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Novel Role for the *Streptococcus pneumoniae* Toxin Pneumolysin in the Assembly of Biofilms

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**ABSTRACT** *Streptococcus pneumoniae* is an important commensal and pathogen responsible for almost a million deaths annually in children under five. The formation of biofilms by *S. pneumoniae* is important in nasopharyngeal colonization, pneumonia, and otitis media. Pneumolysin (Ply) is a toxin that contributes significantly to the virulence of *S. pneumoniae* and is an important candidate as a serotype-independent vaccine target. Having previously demonstrated that a luxS knockout mutant was unable to form early biofilms and expressed less ply mRNA than the wild type, we conducted a study to investigate the role of Ply in biofilm formation. We found that Ply was expressed in early phases of biofilm development and localized to cellular aggregates as early as 4 h postinoculation. *S. pneumoniae ply* knockout mutants in D39 and TIGR4 backgrounds produced significantly less biofilm biomass than wild-type strains at early time points, both on polystyrene and on human respiratory epithelial cells, cultured under static or continuous-flow conditions. Ply’s role in biofilm formation appears to be independent of its hemolytic activity, as *S. pneumoniae* serotype 1 strains, which produce a nonhemolytic variant of Ply, were still able to form biofilms. Transmission electron microscopy of biofilms grown on A549 lung cells using immunogold demonstrated that Ply was located both on the surfaces of pneumococcal cells and in the extracellular biofilm matrix. Altogether, our studies demonstrate a novel role for pneumolysin in the assembly of *S. pneumoniae* biofilms that is likely important during both carriage and disease and therefore significant for pneumolysin-targeting vaccines under development.

**IMPORTANCE** The bacterium *Streptococcus pneumoniae* (commonly known as the pneumococcus) is commonly carried in the human nasopharynx and can spread to other body sites to cause disease. In the nasopharynx, middle ear, and lungs, the pneumococcus forms multicellular surface-associated structures called biofilms. Pneumolysin is an important toxin produced by almost all *S. pneumoniae* strains, extensively studied for its ability to cause damage to human tissue. In this paper, we demonstrate that pneumolysin has a previously unrecognized role in biofilm formation by showing that strains without pneumolysin are unable to form the same amount of biofilm on plastic and human cell substrates. Furthermore, we show that the role of pneumolysin in biofilm formation is separate from the hemolytic activity responsible for tissue damage during pneumococcal diseases. This novel role for pneumolysin suggests that pneumococcal vaccines directed against this protein should be investigated for their potential impact on biofilms formed during carriage and disease.
otic cell membranes and forms 400-Å pores that lead to cell lysis (12). Through its activity at the bacterial cell surface, or upon release through pneumococcal autolysis, Ply is responsible for almost all pneumococcal hemolytic activity (13). As anti-Ply antibodies are generated following carriage or otitis media (14–16) and TLR4-mediated recognition of pneumolysin stimulates a profound immune response (17, 18), a detoxified derivative of pneumolysin is under investigation as a serotype-independent vaccine candidate (19–21). However, it is currently unclear what role Ply plays during asymptomatic carriage that exposes this protein to the host immune system.

Though prior studies have demonstrated that the hemolytic alpha-toxin of Staphylococcus aureus is essential for biofilm formation on plastic and ex vivo porcine mucosa (22, 23), we are unaware of any other hemolysins being directly implicated in biofilm formation. That said, some preliminary data suggest that Ply has a role in pneumococcal biofilm development. A study of S. pneumoniae ATCC 6303 biofilms in a continuous-flow system demonstrated more Ply present in 3-day-old biofilms than in planktonic cultures (24), and more recently, Marks et al. reported that 48-h biofilms formed by a Ply-deficient mutant, grown on NCI-H292 epithelial cells with medium replacement every 12 hours, differed in appearance and biomass from wild-type D39 (25). Furthermore, our previous study of the role of LuxS/AI-2 in biofilm development suggests that genes regulated by LuxS, including ply (7, 26), the putative hemolysin SP1466 (26), and the cbpD gene, encoding choline-binding protein D (CbpD), may be important in biofilm development. While it has been shown that CbpD is required for biofilm formation (8), the role of pneumolysin in biofilm development has not been thoroughly investigated.

To assess whether Ply has a part in pneumococcal biofilm formation, we examined the timing and distribution of Ply expression in pneumococcal biofilms and evaluated biofilm formation of Ply knockout mutants under static and continuous-flow conditions, on both abiotic and human cell substrates. We found that Ply is expressed in pneumococcal biofilms, localizing to the earliest aggregates of cells, and knockout mutants deficient in Ply are impaired in their ability to form biofilms. While biofilm formation appears to mask the hemolytic activity of pneumococcal cells, pneumolysin’s role in biofilm formation appears to be separate from its hemolytic activity. Altogether, our results demonstrate a novel and important role for Ply in the early development of pneumococcal biofilms.

**RESULTS**

**Pneumolysin is expressed in pneumococcal biofilms.** We first examined whether Ply was expressed in wild-type D39 during biofilm development in static cultures on a polystyrene substrate. To visualize Ply in biofilms, we employed a mouse monoclonal anti-Ply antibody followed by a fluorescently labeled anti-mouse antibody. Immunofluorescence images revealed low levels of Ply expression in biofilms 4 h and 6 h postinoculation, followed by maximal expression 8 h postinoculation and decreased expression.
FIG 2 Location of pneumolysin in pneumococcal biofilms. (A) Confocal microscopy images of D39 (antipneumococcal antibody, green) and Ply (anti-Ply antibody, red) indicate that pneumococcal cells and pneumolysin colocalized in D39 biofilms at 24 h. (B) An image of differential interference contrast (Nomarski) with a fluorescent anti-Ply primary antibody and a gold-conjugated secondary antibody reveals that Ply is located on the bacterial surface (arrow) and in the extracellular matrix. (C) Transmission electron microscopy of an 8-h biofilm grown on A549 cells and treated with anti-Ply primary antibody and a fluorescently labeled secondary antibody reveals that Ply is localized to nascent biofilm structures (i.e., early bacterial aggregates) and to collagen (Fig. 2D). Altogether, these results confirm the specific localization of Ply to nascent biofilm structures and suggest a role as a matrix protein.

Early biofilm assembly on polystyrene is impaired in a Ply-deficient knockout. To examine whether Ply is essential for assembly of pneumococcal biofilms, we constructed a Ply-deficient knockout in D39 (see Fig. S1 in the supplemental material) and examined the biofilm formation of this mutant as well as a TIGR4 derivative provided by Andrew Camilli (Table 1). While the planktonic growth of the Ply-deficient mutants did not differ from that of the corresponding wild-type strains (see Fig. S3 in the supplemental material), the ability of the Δply derivatives to form biofilms was drastically impaired. Fluorescence microscopy of GFP-expressing derivatives of D39, D39Δply, AC2394 (TIGR4), and AC4037 (TIGR4Δply) demonstrated that biofilm formation ability of the Δply mutants was inferior to that of the wild-type

TABLE 1 Strains and plasmids used in this study

<table>
<thead>
<tr>
<th>Strain or plasmid</th>
<th>Description</th>
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<tr>
<td><strong>S. pneumoniae strains</strong></td>
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<td>pMV158GFP</td>
<td>S. pneumoniae mobilizable plasmid encoding GFP; confers resistance to tetracycline</td>
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strains at 4 and 8 h (Fig. 3A). When absolute biofilm biomass was examined using a fluorescein isothiocyanate (FITC)-conjugated anti-pneumococcal antibody, we found that D39Δply produced significantly less biomass than the wild type (WT) at 6, 8, 10, and 12 h ($P < 0.05$; Fig. 3B). While a comC knockout mutant formed wild-type levels of biofilm biomass on abiotic surfaces, the luxS and ply knockout mutants formed significantly less biofilm biomass than the wild type ($P < 0.01$) (Fig. 3C). Similar results were obtained when biofilm assays were conducted using polystyrene substrates and Dulbecco’s modified Eagle medium (DMEM) (data not shown). As a control, we examined the previously characterized complemented ply mutant Pn + (13) and found that its ability to form biofilms did not differ significantly from that of the wild type (Fig. 3C).

To ascertain whether Ply released into the medium could be sufficient to restore biofilm formation capability to cells lacking endogenous Ply, we conducted experiments using Transwell permeable supports to separate strains of bacteria while allowing the free flow of nutrients and proteins. We grew three combinations of strains in wells with Transwell supports (given in the format top/bottom), WT/WT, WT/Δply mutant, and Δply mutant/Δply mutant. These wells were incubated at 37°C for 10 h, and biofilms on the bottom of the wells were quantified using GFP fluorescence. We found that the Δply strain with wild-type D39 above the Transwell barrier grew no more biofilms than the Δply strain with Δply organisms above the Transwell barrier (Fig. 3C).

**The pneumolysin deficient mutant forms comparable biofilm biomass by 24 h, and cellular autolysis is delayed.** Quantification of biofilms formed at 18 and 24 h using a fluorescently conjugated antipneumococcal antibody revealed that comparable biofilm biomass was found in wild-type and Δply strains (Fig. 4A). However, when biofilms were resuspended in phosphate-buffered saline (PBS) and plated on blood agar plates (BAPs), we found that more CFUs were recoverable from the wild-type strain at 8 h and 18 h postinoculation, but at 24 h postinoculation, there were no CFUs recoverable from wild-type D39 biofilms, while D39Δply and D39ΔluxS biofilms still yielded $\sim 1 \times 10^7$ CFU/ml (Fig. 4B). Fluorescence microscopy of GFP-expressing D39 and D39Δply at 24 h revealed that the Δply strain was more metabolically active (as evidenced by expression of GFP) than the wild-type strain (Fig. 4C). Electron microscopy images directly illustrated pneumococcal autolysis; while wild-type D39 cells in 8-h biofilms contained electron-dense material, wild-type D39 cells in 24-h biofilms have disrupted cellular membranes and exhibit an absence of electron-dense material (Fig. 4D). TEM of the ply knockout mutant demonstrated healthy electron-dense cells at both 8 h and 24 h and also showed more surface-associated polysaccharide than in the wild type at 8 h (Fig. 4D).

**Ply-deficient D39 forms less biofilm than the wild type on human cell line substrates.** To verify that pneumolysin is necessary for early biofilm formation under more physiologically relevant conditions, we examined biofilm formation and ply expression in biofilms grown on human cell substrates under static and continuous-flow conditions. When biofilm formation was assessed using an A549 human lung cell substrate in static growth conditions, we examined biofilm formation and ply expression in biofilms grown on human cell substrates under static and continuous-flow conditions. When biofilm formation was assessed using an A549 human lung cell substrate in static growth conditions, we observed that wild-type D39 formed confluent biofilms (Fig. 5A), while D39Δply formed very little biofilm (Fig. 5B). Quantitatively, the biofilm biomass of D39Δply was significantly less than that of the wild type 6 h ($P < 0.05$) and 8 h ($P < 0.01$) postinoculation (Fig. 5C). In addition, when the strains

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**FIG 3** Ply knockout mutants form inferior biofilms on polystyrene at early time points. (A) GFP-expressing D39, D39Δply, AC2394 (TIGR4), and AC4037 (TIGR4Δply) were incubated on glass slides for 4 or 8 h, and biofilms were imaged with fluorescence microscopy. (B) Wild-type D39 and D39Δply biofilm biomasses on polystyrene plates at 6, 8, 10, and 12 h were quantified using a polyclonal FITC-conjugated anti-pneumococcal antibody. Asterisks indicate $P$ values of $<0.05$. (C) Biofilm biomass of D39ΔcomC, D39ΔluxS, D39Δply, and the complemented strain Pn + (13) after 8 h of incubation. In addition, biofilm biomass formed on the bottom of the well of Transwell experiments with the wild type above and below, the wild type above and the ply mutant below, and the ply mutant above and below the barrier, after 10 h of incubation. Values from both the 8-h single-well experiments and 10-h Transwell experiments are expressed as percentages of the biofilm biomass of wild-type D39 at the respective time points. Error bars indicate standard errors of the means, and asterisks indicate $P$ values of $<0.01$. 
were grown on top of Detroit 562 pharyngeal cells for 8 h, we found that D39/H9004 and D39/luxS formed significantly less biofilm biomass than the wild type (P < 0.01) (Fig. 5D).

To better simulate in vivo conditions of pneumococcal biofilm growth, we grew D39 biofilms on an A549 substrate in a continuous flow bioreactor previously described (27). Confocal microscopy studies demonstrated that Ply coated the bacterial cell wall, with the top section showing pneumococcal chains surrounded by Ply (Fig. 6A) and a middle section showing abundant localization of Ply within aggregated bacteria (Fig. 6B). In relation to the log-phase planktonic inoculum, ply expression was upregulated ~6-fold in cells grown on immobilized A549 under static conditions for 8 h, ~2-fold in biofilms grown for 8 h in a bioreactor, and ~6-fold in biofilms grown for 24 h in a bioreactor (Fig. 6C).

Pneumococcal hemolytic activity is not essential for biofilm formation. To examine whether the biofilm formation function of pneumolysin was related to its hemolytic function, we examined the biofilm formation capability of serotype 1 strains belonging to sequence type (ST) 306 that produce a variant of pneumolysin with minimal hemolytic activity (28). Although Western blot analysis confirmed expression of Ply in the serotype 1 strains (see Fig. S4 in the supplemental material), hemolytic activity assays demonstrated the absence of hemolytic activity in D39/Dply and the serotype 1 isolates CDC_881, CDC_1403, and CDC_4829 (Fig. 7A). However, these three serotype 1 strains displayed no statistically significant difference in biofilm biomass at 8 h compared to wild-type D39 (Fig. 7B). Biofilm formation capacity is known to vary markedly between pneumococcal strains. Thus, to ensure that the different genetic background of the serotype 1 strains did not affect the experiment, we constructed Ply306, a D39 derivative expressing the nonhemolytic ply allele from ST 306, using methods described previously (29). We found that the biofilm biomass formed by the Ply306 D39 strain did not differ significantly from that of wild-type D39 (Fig. 7B). Taken together, these results suggest that Ply’s role in early biofilm formation is independent of hemolytic activity.

As pneumococcal biofilms are present in the nasopharynx with no apparent epithelial damage, the incorporation of Ply into the biofilm matrix may interfere with its hemolytic and cytotoxic activity. We therefore examined the hemolytic activity of wild-type D39 harvested from planktonic cultures in comparison to wild-type D39 harvested from biofilms. While D39 harvested from planktonic cultures maintained constant hemolytic activity over time (Fig. 7C and D), the hemolytic activity of D39 in biofilms

**FIG 4** Ply-deficient D39 biofilm formation is comparable to that of the wild type by 24 h, but cellular autolysis is delayed. (A) Biofilm biomass of D39 and D39/Dply grown on polystyrene for 12, 18, and 24 h, quantified using a FITC-conjugated antipneumococcal antibody. (B) Biofilms formed by D39, D39/DluxS, and D39/Dply on polystyrene after 8, 16, or 24 h of incubation were resuspended in 1 ml of PBS and plated on BAP to estimate CFU. The limit of detection of this assay was 10 CFU/ml. (C) Fluorescence microscopy of GFP-expressing D39 and D39/Dply confirms that the number of enzymatically active cells is greater in D39/Dply biofilms than in wild-type biofilms following 24 h of static incubation. (D) Transmission electron microscopy of wild-type D39 and the Dply mutant grown on A549 cells demonstrates healthy D39 at 8 h and autolyzed cells at 24 h and healthy Dply cells at both 8 and 24 h. Error bars indicate standard errors of the means.
Discussions

This study is the first to demonstrate that pneumolysin is essential for the production of early pneumococcal biofilms. Not only was Ply expressed in pneumococcal biofilms produced under static and continuous-flow conditions on abiotic and human cell substrates, but Ply knockout mutants produced significantly less biofilm biomass than the wild type at early time points. Early aggregates of pneumococci were the first to express pneumolysin, and expression peaked with maximal biofilm formation at approximately 8 to 10 h postinoculation. Incorporation of cells into biofilms led to a decrease in hemolytic activity, but the hemolytic function of Ply appears to be separate from its role in biofilm formation. In total, we have demonstrated that pneumolysin has a previously unrecognized role, key in the early stages of the biofilm formation process.

Previous studies have alluded to a connection between pneumolysin and biofilm formation. Most evidence has been circumstantial, demonstrating that biofilm formation results in changes in ply expression. In 2006, Oggioni et al. reported extensive regulatory differences between planktonic and sessile phases, including the finding that in comparison to mid-exponential-phase liquid cultures, pneumococcal cells on agar and in biofilms expressed 100-fold and 5-fold less ply, respectively (30). Lizcano et al. concluded that pneumolysin was not important in biofilm formation in TIGR4; however, their study only examined biofilm biomass by measuring crystal violet staining at 18 h postinoculation (31). Our results indicate that after 18 h of growth, cells in wild-type biofilms may have already begun to undergo autolysis, while the Ply knockout mutant continues to form more biofilm (7, 32). By examining biofilm formation of wild-type strains and isogenic mutants using various growth conditions, multiple substrates, and a variety of time points, we have shown that there is an important role for pneumolysin in the early stages of pneumococcal biofilm development.

Confocal and electron microscopy localized pneumolysin to the pneumococcal cell surface and extracellular matrix, suggesting a linking role for this protein during the aggregation phase of biofilm formation. While altered degradation of GFP may be responsible for the difference in autolysis witnessed by fluorescence, our CFU measurements and TEM results clearly demonstrate that the ply knockout mutant has delayed autolysis. This may be a function of metabolic changes, perhaps resulting from altered proximity to other bacterial cells. Previous studies have indicated that presence of a capsule impairs pneumococcal biofilm formation (33, 34) and that genes in the capsule operon are down-regulated during biofilm formation (35). The polysaccharide halo surrounding the ply knockout cells imaged by TEM is consistent with altered capsular regulation, but further studies are needed to understand the role of capsular expression in pneumococcal biofilm formation.

Pneumolysin has been recognized as a virulence determinant for decades (36), and animal and human studies have indicated that the hemolytic activity of pneumolysin is largely responsible for pneumococcal virulence (37). However, the amount of hemolytic activity necessary for virulence is reportedly just 0.1% of the wild-type levels (13). Nevertheless, it was surprising when Kirkham et al. reported the isolation of nonhemolytic serotype 1 strains from invasive pneumococcal disease (28). This result led
those authors to speculate that an immune activation function of pneumolysin may contribute to virulence; this hypothesis is supported by the findings of Malley et al. showing that pneumolysin induces a substantial immune response by macrophages (17). The results of our study raise the possibility that pneumolysin’s contribution to pneumococcal virulence may be partially attributed to its role in biofilm formation. At the same time, our finding that pneumococcal hemolytic activity of cells decreases as biofilms are formed may indicate that incorporation of cells into the biofilm matrix may mask the hemolytic epitope and prevent pneumolysin from provoking a clearing immune response during carriage.

Recently we demonstrated that LuxS/AI-2 regulates early pneumococcal biofilm formation (7, 8) and observed that a luxS null mutant expressed lower mRNA levels of the pneumolysin gene than the wild type. In a follow-up study, we demonstrated that both the LuxS/AI-2 and Com quorum-sensing systems regulate biofilm production in a bioreactor with living cultures of human respiratory cells (27). Our finding that pneumolysin is down-regulated in a luxS knockout mutant but not in a comC knockout mutant appears to indicate that pneumolysin is predominantly under the regulatory control of the LuxS/AI-2 system. From these results we can hypothesize that LuxS-regulated pneumolysin plays a critical role in early aggregation, while other factors regulated by Com contribute to biofilm maturation. The components of biofilm assembly and maturation, and the regulation of those components, are a subject in need of further study.

Our results suggest that pneumolysin directly contributes to early aggregation of pneumococcal cells into early biofilms. Given that pneumolysin is an important candidate antigen for the development of a serotype-independent vaccine (19–21, 38, 39), our

FIG 6 Pneumolysin is expressed in biofilms produced on human lung cells in a continuous-flow bioreactor. (A) The optical top section from a bioreactor with wild-type D39 biofilm cells on an A549 human cell substrate after 8 h of growth. Cells and biofilms were fixed, nuclei were fluorescently stained with TO-PRO-3 (blue), and Ply was stained with an anti-Ply antibody (red). Arrows indicate GFP-expressing pneumococci (green) surrounded by Ply, enlarged in the inset. (B) The middle section from the same 8-h bioreactor demonstrates abundant Ply expressed within pneumococcal biofilm aggregates grown in this model system. (C) Reverse transcription and qPCR of RNA extracted from biofilms grown on immobilized cells in static culture for 8 h or in a continuous-flow bioreactor for 8 or 24 h demonstrate that ply is upregulated relative to the inoculum when the bacteria are grown on A549 cells. Error bars indicate standard errors of the means.

FIG 7 Hemolytic activity and biofilm phenotypes. (A) D39, D39Δply, and three serotype 1 isolates were examined for hemolytic activity. Wild-type D39 possessed hemolytic activity, while all other strains examined had almost none. (B) When biofilm formation capability was assessed, CDC_881, CDC_1403, CDC_4829, and Ply306 exhibited biofilm biomasses similar to that of wild-type D39. (C) Wild-type D39 cells harvested from planktonic and biofilm cultures assayed for hemolytic activity did not differ in hemolytic activity at 6 h postinoculation. (D) Planktonic and biofilm cultures of D39 harvested 10 h postinoculation demonstrate a marked difference in hemolytic activity. Error bars indicate standard deviations, and the asterisk indicates a \( P \) value of <0.01.
characterization of pneumolysin’s role in biofilm formation has important implications. Carriage of the pneumococcus is a necessary precursor to pneumococcal diseases (40) and if Ply is routinely expressed in nasopharyngeal biofilms, its potential as a vaccine antigen is more easily understood. These results highlight the urgent need for further exploration of pneumolysin and the immune response against it, using both in vivo and in vitro models of biofilm formation.

MATERIALS AND METHODS

Strains and bacterial culture methods. The S. pneumoniae reference and derivative strains used in this study are listed in Table 1. Strains were cultured on Trypticase soy agar BAPs or in Todd-Hewitt broth containing 0.5% (wt/vol) yeast extract (THY). When biofilm formation was visualized or quantified using GFP-expressing strains, 2% (wt/vol) maltose was added to the culture medium. Inocula for all biofilm assays were prepared using growth on an overnight BAP to prepare a cell suspension in THY broth at an optical density at 600 nm (OD_{600}) of 0.05. This suspension was incubated at 37°C in a 5% CO_2 atmosphere until the culture reached an OD_{600} of 0.2 to 0.3 (early log phase); glycerol was added to a final concentration of 10% (vol/vol), and the suspension was stored at −80°C until used.

Growth of biofilms on abiotic and biotic surfaces. For experiments conducted on abiotic surfaces, 24-well Costar polystyrene plates (Corning, Tewksbury, MA) were used for quantification experiments, and 8-well Lab-Tek II glass chamber slides (Thermo, Fisher Scientific, Rockford, IL) were used for visualization experiments. Wells or chambers were filled with THY medium, inoculated 1:10 with inocula prepared as described above, and incubated at 37°C for various lengths of time. When indicated, experiments used Transwell permeable supports (Corning, Tewksbury, MA) with 0.4-μm pores to separate strains of bacteria within wells of 24-well plates while allowing the free flow of nutrients and secreted proteins.

To simulate the environment in the human respiratory tract, a human cell line substrate was employed, in both static and continuous-flow cultures, as recently described (27). Briefly, human-derived lung A549 cells (ATCC CCL-185) or human-derived pharyngeal Detroit 562 cells (ATCC CCL-198) were grown on polystyrene plates or Snapwell filters (Corning, Tewksbury, MA) until confluent (4 to 5 days). For static experiments, human cells were fixed with 2% paraformaldehyde (Sigma) for 15 min. Following several washes with PBS, immobilized human cells were inoculated with prepared bacterial inocula (−7 × 10^6 CFU/ml) and incubated in DMEM at 37°C for various times. For continuous-flow experiments, confluent cells on Snapwell filters were inoculated as detailed above and immediately placed in a sterile vertical diffusion chamber (27). Both basolateral and apical sides (inner chamber) were perfused with sterile DMEM with no antibiotics using a Master Flex L/S precision pump system (Cole-Parmer, Vernon, IL) at a low flow rate (0.20 ml/min). Bioreactor chambers containing S. pneumoniae and lung cells were incubated at 37°C in a sterile environment. At the end of the incubation period, inserts containing biofilms were removed, and biomass was analyzed qualitatively and quantitatively.

Visualizing pneumococcal biofilms. When GFP-expressing strains were used, biofilms on microtiter plates were washed with PBS and visualized by fluorescence microscopy, or biofilms produced on 8-chamber slides were washed with PBS, fixed with 2% paraformaldehyde, and mounted with Vectashield (Vector Laboratories, Burlingame, CA), and fluorescence was visualized with an inverted Evos FL microscope (Advanced Microscopy Group, Carlsbad, CA) or confocal microscopy. When non-GFP-expressing strains were used, biofilms were fixed with 2% paraformaldehyde (Sigma-Aldrich, St. Louis, MO) for 15 min, washed with PBS, blocked with 2% bovine serum albumin, and stained for 1 h at room temperature with a polyclonal anti-S. pneumoniae antibody (−40 μg/ml) coupled to fluorescein isothiocyanate (FITC; ViroStat, Portland, ME). Where indicated, Ply was detected using 2.0 μg/ml unconjugated mouse monoclonal anti-Ply antibody (Santa Cruz Biotechnology, Santa Cruz, CA), followed by an Alexa Fluor 568 goat anti-mouse IgG secondary antibody (Molecular Probes, Invitrogen, Carlsbad, CA). For preparations from the human cell bioreactor system, sialic acid residues present on the plasma membranes were stained with 5 μg/ml wheat germ agglutinin conjugated with Alexa Fluor 555 (Molecular Probes, Invitrogen, Carlsbad, CA) for 30 min (41), and nucleic acids were stained with TO-PRO-3 (1 μM), a carboxyfluorescein monomer nucleic acid stain (Molecular Probes, Invitrogen, Carlsbad, CA) for 15 min. Finally, preparations were washed three times with PBS, mounted with Vectashield mounting medium (Vector Laboratories, Burlingame, CA), and analyzed with a Zeiss LSM510 confocal microscope. Confocal images were analyzed with an LSM image browser, version 4.0.2.121.

For immunogold localization of Ply, 8-h biofilms grown on top of A549 lung cells were fixed with either 2 or 4% paraformaldehyde (PFA) in 0.1 M phosphate buffer (PB) overnight at 4°C. Cells were then washed and treated with 0.05% Triton X-100 for 10 min before blocking with PBS containing 5% bovine serum albumin (BSA) and 0.1% cold-water-fish gelatin. Mouse anti-Ply primary antibody was diluted to 8 μg/ml in PBS containing 0.1% acetylated BSA for overnight incubation. After washes, cells were incubated overnight with ultrasmall-collodial-gold-conjugated goat anti-mouse IgG secondary antibody (Aurion, Wageningen, the Netherlands), followed by washes and postfixation in 2.5% buffered glutaraldehyde. Silver enhancement of ultrasmall gold particles was carried out using R-gent SE-EM silver enhancement kit (Aurion, Wageningen, the Netherlands) following the manufacturer’s instruction. Cells were then fixed with 0.5% osmium tetroxide, dehydrated, and embedded in Eponate 12 resin. Ultrathin sections were counterstained with 5% uranyl acetate and 2% lead citrate and examined on a JEOL JEM-1400 transmission electron microscope (JEOL Ltd., Japan) equipped with a Gatan UltraScan US1000 charge-coupled device (CCD) camera (Gatan, Inc., Pleasanton, CA).

Quantification of biofilm biomass. Biofilm biomass was quantified by fluorescence methods as previously described (7, 27) or by serial dilution followed by plating to obtain cell counts (CFU/ml). For GFP-expressing strains and biofilms stained with a FITC-conjugated antibody, arbitrary relative fluorescence units (RFU) were obtained using a Victor X3 multilabel plate reader (PerkinElmer, Waltham, MA). The number of arbitrary fluorescence units of wild-type D39 was set to a biofilm biomass of 100% and used to calculate the biofilm biomass percentages of all of the other S. pneumoniae strains tested and those at different time points. Results of repeated experiments were plotted using GraphPad Prism or SigmaPlot and examined for statistical significance with two-tailed t tests for normal data and Mann-Whitney U tests for nonparametric data.

Western blot assays. To compare the expression of Ply in biofilm and planktonic cells, bacterial cells were treated with the B-PER bacterial protein extraction reagent (Thermo, Fisher Scientific, Rockford, IL) according to the manufacturer’s protocol and quantified using the Bradford protein assay (42). A 5-μg portion of extracted protein was loaded into each well on a 12% polyacrylamide gel, subjected to SDS-PAGE, and transferred onto a nitrocellulose membrane. Those membranes were blocked with PBS with 0.05% (vol/vol) Tween 20 and 5% (wt/vol) nonfat dry milk for 1 h and then probed with a 1:200 dilution (final concentration, 0.5 μg/ml) of mouse anti-Ply monoclonal antibody (Santa Cruz Biotechnology, Santa Cruz, CA) (43). Bound antibody was detected with a horseradish peroxidase-conjugated secondary anti-mouse antibody diluted 1:10,000 and addition of Pierce ECL Western blotting substrate (Thermo, Fisher Scientific, Rockford, IL).

Construction of D39 derivatives with ply knocked out or modified. A ply knockout mutant was generated by PCR ligation mutagenesis as previously described (44) (see Fig. S1 in the supplemental material). Briefly, the construct was generated by PCR amplification of a 5’ segment of the ply gene with primers HPL1 and HPL2 and a 3’ segment with primers HPL3 and HPL6. Primers HPL3 and HPL4 were used to amplify the ermB gene, encoding erythromycin resistance, and to add XbaI and...
TABLE 2 Primers used in this study

<table>
<thead>
<tr>
<th>Primer</th>
<th>Target gene</th>
<th>Sequence&lt;sup&gt;a&lt;/sup&gt;</th>
<th>RE site</th>
<th>PCR product size (bp)</th>
</tr>
</thead>
<tbody>
<tr>
<td>HPL1L</td>
<td>SPD1728-ply</td>
<td>TGGCGGACAAGCATTTTGA</td>
<td>XbaI</td>
<td>800</td>
</tr>
<tr>
<td>HPL2R</td>
<td>ply-SPD1728</td>
<td>CAGTCTAGACCACTAGGAGATGCTCCA</td>
<td>XbaI</td>
<td>796</td>
</tr>
<tr>
<td>HPL3L</td>
<td>ermB</td>
<td>CAGTCTAGAAATTTTGGTAATAGAGGAGT</td>
<td>XbaI</td>
<td>800</td>
</tr>
<tr>
<td>HPL4R</td>
<td>ermB</td>
<td>CAGCTCGAGCCAAATTTACCAAAAGGGACTCA</td>
<td>XbaI</td>
<td>800</td>
</tr>
<tr>
<td>HPL5L</td>
<td>ply-flank</td>
<td>CAGCTCGAGCTTTAAAAGGGAATGTTCGTAATCTCT</td>
<td>XbaI</td>
<td>800</td>
</tr>
<tr>
<td>HPL6R</td>
<td>Flank-ply</td>
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<td>XbaI</td>
<td>800</td>
</tr>
<tr>
<td>JVS35L</td>
<td>16S rRNA</td>
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<td>XbaI</td>
<td>126</td>
</tr>
<tr>
<td>JVS36R</td>
<td>16S rRNA</td>
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<td>XbaI</td>
<td>225</td>
</tr>
<tr>
<td>JVS59L</td>
<td>ply</td>
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<td>XbaI</td>
<td>225</td>
</tr>
<tr>
<td>JVS60R</td>
<td>ply</td>
<td>CTAATTGGACAGAGAGATTTGAGTAAGA</td>
<td>XbaI</td>
<td>257</td>
</tr>
<tr>
<td>luxS-L</td>
<td>luxS</td>
<td>ACATCACCTCCAAATTTGATATTC</td>
<td>XbaI</td>
<td></td>
</tr>
<tr>
<td>luxS-R</td>
<td>luxS</td>
<td>GACATCTTCCAAATTTGATATTC</td>
<td>XbaI</td>
<td></td>
</tr>
</tbody>
</table>

<sup>a</sup> Underlining indicates the restriction enzyme (RE) site used for cloning.

XhoI restriction sites to the ends (7). The 5′ and 3′ segments were ligated to the gene encoding erythromycin resistance using T4 DNA ligase, and the cassette was generated by amplification with primers HPL1 and HPL6. A 100-ng portion of the cassette was then transformed into competent D39 cells, and the SPJV14 (D39<sup>ply</sup>) recombinants were selected for on BAP containing 0.5 μg/ml of erythromycin. Knockouts were confirmed by PCR (see Fig. S1 in the supplemental material), sequencing (data not shown), and Western blotting (see Fig. S4 in the supplemental material). A derivative of D39 expressing the nonhemolytic ply allele from an <i>S. pneumoniae</i> type 1 ST 306 strain was constructed by allelic replacement using an analogous protocol and the primers employed for construction of a derivative expressing the ply<sup>ply4496</sup> allele, which has significantly less hemolytic activity (29).

**Hemolytic activity assay.** Pneumococcal cells were prepared for assay by growth in THY to mid-log phase (OD<sub>600</sub> 0.3 to 0.4) and 10<sup>8</sup> concentration before resuspension in PBS. Bacterial cells were lysed with a 10-min incubation at 37°C in 0.1% sodium deoxycholate solution, and then protein was quantified using the Bradford assay. Sheep red blood cells (RBCs) were washed three times using PBS. RBCs (2 ml) were added to protein was quantified using the Bradford assay. Sheep red blood cells (2 ml) were added to

**Supplemental Material**

Supplemental material for this article may be found at http://mbio.asm.orglookup/suppl/doi:10.1128/mBio.00655-13/-/DCSupplemental.

Figure S1, EPS file, 1.2 MB.
Figure S2, EPS file, 14.8 MB.
Figure S3, EPS file, 1.2 MB.
Figure S4, EPS file, 1.2 MB.

**Acknowledgments**

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**References**


