rRNA from <i>Spirulina</i> sp. | 80 |
| Figure 4.2: PCR amplification of 16S rRNA from five <i>Spirulina</i> isolates..... | 81 |
| Figure 4.3: The sequence of 16S rRNA for: J: <i>Spirulina</i> (J), W: <i>Spirulina</i> (S), M: <i>Spirulina</i> (M)..... | 82 |

| | |
|---|-----|
| Figure 4.4: The sequence of 16S rRNA for: P: <i>Spirulina</i> (P), and S: <i>Spirulina</i> sp. | 83 |
| Figure 4.5: Duration of exposure under ultraviolet radiation | 85 |
| Figure 4.6: <i>Spirulina</i> sp. lost its green pigmentation after 120 minutes irradiation | 86 |
| Figure 4.7: Growth characteristic of <i>Spirulina</i> sp..... | 86 |
| Figure 4.8: <i>Spirulina</i> sp. culture in different media..... | 87 |
| Figure 4.9: <i>Spirulina</i> sp. culture in MLA/seawater media at different temperatures | 88 |
| Figure 4.10: Fatty acid content in different isolates of <i>Spirulina</i> | 91 |
| Figure 4.11: Gas chromatographic separation of the methyl ester derivatives of the fatty acids of the external standard | 91 |
| Figure 5.1: Results of NRPS and PKS PCR amplification..... | 101 |
| Figure 5.2: The nucleotide sequence of unspecific band for <i>Spirulina</i> sp..... | 102 |
| Figure 5.3: PCR in different condition for different isolates of <i>Spirulina</i> using sqdB. | 103 |
| Figure 5.4: Purified sqdB product from all isolates of <i>Spirulina</i> | 104 |
| Figure 5.5: The sequence for sqdB for J: <i>Spirulina</i> (J), P: <i>Spirulina</i> (P), W: <i>Spirulina</i> (S), S: <i>Spirulina</i> sp. | 105 |
| Figure 5.6: An erroneous DNA sequence amplified from <i>Synechocystis</i> PCC 6803 .. | 106 |
| Figure 5.7: The initial trial on annealing temperature using PCR program | 107 |
| Figure 5.8: The PCR condition with annealing temperature | 107 |
| Figure 5.9: The sequence for sqdX for J: <i>Spirulina</i> (J), P: <i>Spirulina</i> (P), W: <i>Spirulina</i> (S), S: <i>Spirulina</i> sp. | 108 |
| Figure 5.10: Comparison between the organisation of sqd genes in <i>R. sphaeroides</i> , <i>Synechococcus</i> and <i>Synechocystis</i> (Benning, 1998) | 109 |
| Figure 5.11: Phylogenetic tree based on sqdB homologues | 111 |
| Figure 5.12: Phylogenetic tree based on sqdX homologues..... | 112 |
| Figure 6.1: Appearance of <i>Spirulina</i> in powder form and view of <i>Spirulina</i> under a microscope | 123 |
| Figure 6.2: <i>Spirulina</i> sp. using different oven temperature for drying | 124 |
| Figure 6.3: Effect of extraction systems on different isolates of <i>Spirulina</i> | 125 |
| Figure 6.4: Effect of solvent systems on different isolates of <i>Spirulina</i> | 126 |
| Figure 6.5: Effect of different ratio of chloroform:methanol on different isolates of <i>Spirulina</i> | 129 |
| Figure 6.6: Cumulative yield of supercritical extraction of lipids from <i>Spirulina</i> (M) isolate | 130 |
| Figure 6.7: Supercritical carbon dioxide extraction conducted in two different models | 130 |
| Figure 6.8: Cumulative yield of lipids from the <i>Spirulina</i> (M) isolate as a function of carbon dioxide mass | 131 |
| Figure 6.9: TLC for the identification of sulfolipid from different strains of algae by using the Bligh and Dyer lipid extraction method | 132 |
| Figure 7.1: Sulfolipids recovered based on different solvent systems used | 144 |
| Figure 7.2: Sulfolipids recovered based on different solvent systems used | 145 |
| Figure 7.3: HPLC Chromatogram using UV/VIS detector at wavelength 208nm | 146 |

| | |
|--|-----|
| Figure 7.4: HPLC/ELSD chromatogram for the chloroform: ethanol (2:1, v/v) extract of <i>Spirulina</i> (M) | 147 |
| Figure 7.5: Sequential extracts in developing system..... | 150 |
| Figure 7.6: Serial dilution of the SQDG was observed under microscope for cytotoxicity effect | 152 |

LIST OF TABLES

| | |
|--|-----|
| Table 1.1 Anti-HIV products from algae and cyanobacteria..... | 5 |
| Table 1.2: Global summary of the AIDS epidemic December 2010..... | 8 |
| Table 2.1: Chemical composition of different strains of <i>Spirulina</i> | 19 |
| Table 2.2: Fatty acid composition of different strains and species of <i>Spirulina</i> | 21 |
| Table 2.3: Cyanobacteria strains analysed by (NRPS and PKS) polymerase chain reaction (PCR) and bioassay results | 39 |
| Table 2.4: Review of advantages and disadvantages of extraction methods..... | 43 |
| Table 2.5: Extraction methods for <i>Spirulina</i> | 47 |
| Table 3.1: Details of algae investigated in this study | 50 |
| Table 3.2: Various growth conditions for <i>Spirulina</i> sp. | 52 |
| Table 3.3: Cyanobacterial 16S rRNA gene amplification and sequencing primers | 57 |
| Table 3.4: Reaction mixture recipe | 58 |
| Table 3.5: Designed degenerate primers for sqdB and sqdX genes | 58 |
| Table 3.6: PCR cycles..... | 59 |
| Table 3.7: Reaction mixture for sequencing PCR | 61 |
| Table 3.8: Dielectric constants of solvents used for the sequential extraction..... | 64 |
| Table 3.9: Combination system of the organic solvent used for extraction | 66 |
| Table 3.10: TLC developing systems for SQDG..... | 70 |
| Table 3.11: Gradient elution | 73 |
| Table 3.12: Bacteria and fungi used in the anti-microbial testing | 77 |
| Table 4.1: Similarity to the closest relatives in GenBank of 16S rDNA | 84 |
| Table 4.2: The classification of different isolates of <i>Spirulina</i> | 84 |
| Table 4.3: Growth of bacteria and appearance of <i>Spirulina</i> sp. after ultraviolet exposure..... | 85 |
| Table 4.4: Chemical composition of different isolates of <i>Spirulina</i> | 89 |
| Table 4.5: Fatty acid profile for <i>Spirulina</i> isolates | 90 |
| Table 5.1: Sequence analysis of the sqd genes amplified from the isolates of <i>Spirulina</i> | 110 |
| Table 6.1: The estimation of yield of different sequential extracts from the <i>Spirulina</i> isolates | 119 |
| Table 6.2: The characteristics of the sequential extracts of the <i>Spirulina</i> isolates | 120 |
| Table 6.3: Preliminary phytochemical screening of different sequential extracts..... | 121 |
| Table 6.4: Effect of drying conditions for the <i>Spirulina</i> sp. | 124 |
| Table 6.5: Fatty acid composition of all isolates of <i>Spirulina</i> using various forms of organic solvents..... | 127 |
| Table 6.6: Sulfolipid extraction from different isolates of <i>Spirulina</i> by using various solvent systems..... | 133 |
| Table 7.1: Quantity of SQDG from different isolates of <i>Spirulina</i> | 148 |

| | |
|--|-----|
| Table 7.2: Quantity and recovery of SQDG by using various forms of organic solvents for the <i>Spirulina</i> (M) | 149 |
| Table 7.3: Collection of different fraction through the column..... | 151 |
| Table 7.4: Anti-bacterial activity of sequential extracts of <i>Spirulina</i> | 151 |

ABBREVIATION

| | |
|--------------------------------|--|
| °C | Degree Celsius |
| α | Alpha |
| AIDS | Acquired Immunodeficiency Syndrome |
| AFDW | Ash free dry weight |
| ATCC | American Type Culture Collection |
| BLAST | Basic Local Alignment Search Tool |
| bp | Base pairs |
| BuOH | Butanol |
| C | Cytosine |
| CaCl ₂ | Calcium chloride |
| cART | Combination Anti-retroviral Therapy |
| CHCl ₃ | Chloroform |
| chl | Chlorophyll |
| CSIRO | Commonwealth Scientific and Industrial Research Organization |
| DGDG | Digalactosyldiacylglycerol |
| DMSO | Dimethyl sulphoxide |
| DNA | Deoxyribonucleic acid |
| dNTP | Deoxyribonucleotide triphosphate |
| EDTA | Ethylenediaminetetraacetic acid |
| ELSD | Evaporative light scattering detector |
| EPA | Eicosapentaenoic acid |
| EtOH | Ethanol |
| FAME | Fatty acid methyl esters |
| g | Gram |
| G | Guanine |
| GC | Gas chromatography |
| GLA | Gamma linolenic acid |
| H ₂ SO ₄ | Sulphuric acid |
| HCl | Hydrochloric acid |
| HIV | Human immunodeficiency virus |

| | |
|-------------------|---|
| HPLC | High pressure liquid chromatography |
| HSV | Herpes simplex virus |
| IMVS | Institute of Medical and Veterinary Science |
| IUPAC | International Union of Pure and Applied Chemistry |
| kb | Kilobases |
| L | Litre |
| m | Milli |
| MeOH | Methanol |
| mg | Milligrams |
| MgCl ₂ | Magnesium chloride |
| MGDG | Monogalactosyldiacylglycerol |
| min | Minute |
| MS | Mass spectrometry |
| NaAc | Sodium acetate |
| NaCl | Sodium chloride |
| NCBI | National Centre for Biotechnology Information |
| NMR | Nuclear magnetic resonance |
| NNRTIs | Non-nucleoside reverse-transcriptase inhibitors |
| NRPS | Non-ribosomal peptide synthase |
| NRTIs | Nucleoside reverse-transcriptase inhibitors |
| PCC | Pasteur culture collection |
| PCR | Polymerase chain reaction |
| PKS | Polyketide synthase |
| PUFA | Poly unsaturated fatty acids |
| rRNA | Ribosomal RNA |
| RNA | Ribonucleic acid |
| rpm | Revolution per minute |
| s | Second |
| SARDI | South Australian Research and Development Institute |
| SDS | Sodium dodecyl sulphate |
| SQDG | Sulfoquinovosyldiacylglycerol |
| TAE | Tris-acetate-EDTA |
| TE | Tris-EDTA |

| | |
|------|---------------------------------|
| Tris | Tris hydroxymethyl aminomethane |
| TLC | Thin layer chromatography |
| UDP | Uridine diphosphate |
| UNSW | University of New South Wales |
| UV | Ultraviolet |
| V | Volt |
| v/v | Volume per volume |
| WH | Woods Hole |
| w/v | Weight per volume |
| XS | Xanthogenate-SDS |