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# Deletion of Interleukin-6 Signal Transducer gp130 in Small Sensory Neurons Attenuates Mechanonociception and Down-Regulates TRPA1 Expression

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Glycoprotein 130 (gp130) is the signal transducing receptor subunit for cytokines of the interleukin-6 (IL-6) family, and it is expressed in a multitude of cell types of the immune and nervous system. IL-6-like cytokines are not only key regulators of innate immunity and inflammation but are also essential factors for the differentiation and development of the somatosensory system. Mice with a null mutation of gp130 in primary nociceptive afferents (*SNS-gp130*<sup>−/−</sup>) are largely protected from hypersensitivity to mechanical stimuli in mouse models of pathological pain. Therefore, we set out to investigate how neuronal gp130 regulates mechanonociception. *SNS-gp130*<sup>−/−</sup> mice revealed reduced mechanosensitivity to high mechanical forces in the von Frey assay *in vivo*, and this was associated with a reduced sensitivity of nociceptive primary afferents *in vitro*. Together with these findings, transient receptor potential ankyrin 1 (TRPA1) mRNA expression was significantly reduced in DRG from *SNS-gp130*<sup>−/−</sup> mice. This was also reflected by a reduced number of neurons responding with calcium transients to TRPA1 agonists in primary DRG cultures. Downregulation of *Trpa1* expression was predominantly discovered in nonpeptidergic neurons, with the deficit becoming evident during stages of early postnatal development. Regulation of *Trpa1* mRNA expression levels downstream of gp130 involved the classical Janus kinase family-signal transducer and activator of transcription pathway. Our results closely link proinflammatory cytokines to the expression of TRPA1, both of which have been shown to contribute to hypersensitive pain states. We suggest that gp130 has an essential role in mechanonociception and in the regulation of TRPA1 expression.

**Key words:** cytokine; IL-6 signal transducer; JAK/STAT signaling; mechanonociception; mechanotransduction; pain

## Introduction

Neurotrophic cytokines of the interleukin-6 (IL-6) family are essential regulators of innate immunity and nerve regeneration.

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The cytokines act on their target cells via heteromeric receptor complexes that require glycoprotein 130 (gp130) as signal transducer subunit (Heinrich et al., 2003). In addition to IL-6, the family consists of IL-11, IL-27, leukemia inhibitory factor (LIF), ciliary neurotrophic factor (CNTF), oncostatin M (OSM), neurotrophin, cardiotrophin-1, and cardiotrophin-like cytokine (Bauer et al., 2007).

In addition to their role in innate immunity, gp130-targeting cytokines are important survival and differentiation factors. IL-6, CNTF, OSM, and LIF play crucial roles in the brain and in the peripheral nervous system (Murphy et al., 1991, 1993; Thompson et al., 1996; Banner et al., 1998; Nakashima et al., 1999; Zhong et al., 1999). OSM, for example, is essential for the development and survival of a subpopulation of TRPV1/P2X3-positive sensory neurons (Morikawa et al., 2004). However, OSM and IL-6, but not LIF, sensitize nociceptors to heat (Langeslag et al., 2011).

Thermal sensitization occurs as a consequence of fast modulation of the heat and capsaicin-sensitive ion-channel TRPV1, whereas ablation of gp130 in Na<sub>v</sub>1.8-expressing small-diameter

DRG neurons (*SNS-gp130<sup>-/-</sup>*) reduces thermal hypersensitivity in models of pathological pain (Obreja et al., 2002, 2005; Andratsch et al., 2009). Furthermore, IL-6 sensitizes nociceptors to noxious mechanical stimuli, and *SNS-gp130<sup>-/-</sup>* mice are largely protected and recover faster from mechanical hypersensitivity in models of inflammatory, neuropathic, and cancer pain (Brenn et al., 2007; Quarta et al., 2011).

It is generally accepted that the detection of noxious mechanical stimuli (mechanonociception) relies on the presence of mechanically activated ion channels in nociceptive neurons. Differential expression and sensitization of these putative ion channels very likely contribute to the mechanical hypersensitivity associated with chronic pain states. Despite the anticipated importance of such channels, the cellular and molecular mechanisms of mechanonociception are still incompletely understood (Lumpkin and Caterina, 2007; Dubin and Patapoutian, 2010; Delmas and Coste, 2013).

In the present study, we assessed mechanonociception in the *SNS-gp130<sup>-/-</sup>* mouse model *in vivo* and with an *in vitro* skin-nerve preparation. Moreover, a genome-wide mRNA expression profiling assay was used to screen for differential expression of nociceptor-specific ion-channel mRNA. The expression of candidates was assessed with quantitative TaqMan RT-PCR, ISH of DRG sections, as well as correlative functional assays. Here, we demonstrate that ablation of gp130 reduces the sensitivity of nociceptive fibers to noxious mechanical stimuli and that the presence of gp130 in small DRG neurons is essential for the expression of the TRPA1 ion channel.

## Materials and Methods

**Animals.** *SNS-gp130<sup>-/-</sup>* and *gp130<sup>fl/fl</sup>* mice were generated, bred, and genotyped as described previously (Andratsch et al., 2009). Briefly, *gp130<sup>fl/fl</sup>* mice were cross-bred with *SNS-Cre* mice (Agarwal et al., 2004) to obtain *SNS-Cre-gp130<sup>fl/fl</sup>* (*SNS-gp130<sup>-/-</sup>*) and *gp130<sup>fl/fl</sup>* mice. All mice were maintained under specific pathogen free conditions. *TRPA1<sup>-/-</sup>* mice were a generous gift from Dr. David Corey (Harvard Medical School, Boston) and used as controls where appropriate (Kwan et al., 2006). Unless otherwise stated, animals of either sex and older than 8 weeks were used in all experiments. For postnatal development studies, animals were staged by their date of birth. Day of birth was counted as postnatal day 0 (P0). Behavioral measurements and analyses were performed in awake, unrestrained, age-matched, male mice with an age of 8–16 weeks by examiners who were blinded to the genotype of the mice. All animal procedures were in accordance with ethical guidelines and animal welfare regulations according to Austrian law and with formal permission of the Austrian BMWF ministry (BMWF-66.011/0051-II/10b/2008; BMWF-66.011/0113-II/3b/2010).

**Mechanical sensitivity.** Mice were placed in a plastic chamber with a wire mesh floor and allowed to habituate for 1 h before starting the test. The lateral side of the plantar surface of the paw (sural nerve innervation territory) was stimulated by calibrated von Frey monofilaments. Filaments were applied in an ascending force order (1.4, 4, 8, 16, 22.6, 32, and 45.3 mN), and the frequency of paw withdrawal to 5 stimuli for each monofilament was recorded. The threshold was taken as the lowest force that evoked a brisk withdrawal response to two of five repetitive stimuli. Mechanical sensitivity was assessed on two separate days, and threshold values were averaged per animal. One animal in the *SNS-gp130<sup>-/-</sup>* group failed to fulfill the threshold criterion and was assigned the highest mechanical force (45.3 mN) as threshold.

**Nocifensive behavior.** Spontaneous pain-like behavior was monitored after intracutaneous injection of 20  $\mu$ l of 1% cinnamaldehyde (CA, Sigma) or vehicle (20% Tween 80, Sigma) in PBS (PAA) into the hindpaw of *gp130<sup>fl/fl</sup>* or *SNS-gp130<sup>-/-</sup>* mice. Mice were placed in a plastic chamber and allowed to habituate for at least 1 h before the experiment. Movies were taken with a Zx1 (Kodak) or an IN-3011 (Instar) camera through the translucent floor of the box so that the hindpaws were visible

at all times. Behavioral responses were scored for 5 min after injection. Vehicle controls were injected into the hindpaw contralateral to CA 1 week after the initial treatment, and the mice did not show a response during the analyzed time frame.

**Single fiber recordings.** An *in vitro* skin nerve preparation was used to investigate the properties of unmyelinated afferent nerve fibers innervating the skin of the mouse dorsal hindpaw as previously reported (Andratsch et al., 2009; Quarta et al., 2011). The saphenous nerve was dissected with the skin of the dorsal hindpaw attached and mounted in an organ bath “inside-up” to expose the corium side. The preparation was superfused (15 ml/min) with an oxygen-saturated modified synthetic interstitial fluid solution containing (in mM) as follows: 108 NaCl, 3.48 KCl, 3.5 MgSO<sub>4</sub>, 26 NaHCO<sub>3</sub>, 1.7 NaH<sub>2</sub>PO<sub>4</sub>, 2.0 CaCl<sub>2</sub>, 9.6 sodium glutamate, 5.5 glucose, and 7.6 sucrose at a temperature of 31  $\pm$  1°C and pH 7.4  $\pm$  0.05. The saphenous nerve was pulled into a separate chamber of the organ bath and placed on a small mirror. Using sharpened watchmakers’ forceps, fine filaments were teased from the desheathed nerve and placed on a gold wire recording electrode. Action potentials of single-nerve strands were recorded extracellularly, amplified (5000 $\times$ ), filtered (low pass 1 kHz, high pass 100 Hz), visualized on an oscilloscope, and stored on a PC-type computer with Spike/Spidi software package for offline analysis using a template-matching procedure. The fibers were characterized as unmyelinated (C) according to their conduction velocity (<2 m/s, calculated from the latency of unitary action potential to electrical stimulus at receptive field and distance of receptive field to recording electrode) and on the basis of the shape of the action potential. The receptive field of the primary afferent fiber was located by mechanical probing of the skin with a blunt glass rod. Only units with a signal-to-noise ratio >2 were used for further analysis. Fibers were subject to a standard protocol of adequate mechanical stimuli. The mechanical threshold of each unit was determined with a set of calibrated von Frey monofilaments with uniform tip diameter 1.1 mm and bending forces ranging from 1 to 362 mN. The strength of the finest filament that evoked at least 3 action potentials was defined as activation threshold.

**DRG culture.** DRG containing the cell bodies of primary afferents were harvested from adult mice as previously published (Andratsch et al., 2009; Camprubí-Robles et al., 2013). After removal of the connective tissue, ganglia were incubated in Liberase Blendzyme 1 (9 mg/100 ml DMEM, Roche) for 2 times at 30 min. After washing with PBS (PAA), 1 $\times$  trypsin-EDTA (Invitrogen) was added for 15 min, and DRG were washed three times with DMEM and once with TNB medium (Biochrom) supplemented with Protein-Lipid-Complex (Biochrom), L-glutamine, penicillin G sodium, and streptomycin sulfate (all from Invitrogen). The DRG were dissociated in TNB medium with a fire-polished Pasteur pipette and centrifuged through a 3.5% BSA gradient (Sigma) to eliminate non-neuronal cells and debris. The sensory neurons were resuspended, plated on either 6-well or 24-well culture dishes (Nunc) coated with poly-L-lysine/laminin-1 (Sigma), and cultivated in supplemented TNB at 37°C in 5% CO<sub>2</sub> for 24 h, unless otherwise indicated. For microfluorimetric calcium measurements, cells were plated on glass-bottom dishes coated with poly-L-lysine/laminin-1 (Sigma) and cultivated for 18–32 h. Exogenous mNGF 2.5S (Alomone Labs, 10  $\mu$ g/100 ml TNB medium) was added whenever indicated.

**In vitro inhibitor studies.** DRG were harvested from spinal segments T1 to L6, and cultures were prepared as described above. Dissociated neurons of one to three *gp130<sup>fl/fl</sup>* mice were pooled and subsequently divided into control and inhibitor samples. Inhibitor samples were first pretreated with non-toxic concentrations of Janus kinase family (JAK) inhibitor I (1  $\mu$ M, Calbiochem/Millipore) (Loy et al., 2011) or signal transducer and activator of transcription (STAT) inhibitor Stattic (1.5  $\mu$ M, Sigma) (S. Quarta, Medical University Innsbruck, Austria, personal communication) in TNB medium for 1 h and then incubated with the same concentration of inhibitor diluted in TNB medium containing exogenous mNGF 2.5S for 72 h. Inhibitors were dissolved in DMSO in a maximal final dilution of 1:1000. Control samples received an equal treatment with medium containing the appropriate concentration of vehicle.

**Genome-wide mRNA expression profiling assay.** DRG explants were prepared as described above. DRG samples of two littermate mice were always pooled, and a total of 10 *gp130<sup>fl/fl</sup>* and 10 *SNS-gp130<sup>-/-</sup>* mice

were divided into five groups per genotype. RNA isolation, cRNA preparation, and microarray measurement with subsequent analysis were performed by IMGM Laboratories using Mouse Genome Survey Microarray V2.0 with the 1700 Chemiluminescent Microarray Analyzer (Applied Biosystems). For genes to be included in the analysis, all five samples of at least one genotype had to obtain a signal-to-noise ratio of  $\geq 3$ . Functional annotation of the genes was based on the Panther2.0 database (www.pantherdb.org). All genes that belonged to the Panther\_Function “ion channel” or “ligand-gated ion channel” were grouped as “ion channels.” Only genes with net fold changes  $> 1.5$  were considered as differentially expressed between the two genotypes. Student's *t* test was applied to compare *SNS-gp130<sup>-/-</sup>* and *gp130<sup>fl/fl</sup>* groups. The *p* values were corrected for multiple testing using the algorithm of Benjamini and Hochberg, which is based on a control of the false discovery rate (FDR) (Benjamini and Hochberg, 1995). Microarray data are available in the ArrayExpress database (www.ebi.ac.uk/arrayexpress) under accession number E-MTAB-2584.

**qRT-PCR.** For analysis of mRNA expression levels, total RNA was isolated from murine DRG explants or primary cultures of DRG neurons by using TRI Reagent (Sigma) or peqGOLD TriFast (PeqLab) as previously published (Andratsch et al., 2009). DRG explants were lysed with a Tissue Lyser II (QIAGEN) or manually using plastic pestles. Primary cultures of DRG neurons were directly lysed on the culture dish and subsequently used for RNA extraction. The quantity of RNA was analyzed using a BioPhotometer (Eppendorf) or a Nanodrop 2000 (Thermo Scientific) and, where applicable, RNA quality was verified by gel electrophoresis. Reverse transcription to cDNA was performed using MuLV reverse transcriptase (2.5 U/ $\mu$ l, Applied Biosystems) with random hexamer primers (10 ng/ $\mu$ l), RiboLock (2 U/ $\mu$ l), 1 $\times$  Taq Buffer (all from Thermo Scientific), MgCl<sub>2</sub> (5 mM), and dNTPs (1 mM, both Fermentas). cDNA samples were analyzed for expression of target genes by quantitative real-time TaqMan PCR using TaqMan 5' nuclease assays (all from Applied Biosystems). With the exception of 18S (18S ribosomal RNA, Hs99999901\_s1), the following exon spanning assays were used: Mm01352363\_m1 (succinate dehydrogenase subunit A, *Sdha*), Mm00625268\_m1 (transient receptor potential ankyrin 1, *Trpa1*), Mm01246302\_m1 (transient receptor potential vanilloid 1, *Trpv1*), Mm00499025\_m1 (transient receptor potential vanilloid 4, *Trpv4*), and Mm99999915\_g1 (glyceraldehyde-3-phosphate dehydrogenase, *Gapdh*). Reactions were performed in a MicroAmp Fast Optical 96-Well Reaction Plate (Applied Biosystems) using the 7500 Fast Real-Time PCR System (Applied Biosystems) for thermal cycling and real-time fluorescence measurements. The PCR cycle protocol consisted of 10 min at 95°C, and 50 two-step cycles of 15 s each at 95°C and of 1 min at 60°. Positive and negative controls (no-template controls for RNA extraction, RT-PCR, and TaqMan-qPCR) were included in the experiments. Each sample was run in duplicates or triplicates. Threshold cycle (*C<sub>T</sub>*) values were recorded as a measure of initial template concentration. For duplicates, only samples with a *C<sub>T</sub>* value range  $< 0.5$  for the replicates were included in the analysis. Exceptions were made for *Trpa1* expression in DRG explants during postnatal days 1 and 2 because of the low expression of *TRPA1*. Relative fold changes in mRNA levels were calculated by the  $\Delta\Delta C_T$  method using *Sdha* as a reference standard. The range for the target gene relative to *gp130<sup>fl/fl</sup>* samples as calibrator was calculated by  $2^{-\Delta\Delta C_T}$ .

**ISH: probes.** A region complementary to base pairs 957–1761 of murine *Trpa1* mRNA (NM\_177781.4) was amplified from full-length cDNA by PCR. Specifically designed primer pairs introduced T7 (antisense probe: 5'-TAATACGACTCACTATAGGGGTTTCAGGAGGATGTCAG-CATTG-3') and SP6 (sense probe: 5'-CATACGATTTAGGTGACAC-TATAGGAGACCTGCTTCACAGAG-3') RNA polymerase promoter regions at the 5' ends of the amplified region. Amplified DNA fragments were visualized by gel electrophoresis and purified by gel extraction using the MinElute Gel Extraction Kit (QIAGEN). Digoxigenin-labeled cRNA was synthesized by *in vitro* transcription using the DIG RNA Labeling Kit (SP6/T7) according to the manufacturer's instructions (Roche).

**Tissue preparation.** Lumbar DRGs (L1–L6) were harvested, fixed in 4% PFA (Fluka/Sigma) for 3–4 h at room temperature, washed three times in 1 $\times$  PBS for 10–30 min, and cryoprotected in 25% sucrose (Merck)/PBS

overnight at 4°C. Tissue samples were embedded in Tissue-Tek OCT Compound (Sakura), snap-frozen in liquid nitrogen, and kept at  $-80^\circ\text{C}$  until use.

Cryostat sections (12  $\mu$ m) were mounted on Super Frost Plus slides (Thermo Scientific), air-dried for 1–3 h at room temperature, fixed with 4% PFA for 30 min, and prehybridized with equilibration buffer containing the following: 12.6 mM Tris (Roth) at pH 7.5, 185 mM NaCl (Sigma), 10 mM NaH<sub>2</sub>PO<sub>4</sub> (Merck), 5 mM EDTA (Sigma), 50% deionized formamide (Sigma), 0.5 $\times$  Denhardt's (Sigma) in DEPC-treated H<sub>2</sub>O for 5–15 min at room temperature. Hybridization was performed overnight at 68°C in a humidified chamber. cRNA probes were diluted 1:50 in hybridization buffer containing 12.6 mM Tris (Roth), pH 7.5, 185 mM NaCl (Sigma), 10 mM NaH<sub>2</sub>PO<sub>4</sub> (Merck), 5 mM EDTA (Sigma), 50% deionized formamide (Sigma), 10% dextrane sulfate (Sigma), 1 mg/ml yeast RNA (Roche), and 0.5 $\times$  Denhardt's (Sigma) in DEPC-treated H<sub>2</sub>O. Sections were then washed two times for 30 min at 68°C in 1 $\times$  SSC (Sigma), pH 7.0, 25% formamide, 0.1% Tween 20 (Sigma), twice for 30 min and once for 1 h at room temperature in 100 mM maleic acid (Sigma), 150 mM NaCl (Sigma), and 0.1% Tween 20 (Sigma), pH 7.5 (MABT). Sections were blocked for 1 h at room temperature with 20% FBS (Biochrom) in MABT before overnight incubation with alkaline phosphatase-conjugated anti-digoxigenin antibody (Roche; 1:3000) diluted in 20% FBS/MABT. Sections were washed 3 $\times$  20 min in MABT at room temperature and twice 10 min with alkaline phosphatase buffer containing 100 mM Tris, pH 9.5, 50 mM MgCl<sub>2</sub> (Merck), 100 mM NaCl, 0.1% Tween 20, and 0.024% levamisole (Fluka/Sigma). Revelation was performed using NBT/BCIP (Roche) in alkaline phosphatase buffer for 22–26 h at room temperature in the dark. The reaction was stopped by 2 washes with 5 mM EDTA/PBS, before postfixation with 4% PFA for 30 min at room temperature and subsequent embedding in Mowiol (Calbiochem).

For FISH, the same protocol was applied for hybridization and subsequent washing and blocking steps. Sections were incubated with alkaline phosphatase-conjugated anti-digoxigenin antibody (Roche; 1:1000), and revelation was performed using 2-hydroxy-3-naphthoic acid-2'-phenylamide phosphate (HNPP)/Fast Red (Roche). Briefly, sections were incubated with a freshly prepared solution of 0.1 mg/ml HNPP and 0.25 mg/ml Fast Red TR for 30 min at room temperature and then washed in alkaline phosphatase buffer (without levamisole). This procedure was repeated three times. Afterward, sections were washed once in PBS and stained with isolectin B4 (IB4)-Alexa-488 (1:200, Invitrogen) in PBS for 30 min at room temperature before embedding in Mowiol. For comparisons between genotypes, sections from equal numbers of animals per genotype (*gp130<sup>fl/fl</sup>* or *SNS-gp130<sup>-/-</sup>*) were placed on one slide and therefore subject to the same treatment. Sections of four to six DRG were used per animal.

Pictures were taken through an Axiovert 200M (Zeiss) with a Neofluar 40 $\times$  oil-immersion objective. NBT/BCIP-stained sections were illuminated using the bright field modus with a constant light intensity and 50 ms exposure time. Staining intensity was measured as mean gray value per neuron. Per section, gray values of one nonstained neuron were measured as background control. Because gray values correlated inversely with staining intensities, neurons with staining intensities exceeding background by 3.5 times the SD were considered as *Trpa1*-positive. Sections incubated with sense probe served as negative control and did not reveal any specific staining. HNPP/Fast Red TR staining was visualized using a matching filter set (BP 546/10, Omega; FT 560, BP 575–640, Zeiss) with 300 or 500 ms exposure time. All pictures were normalized to 500 ms exposure time and a threshold was applied to discriminate *Trpa1*-positive neurons. Sections hybridized with sense cRNA probes served as negative control and yielded  $< 4\%$  of positive neurons with the applied threshold. Approximately 300–1000 neurons per animal were analyzed, and data are displayed as means per animal. Quantification was performed using ImageJ (National Institutes of Health) (Schneider et al., 2012). All analyses were performed by an examiner who was blinded to genotype and sense/antisense staining.

**Microfluorimetric calcium measurements.** Twenty-four hour DRG neuron cultures were incubated with 6  $\mu$ M calcium ( $\text{Ca}^{2+}$ )-sensitive dye fura-2 AM (Invitrogen) in TNB medium (Biochrom) for 30 min at 37°C in 5% CO<sub>2</sub> in a humidified incubator. Then cells were washed and kept in



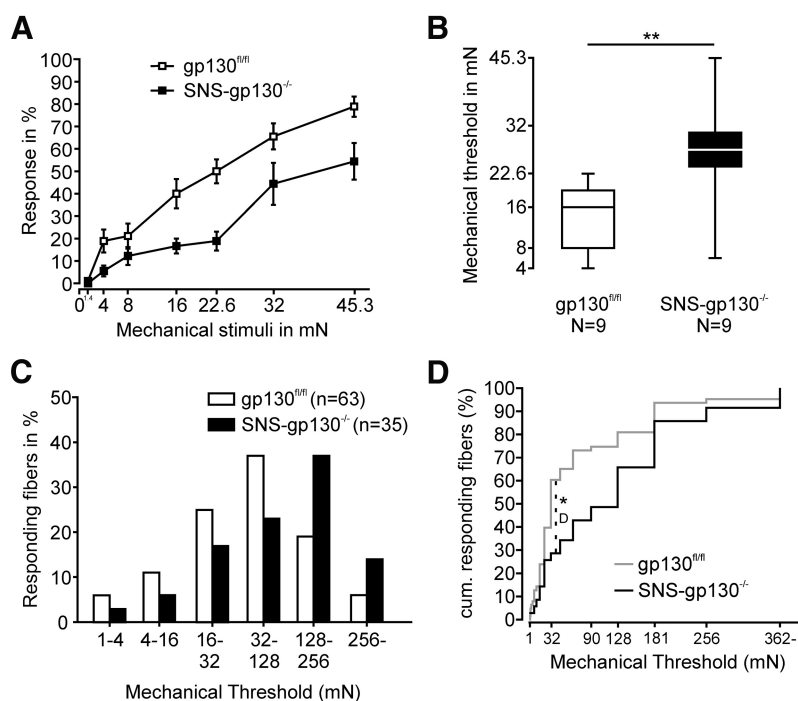
extracellular solution containing the following (in mM): 145 NaCl, 5 KCl, 2 CaCl<sub>2</sub>, 1 MgCl<sub>2</sub>, 10 D-glucose (all from Merck) and 10 HEPES (Roth), at pH 7.3 adjusted with NaOH (Merck). Ratiometric measurements were performed using a Zeiss Axiovert 200 microscope (Zeiss) with a Fluor 20×/0.75 NA objective (Zeiss). Fura-2 was excited consecutively at 340 and 380 nm (excitation time: 55 ms) with a polychrome IV monochromator (TILL Photonics), and fluorescence intensities were filtered by a 510 nm long pass filter and recorded with a CCD camera (Roper Scientific) at 1 s intervals. For data acquisition, MetaFluor 7.1.2.0 (Molecular Devices) was used, and off-line analysis was performed with Excel 2007 (Microsoft). Data traces were filtered by a simple 3-point moving average. All chemicals were diluted in extracellular solution and applied by a gravity-driven perfusion system (Dittert et al., 1998). Only cells with a low and stable baseline Ca<sup>2+</sup> signal within the first 120 s of the protocol were used for analysis. The inclusion criterion for cells responsive to CA or capsaicin was set to 125% of the baseline Ca<sup>2+</sup> signal 10 s before stimulation. The threshold for a positive response to 5-nitro-2-(3-phenylpropylamino) benzoic acid (NPPB) was defined by the highest Ca<sup>2+</sup> signal in *TRPA1<sup>-/-</sup>* DRG neurons during stimulation. Viability of the neurons was tested with a 10 s pulse of 25 mM potassium chloride solution in each experiment. All chemicals were purchased from Sigma.

**Statistical analysis.** Statistical analysis was performed per animal (*N*), and per nerve fiber or number of cells (*n*) for the single-nerve fiber recording and calcium imaging data. For statistical analysis, Sigma Stat 3 and Origin Pro 8 software were used, and the data are presented as mean ± SEM unless otherwise indicated. Depending on sample size, distribution, and number of variables, one-way ANOVA with post Tukey test, unpaired Student's *t* test, two-way ANOVA, or the nonparametric Mann–Whitney *U* test for comparison between groups, and  $\chi^2$  test with Yates correction as well as Kolmogorov–Smirnov test were used as stated in the respective text. Differences were considered statistically significant at *p* < 0.05.

## Results

### Reduced mechanonociception in *SNS-gp130<sup>-/-</sup>* mice

Because IL-6 sensitizes nociceptors to mechanical stimuli and its receptor gp130 is required for the maintenance of mechanical hypersensitivity in several rodent models of pathological pain (Brenn et al., 2007; Quarta et al., 2011), we hypothesized that gp130 may be an important switch for the function of mechanonociceptors. Therefore, we assessed the reflex withdrawal thresholds to mechanical stimuli of increasing strength of *gp130<sup>fl/fl</sup>* and *SNS-gp130<sup>-/-</sup>* mice with a calibrated set of von Frey filaments. Response frequencies of *SNS-gp130<sup>-/-</sup>* mice were significantly lower than those of *gp130<sup>fl/fl</sup>* mice (two-way ANOVA: genotype:  $F_{(1,125)} = 38.99051$ ; *p* < 0.00001, force:  $F_{(6,125)} = 40.08017$ ; *p* < 0.00001, genotype × force:  $F_{(6,125)} = 1.91561$ , *p* = 0.08432; *N* = 9; Fig. 1A), and the median mechanical withdrawal threshold was significantly higher for *SNS-gp130<sup>-/-</sup>* than for *gp130<sup>fl/fl</sup>* mice (*gp130<sup>fl/fl</sup>*: 16 mN with 8 mN and 19.3 mN as lower and upper quartile vs *SNS-gp130<sup>-/-</sup>*: 27.3 mN with 24 mN and 30.65 mN as lower and upper quartile; Mann–Whitney *U* test, *T* = 54, *p* = 0.00619; *N* = 9; Fig. 1B). Both parameters indicated

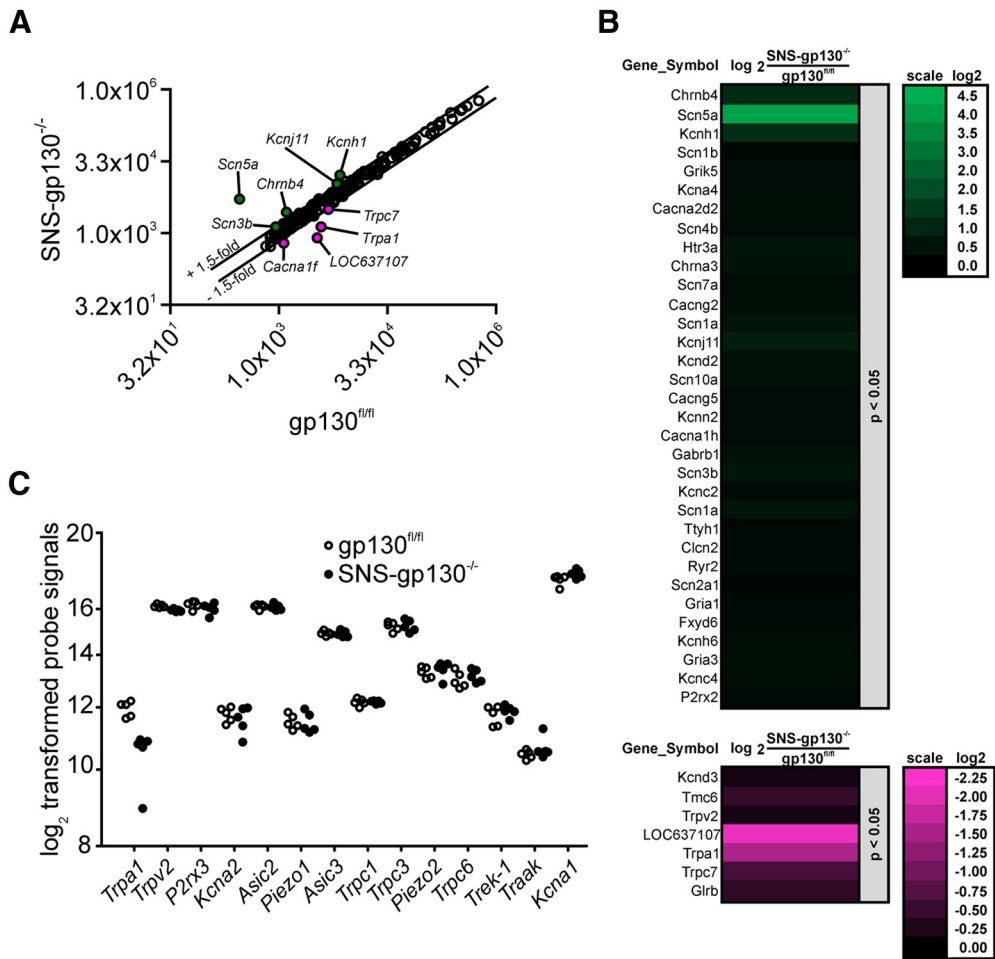


**Figure 1.** *SNS-gp130<sup>-/-</sup>* mice were less sensitive to mechanical stimulation. **A**, *SNS-gp130<sup>-/-</sup>* mice responded with significantly lower frequencies to mechanical stimulation with von Frey hairs than control *gp130<sup>fl/fl</sup>* mice (*N* = 9, two-way ANOVA, genotype: *p* < 0.001, forces: *p* < 0.001, genotype × forces: *p* = 0.084). **B**, Stimulus–response relationship of *SNS-gp130<sup>-/-</sup>* mice in the von Frey test was shifted toward stronger forces as *SNS-gp130<sup>-/-</sup>* mice exhibited significantly higher mechanical thresholds than *gp130<sup>fl/fl</sup>* mice. Boxes represent median as well as lower and upper quartile; whiskers represent the 10th and 90th percentiles. **C**, *In vitro*, the distribution of mechanical thresholds was significantly shifted toward stronger stimuli for unmyelinated mechanosensitive afferents obtained from *SNS-gp130<sup>-/-</sup>* (black columns) compared with those from *gp130<sup>fl/fl</sup>* mice (white columns; *gp130<sup>fl/fl</sup>*: median: 32 mN, *n* = 63, *N* = 30; black columns: *SNS-gp130<sup>-/-</sup>*: median: 128 mN, *n* = 35, *N* = 23; Mann–Whitney *U* test, *p* < 0.05). **D**, The cumulative distribution showed that a significantly lower percentage of fibers responded to medium to high mechanical forces in *SNS-gp130<sup>-/-</sup>* mice. \**p* < 0.05. \*\**p* < 0.01.

that *SNS-gp130<sup>-/-</sup>* mice were less sensitive to mechanical stimulation than control mice. Because the deletion of gp130 was restricted to Na<sub>v</sub>1.8-expressing unmyelinated sensory neurons, we analyzed the mechanical thresholds of C-fibers in *SNS-gp130<sup>-/-</sup>* mice using single-nerve fiber recordings in a skin-saphenous nerve *in vitro* preparation. Mechanical probing of the receptive field with von Frey filaments revealed that the mechanical activation thresholds of C-mechano- and polymodal nociceptors were shifted to stronger forces in *SNS-gp130<sup>-/-</sup>* mice (activation threshold: *gp130<sup>fl/fl</sup>*: median: 32 mN with 22.6 mN and 109 mN as lower and upper quartile vs *SNS-gp130<sup>-/-</sup>*: median: 128 mN with 27.3 mN and 181 mN lower and upper quartile, Mann–Whitney *U* test, *T* = 2065.5, *p* = 0.01369; Fig. 1C), suggesting that nociceptors were less sensitive to mechanical stimulation in *SNS-gp130<sup>-/-</sup>* mice compared with *gp130<sup>fl/fl</sup>* mice. In accordance with the behavioral experiments, the greatest differences were observed in the medium to high force range with the maximal accumulated difference at 32 mN (*gp130<sup>fl/fl</sup>* vs *SNS-gp130<sup>-/-</sup>*, Kolmogorov–Smirnov Test, *D*: 0.32381, *p* = 0.01219; Fig. 1D).

### Reduced expression of *Trpa1* mRNA in *SNS-gp130<sup>-/-</sup>* DRG neurons

gp130 signaling regulates mRNA expression in target cells classically via the JAK–STAT or via alternative pathways (Heinrich et al., 2003). A commercial genome-wide mRNA expression screen was used to compare expression levels of ion channels between *gp130<sup>fl/fl</sup>* and *SNS-gp130<sup>-/-</sup>* DRG explants. Out of the 177 ana-



**Figure 2.** Genome-wide mRNA expression profiling assay. **A**, Average ion channel expression intensities in *SNS-gp130<sup>-/-</sup>* compared with *gp130<sup>fl/fl</sup>* DRG explants ( $N = 5$ ). Depicted are the means of the quantile-normalized signals. The linear values are shown in a log<sub>2</sub> scaling. FDR-adjusted  $p$  values for deregulated genes: *Scn5a*,  $p = 0.162797384$ ; *Kcnh1*,  $p = 0.309818818$ ; *Chmb4*,  $p = 0.112616368$ ; *Kcnj11*,  $p = 0.36698117$ ; *Scn3b*,  $p = 0.396426446$ ; *Trpc7*,  $p = 0.378056322$ ; *Cacna1f*,  $p = 0.5246788$ ; *Trpa1*,  $p = 0.363034703$ ; *LOC637107*,  $p = 0.31986923$ . **B**, Heat map showing ion channel gene expression in *SNS-gp130<sup>-/-</sup>* versus *gp130<sup>fl/fl</sup>* DRG explants. Genes, which are upregulated (green, left) or downregulated (magenta, right), are sorted by their unadjusted  $p$  value. Only genes with  $p < 0.05$  are shown. **C**, Expressed ion channels with a putative function in mechanotransduction. Expression levels are shown as log<sub>2</sub> quantile-normalized signal values for each candidate and genotype (open circles represent *gp130<sup>fl/fl</sup>*; filled circles represent *SNS-gp130<sup>-/-</sup>*), and variation between individuals is minimal. Among the 14 genes, *Trpa1* is the only deregulated ion channel. For fold change in expression levels, see Table 1.

lyzed genes, 5 candidates were at least 1.5-fold upregulated, and 4 candidates demonstrated a 1.5-fold (or higher) downregulation in *SNS-gp130<sup>-/-</sup>* mice (Fig. 2A,B). Within the group of ion channels with a putative mechanosensitive function, the screen detected 14 of 17 possible targets, but only *Trpa1* mRNA expression levels were >1.5-fold lower in DRG explants of *SNS-gp130<sup>-/-</sup>* mice compared with *gp130<sup>fl/fl</sup>* mice (Fig. 2C; Table 1). Because the deregulation of *Trpa1* mRNA between *gp130<sup>fl/fl</sup>* and *SNS-gp130<sup>-/-</sup>* mice revealed a relatively high FDR-adjusted  $p$  value, we confirmed this result by quantifying *Trpa1* mRNA levels in thoracic and lumbar DRG explants of *gp130<sup>fl/fl</sup>* and *SNS-gp130<sup>-/-</sup>* mice with TaqMan qRT-PCR. Explants of *SNS-gp130<sup>-/-</sup>* mice expressed significantly less *Trpa1* mRNA than *gp130<sup>fl/fl</sup>* mice (*gp130<sup>fl/fl</sup>*:  $1.03 \pm 0.13$  vs *SNS-gp130<sup>-/-</sup>*:  $0.38 \pm 0.05$ ,  $N = 5-7$ ; Mann–Whitney  $U$  test,  $T = 50$ ,  $p = 0.00253$ ; Fig. 3A). Furthermore, we assessed the *Trpa1* mRNA expression levels in primary neuron cultures of *SNS-gp130<sup>-/-</sup>* and *gp130<sup>fl/fl</sup>* DRG. *Trpa1* expression levels were three times lower in neuron cultures of *SNS-gp130<sup>-/-</sup>* compared with *gp130<sup>fl/fl</sup>* mice (*gp130<sup>fl/fl</sup>*:  $1.00 \pm 0.04$  vs *SNS-gp130<sup>-/-</sup>*:  $0.32 \pm 0.06$ ,  $N = 3-5$ ; Mann–Whitney  $U$  test,  $T = 21$ ,  $p = 0.0357$ ; Fig. 3B). In contrast, expres-

**Table 1.** Genome-wide mRNA expression screen of mechanotransducer ion channel candidates

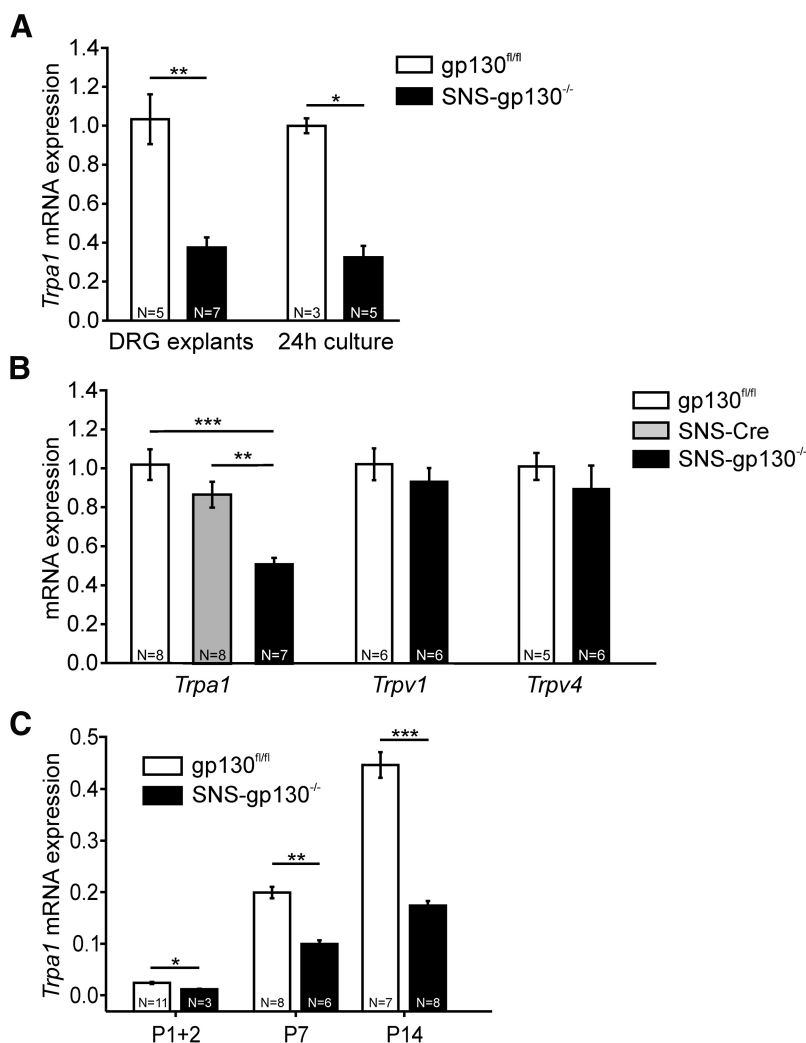
Gene symbol	Linearized fold change: <i>SNS-gp130<sup>-/-</sup></i> versus <i>gp130<sup>fl/fl</sup></i>	Net fold change >1.5
<i>Trpa1</i>	−2.8538	+
<i>Trpv2</i>	−1.1479	—
<i>P2rx3</i>	−1.1412	—
<i>Kcna2</i>	−1.1381	—
<i>Asic2</i>	−1.0220	—
<i>Piezo1</i>	−1.0216	—
<i>Asic3</i>	−1.0137	—
<i>Trpc1</i>	1.0033	—
<i>Trpc3</i>	1.0173	—
<i>Piezo2</i>	1.0765	—
<i>Trpc6</i>	1.1097	—
<i>Trek-1</i>	1.1177	—
<i>Traak</i>	1.1525	—
<i>Kcna1</i>	1.2410	—
<i>Asic1</i>	Not detectable	—
<i>Kcnq4</i>	Not detectable	—
<i>Trpv4</i>	Not detectable	—

sion levels of *Trpv1* and *Trpv4*, two members of the TRP superfamily of ion channels expressed in nociceptive neurons, were unaltered in DRG explants of *SNS-gp130<sup>-/-</sup>* mice (Fig. 3B). This suggests that gp130 deletion specifically targets *Trpa1* expression, whereas expression of other ion channels of the TRP family, which substantially contribute to hypernociception in mouse models of pathological pain, is not regulated by gp130 (for review, see Dubin and Patapoutian, 2010). Moreover, *OSM* knock-out mice reveal a reduced response in the formalin test, which largely depends on TRPA1 (Morikawa et al., 2004; McNamara et al., 2007). Last, to rule out an unspecific effect of the Cre-recombinase on *Trpa1* mRNA expression, we assessed *Trpa1* mRNA expression levels in lumbar DRG explants of *SNS-Cre* mice. *Trpa1* expression was similar for *gp130<sup>fl/fl</sup>* and *SNS-Cre* mice, in contrast to *SNS-gp130<sup>-/-</sup>* mice, which expressed significantly less *Trpa1* mRNA than both control strains (*gp130<sup>fl/fl</sup>*:  $1.02 \pm 0.08$  vs *SNS-Cre*:  $0.87 \pm 0.07$  vs *SNS-gp130<sup>-/-</sup>*:  $0.51 \pm 0.03$ ,  $N = 7$  or  $8$ ; one-way ANOVA  $F_{(2,22)} = 15.96582$ ,  $p = 0.00007$ ; Tukey post test *gp130<sup>fl/fl</sup>* vs *SNS-Cre*  $p = 0.21981$ , *gp130<sup>fl/fl</sup>* vs *SNS-gp130<sup>-/-</sup>*  $p = 0.00019$ , *SNS-Cre* vs *SNS-gp130<sup>-/-</sup>*  $p = 0.00269$ ; Fig. 3B).

Functional recombination and deletion of gp130 in Na<sub>v</sub>1.8-Cre mice start at the day of birth (P0), nearly coinciding with the occurrence of *Trpa1* expression (Agarwal et al., 2004; Hjerling-Leffler et al., 2007). We therefore assessed whether the deficit in *Trpa1* expression was already present during this early postnatal period. In DRG of *SNS-gp130<sup>-/-</sup>* and control mice, *Trpa1* mRNA expression started to be detectable within the first postnatal week. In control mice, *Trpa1* mRNA expression increased to 42.9% of the adult expression, at up to postnatal day P14. However, in *SNS-gp130<sup>-/-</sup>* DRG, *Trpa1* mRNA was low from P0 and remained significantly reduced at postnatal days P7 and P14 (*gp130<sup>fl/fl</sup>* vs *SNS-gp130<sup>-/-</sup>*: P1 + 2:  $0.023 \pm 0.002$  vs  $0.011 \pm 0.000$ ,  $T = 6$ ,  $p = 0.01273$ ; P7:  $0.191 \pm 0.011$  vs  $0.095 \pm 0.007$ ,  $T = 21$ ,  $p = 0.00179$ ; P14:  $0.429 \pm 0.024$  vs  $0.167 \pm 0.009$ ,  $T = 84$ ,  $p = 0.00031$ , all Mann–Whitney  $U$  test; Fig. 3C). These results indeed suggest that gp130 acts as a switch for *Trpa1* expression and the postnatal development of the nociceptive system.

#### Deregulation of *Trpa1* mRNA expression in nonpeptidergic neurons of *SNS-gp130<sup>-/-</sup>* mice

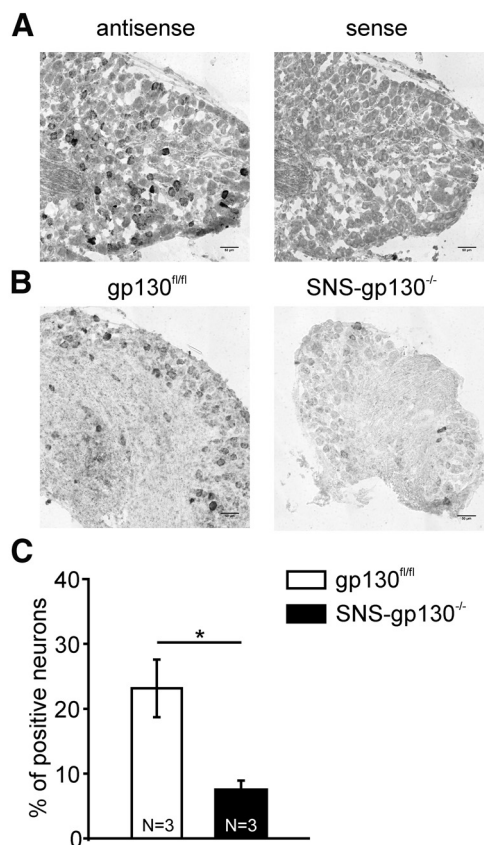
DRG contain all populations of sensory neurons, which can be defined by their function (e.g., as mechanonociceptors of the C-fiber class) and by certain immunohistochemical markers that are associated with certain neuron types. Even nociceptive primary afferents represent heterogeneous neuronal subpopulations. These can be primarily divided into peptidergic neurons,



**Figure 3.** Neuronal gp130 was essential for *Trpa1* mRNA expression in DRG. **A**, TaqMan mRNA analysis revealed significantly lower *Trpa1* mRNA levels in DRG explants and primary neuronal cultures of *SNS-gp130<sup>-/-</sup>* mice (DRG from thoracic and lumbar levels). **B**, *Trpa1* mRNA expression was significantly reduced in lumbar DRG explants from *SNS-gp130<sup>-/-</sup>* mice compared with *gp130<sup>fl/fl</sup>* and *SNS-Cre*, which were not significantly different. Furthermore, *Trpv1* and *Trpv4* mRNA levels were similar between *gp130<sup>fl/fl</sup>* and *SNS-gp130<sup>-/-</sup>* mice. **C**, *Trpa1* mRNA expression of lumbar DRG explants within the first 14 postnatal days. In *SNS-gp130<sup>-/-</sup>* mice, *Trpa1* mRNA expression was already reduced at P1–P14. Expression levels are normalized to adult *gp130<sup>fl/fl</sup>* mice. \* $p < 0.05$ . \*\* $p < 0.01$ . \*\*\* $p < 0.001$ .

expressing the neuropeptides substance P or calcitonin gene-related peptide, and nonpeptidergic neurons, which bind IB4. At the same time, the differential expression of specific receptor molecules determines the function of the nociceptor subpopulations (for review, see Dubin and Patapoutian, 2010). Therefore, it was important to identify the DRG subpopulation that requires gp130 for *Trpa1* expression. We used nonradioactive ISH to assess and localize *Trpa1* mRNA expression in subpopulations of lumbar DRG (L1–L6) of *SNS-gp130<sup>-/-</sup>* and control mice. First, chromogenic ISH with NBT/BCIP was used to determine riboprobe specificity and general *Trpa1* mRNA expression. In contrast to the sense probe, the *Trpa1* antisense probe produced a specific staining in the DRG neurons (Fig. 4A). In *gp130<sup>fl/fl</sup>* sections,  $23.1 \pm 4.4\%$  of the neurons stained positively for *Trpa1* mRNA, whereas *SNS-gp130<sup>-/-</sup>* mice showed a significantly lower percentage of *Trpa1*-positive neurons ( $7.5 \pm 1.4\%$ ;  $N = 3$ , 4–6 DRG per animal, sections/DRG; unpaired  $t$  test,  $t = 3.3577706$ ,  $p = 0.02836$ ; Fig. 4B,C). These results support the





**Figure 4.** Reduced *Trpa1* mRNA levels in DRG sections of *SNS-gp130<sup>-/-</sup>* mice. **A**, ISH with *Trpa1* antisense riboprobe produced a selective staining of *Trpa1*-expressing neurons, whereas sense probes did not show specific staining in the lumbar DRG sections. Scale bar, 50  $\mu$ m. **B**, Samples of *gp130<sup>fl/fl</sup>* and *SNS-gp130<sup>-/-</sup>* lumbar DRG sections with *Trpa1* antisense riboprobe display reduced *Trpa1* mRNA labeling in *SNS-gp130<sup>-/-</sup>* sections. Scale bar, 50  $\mu$ m. **C**, Quantification of the ISH labeling in lumbar *gp130<sup>fl/fl</sup>* and *SNS-gp130<sup>-/-</sup>* sections. *SNS-gp130<sup>-/-</sup>* sections reveal significantly lower numbers of *Trpa1* expressing neurons. \* $p < 0.05$ .

TaqMan qPCR data showing lower *Trpa1* mRNA expression levels in *SNS-gp130<sup>-/-</sup>* DRG explants compared with *gp130<sup>fl/fl</sup>* mice.

As mentioned above, nociceptive neurons can generally be classified into peptidergic and nonpeptidergic neurons. Although previous studies report contrasting results about *Trpa1* mRNA expression in nonpeptidergic neurons, a recent study by Barabas et al. (2012) suggests that TRPA1 function is primarily associated with IB4-positive DRG neurons (Story et al., 2003; Bautista et al., 2005; Obata et al., 2005; Caspani et al., 2009; Kim et al., 2010). To assess the differential *Trpa1* expression in nonpeptidergic versus all DRG neurons, DRG sections were colabeled with AlexaFluor-488-conjugated IB4. Because the dark blue NBT/BCIP precipitate can quench fluorescent dyes, HNPP/ FastRed TR was used as an alternative substrate for the alkaline phosphatase for these experiments. The percentage of IB4-positive neurons was unchanged between *gp130<sup>fl/fl</sup>* and *SNS-gp130<sup>-/-</sup>* mice (*gp130<sup>fl/fl</sup>*:  $34.2 \pm 3.3\%$  vs *SNS-gp130<sup>-/-</sup>*:  $33.6 \pm 2.1\%$ , 4–6 sections per animal,  $N = 5$ ; Fig. 5A–C), which is in accordance with our previous study (Andratsch et al., 2009). In *gp130<sup>fl/fl</sup>* lumbar DRG sections,  $18.9 \pm 3.6\%$  of the neurons were *Trpa1*-positive, whereas *SNS-gp130<sup>-/-</sup>* mice revealed a significantly lower percentage of *Trpa1*-positive neurons ( $8.3 \pm 1.7\%$ ; 4–6 sections per animal,  $N = 5$ ; Mann–Whitney  $U$  test,  $T = 39$ ,  $p = 0.01587$ ; Fig. 5D–F). A total of 40% of the IB4-positive neurons expressed *Trpa1* mRNA in DRG sections from *gp130<sup>fl/fl</sup>* mice. In *SNS-gp130<sup>-/-</sup>* mice, this percentage

was threefold lower (*gp130<sup>fl/fl</sup>*:  $40.0 \pm 6.0\%$  vs *SNS-gp130<sup>-/-</sup>*:  $13.1 \pm 3.2\%$ ; Mann–Whitney  $U$  test,  $T = 40$ ,  $p = 0.00794$ ; Fig. 5G–I). On the other hand, only 5%–9% of IB4-negative DRG neurons expressed *Trpa1*, and no significant difference in this neuron population was detected between the two genotypes (*gp130<sup>fl/fl</sup>*:  $9.0 \pm 3.2\%$  vs *SNS-gp130<sup>-/-</sup>*:  $4.8 \pm 1.4\%$ ; Mann–Whitney  $U$  test,  $T = 34$ ,  $p = 0.22222$ ). Together, these findings support the hypothesis that the presence of gp130 is essential for *Trpa1* expression in IB4-positive nociceptive neurons.

#### Regulation of *Trpa1* mRNA expression via JAK-STAT signaling

gp130-dependent cytokines signal via activation of the JAK-STAT and MAP kinase pathway (Heinrich et al., 2003; Pellegrino and Habecker, 2013). In primary cultures of DRG neurons, both the JAK inhibitor I and the STAT inhibitor (Stattic) significantly decreased *Trpa1* mRNA expression levels (JAK inhibitor I group: control:  $1.03902 \pm 0.1421$  vs inhibitor:  $0.33592 \pm 0.03694$ ,  $T = 57$ ,  $p = 0.00216$ ; Stattic group: control:  $1.01725 \pm 0.08066$  vs inhibitor:  $0.63475 \pm 0.08534$ ,  $T = 54$ ,  $p = 0.01515$ ;  $N = 6$ ; all Mann–Whitney  $U$  test; Fig. 6). Expression levels of the house-keeping genes *Sdh*a, *Gapdh*, and *18S* were similar in Stattic-treated and control samples ( $N = 2$ ; data not shown). Our data indicate that *Trpa1* mRNA expression is regulated by JAK-STAT signaling pathways. Together with the reduced *Trpa1* mRNA expression in *SNS-gp130<sup>-/-</sup>* DRG neurons, this suggests that intact gp130-JAK-STAT signal transduction is critically important for *Trpa1* mRNA transcription.

#### Reduced responsiveness to TRPA1 agonists in *SNS-gp130<sup>-/-</sup>* cultured sensory neurons

TRPA1 is a calcium-permeable ion channel that is activated by numerous pungent substances. Mustard oil and CA are the most commonly used activators of TRPA1 (Bandell et al., 2004; Jordt et al., 2004; Macpherson et al., 2007). In *gp130<sup>fl/fl</sup>* cultures, 41.9% (52 of 124,  $N = 3$ ) of the neurons responded to a 90 s pulse of 200  $\mu$ M CA with a calcium transient, whereas a significantly smaller proportion of neurons was activated by CA in *SNS-gp130<sup>-/-</sup>* cultures (11.6%, 14 of 120,  $N = 3$ ;  $\chi^2 = 26.80203$  with 1 degree of freedom:  $p = 0.0000002$ ; Fig. 7A,B). At the same time, responsiveness to 0.3  $\mu$ M of the TRPV1 agonist capsaicin was similar in *gp130<sup>fl/fl</sup>* and *SNS-gp130<sup>-/-</sup>* cultures (*gp130<sup>fl/fl</sup>*: 43 of 124 neurons = 34.6% vs *SNS-gp130<sup>-/-</sup>*: 39 of 110 neurons = 35.5%,  $N = 3$ ; Fig. 7A,B). This provides functional and translational evidence supporting the differences in *Trpa1* mRNA expression profiles reported above.

Apart from CA, the chloride channel blocker NPPB has been reported to selectively activate heterologously expressed TRPA1. However, reports on sensory neurons are as yet not available for this compound (Liu et al., 2010). NPPB caused a transient increase in intracellular  $\text{Ca}^{2+}$  concentration in cultured DRG neurons obtained from control (*TRPA1<sup>+/+</sup>*) mice, but responses to NPPB were virtually absent in neurons obtained from *TRPA1<sup>-/-</sup>* mice (median ratio increase: *TRPA1<sup>-/-</sup>*: 0.12169 with 0.08273 and 0.16727 as lower and upper quartile,  $n = 101$ ,  $N = 3$ ; *TRPA1<sup>+/+</sup>*: 0.23691 with 0.09383 and 0.7028 as lower and upper quartile,  $n = 53$ ,  $N = 2$ ; Fig. 7C). This suggests that the calcium transient observed was in response to NPPB exposure and was indeed due to its agonistic action on TRPA1. Using NPPB as a second TRPA1 agonist, we obtained a reduction of TRPA1 responsiveness in *SNS-gp130<sup>-/-</sup>* DRG cultures, with a similar degree to the one observed with CA. In *gp130<sup>fl/fl</sup>* cultures, 26.3% (31 of 118,  $N = 3$ ) of the neurons responded to NPPB with a mean



suprathreshold ratio increase of  $1.52 \pm 0.12$  (Fig. 7D). However, only 10.8% (18 of 167,  $N = 4$ ) of the *SNS-gp130<sup>-/-</sup>* neurons responded with a mean TRPA1-mediated ratio increase of  $1.11 \pm 0.11$  to the 60 s 100  $\mu\text{M}$  NPPB pulse. The proportion and the magnitude of the TRPA1-mediated ratio increase were significantly lower than in control cultures (proportion:  $\chi^2 = 10.59433$  with 1 degree of freedom:  $p = 0.00113$ ; ratio increase: unpaired  $t$  test:  $t = 2.33996$  with 47 degrees of freedom:  $p = 0.02358$ ; Fig. 7D). Therefore, we conclude from these data, with two different agonists, that the functional expression of TRPA1 is reduced in primary DRG cultures of *SNS-gp130<sup>-/-</sup>* mice.

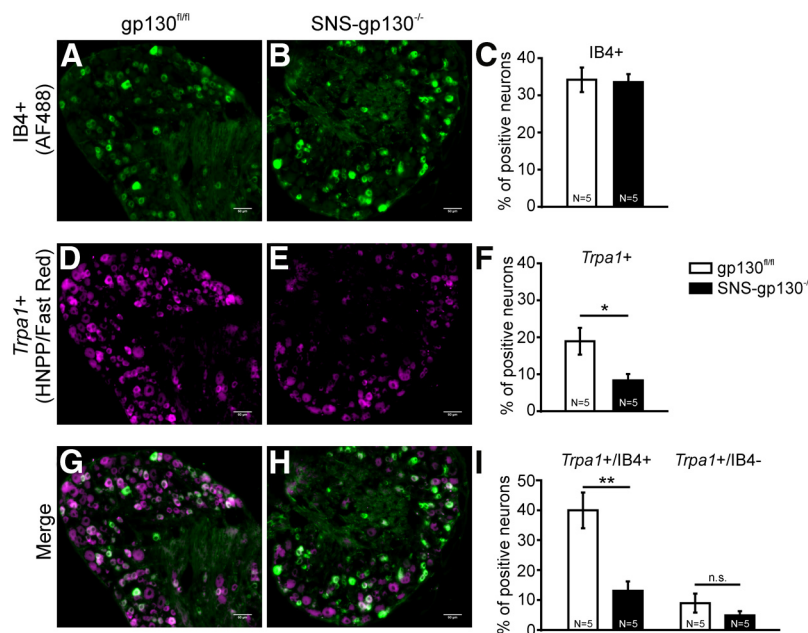
### Attenuated nocifensive response to the TRPA1 agonist CA in *SNS-gp130<sup>-/-</sup>* mice in vivo

Finally, we assessed whether the downregulation of TRPA1 expression in transgenic gp130 mice was functionally relevant *in vivo*. Injection of CA into the hindpaw of control mice induced spontaneous nocifensive behavior as reflected by a mean licking duration of  $25.9 \pm 8.4$  s and mean number of licking events of  $7.3 \pm 1.6$ . Control vehicle injection did not evoke nocifensive reactions during the 5 min observation period (mean licking duration  $< 1$  s; mean number of licking events  $< 1$ ; data not shown). *SNS-gp130<sup>-/-</sup>* mice revealed a significantly attenuated response to the TRPA1 agonist (licking duration:  $7.0 \pm 4.0$  s,  $T = 75$ ,  $p = 0.02890$ ; licking events:  $1.8 \pm 0.9$ ,  $T = 79$ ,  $p = 0.00591$ ; Mann–Whitney  $U$  test;  $N = 7$  or  $8$ ; Fig. 8). These data indicate that downregulation of TRPA1 in mice with a null mutation of gp130 in  $\text{Na}_v1.8$ -expressing nociceptors is relevant for responsiveness to noxious stimuli requiring TRPA1 *in vivo*. Overall, our data suggest that gp130 in nociceptive neurons is an essential switch for TRPA1 expression and absence of gp130 from nociceptive primary afferents causes severe deficits in acute mechanonociception and nocifensive reactions to TRPA1 agonists.

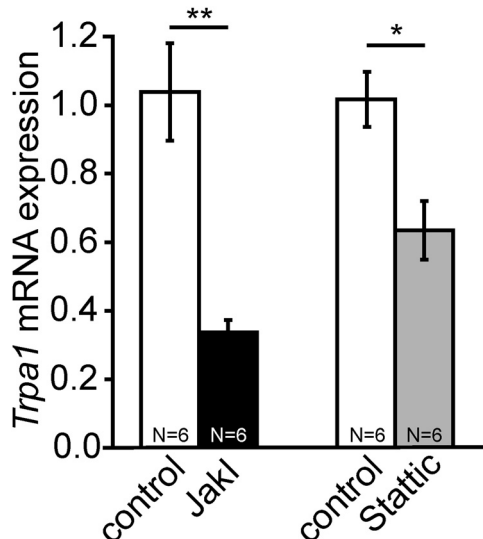
### Discussion

In the present study, we addressed the importance of neuronally expressed gp130 for mechanonociception, using a conditional transgenic gp130 knock-out mouse model previously generated in our group (Andratsch et al., 2009; Quarta et al., 2011). In behavioral tests and electrophysiological recordings, *SNS-gp130<sup>-/-</sup>* mice revealed reduced mechanonociception compared with control mice (Fig. 1). This deficit in mechanonociception was accompanied by low *Trpa1* mRNA expression levels, predominantly in nonpeptidergic neurons of *SNS-gp130<sup>-/-</sup>* mice from early postnatal development onwards (Figs. 234–5). Downstream of gp130, JAK and STAT were involved in the regulation of *Trpa1* transcription (Fig. 6), and the reduction in *Trpa1* expression in *SNS-gp130<sup>-/-</sup>* mice was associated with functional deficits *in vitro* and *in vivo* (Figs. 7 and 8).

Cytokines of the IL-6 family use a combination of ligand binding  $\alpha$ -receptors (IL-6R $\alpha$ , IL-11R $\alpha$ , and CNTFR $\alpha$ ) and the signal transducing receptor subunits gp130, LIFR, and OSMR. Com-

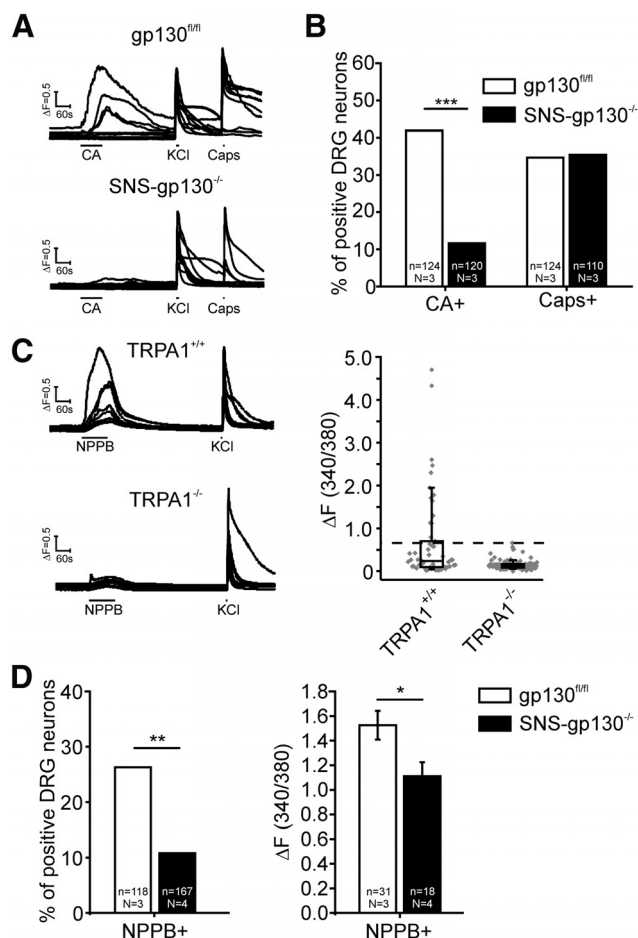


**Figure 5.** *Trpa1* deficiency was predominantly observed in IB4-binding nociceptive neurons. **A, B**, Staining for IB4+ neurons in *gp130<sup>fl/mi</sup>* and *SNS-gp130<sup>-/-</sup>* DRG sections. Scale bar, 50  $\mu\text{m}$ . **C**, The percentage of IB4-binding neurons was similar in both genotypes. **D, E**, FISH for *Trpa1* mRNA in *gp130<sup>fl/mi</sup>* and *SNS-gp130<sup>-/-</sup>* DRG sections. Scale bar, 50  $\mu\text{m}$ . **F**, Quantification of FISH shows that the number of *Trpa1* mRNA-expressing neurons (*Trpa1*+) was significantly reduced in *SNS-gp130<sup>-/-</sup>* mice. **G, H**, Colabeling of *Trpa1* mRNA expressing (magenta) and IB4-binding (green) neurons in both genotypes. Scale bar, 50  $\mu\text{m}$ . **I**, In control mice, *Trpa1* mRNA was predominantly expressed in IB4-positive neurons. In *SNS-gp130<sup>-/-</sup>* mice, reduction of *Trpa1* was most noticeable in the IB4-positive population. n.s., Not significant ( $p > 0.05$ ). \* $p < 0.05$ . \*\* $p < 0.01$ .

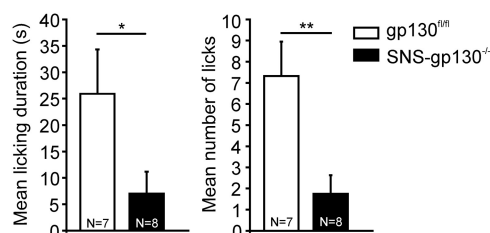


**Figure 6.** Expression of *Trpa1* mRNA was controlled by the JAK-STAT pathway. TaqMan mRNA analysis revealed significantly reduced *Trpa1* expression levels in DRG neurons incubated with a selective JAK inhibitor I (JAKI, 1  $\mu\text{M}$ ) or a STAT inhibitor, with selectivity for STAT3 (Stattic, 1.5  $\mu\text{M}$ ). \* $p < 0.05$ . \*\* $p < 0.01$ .

mon to all these receptor combinations is the essential requirement of at least one gp130 subunit for signal transduction. After activation, gp130 acts via classical JAK-STAT signaling and activation of the mitogen-activated protein kinase (MAPK) pathway (for review, see Heinrich et al., 2003). In addition to transcriptional regulation, IL-6 has been shown to enhance cap-dependent translation by mediating eIF4F cap complex formation, a process that allows control of translation in DRG neurons and is relevant for mechanical allodynia (Melemedjian et al.,



**Figure 7.** Functional TRPA1 expression was reduced in cultured primary DRG neurons of *SNS-gp130<sup>-/-</sup>* mice. **A**, Calcium imaging sample traces of primary neurons stimulated with the TRPA1-agonist CA (200  $\mu$ M), high potassium chloride (KCl, 25 mM) and TRPV1-agonist capsaicin (0.3  $\mu$ M). **B**, The percentage of neurons responding to CA was significantly decreased in *SNS-gp130<sup>-/-</sup>* cultures. In contrast, the responsiveness to capsaicin (Caps) was unchanged. **C**, Calcium imaging sample traces of primary neurons stimulated with NPPB (100  $\mu$ M) and KCl (25 mM). Responses to NPPB were greatly diminished in *TRPA1<sup>-/-</sup>* neurons. Magnitude of ratio increase in response to stimulation with 100  $\mu$ M NPPB in *TRPA1<sup>+/+</sup>* ( $n = 53$ ,  $N = 2$ ) and *TRPA1<sup>-/-</sup>* ( $n = 101$ ,  $N = 3$ ) neuronal cultures. The highest ratio increase in *TRPA1<sup>-/-</sup>* cultures was considered as threshold for TRPA1-mediated responses to NPPB. *TRPA1<sup>+/+</sup>* and *TRPA1<sup>-/-</sup>*: boxes represent median as well as lower and upper quartile; whiskers represent the 10th and 90th percentiles. **D**, Percentage and ratio increase of neurons responding to NPPB with a suprathreshold ratio increase in *gp130<sup>fl/m</sup>* and *SNS-gp130<sup>-/-</sup>* neuron cultures. In *SNS-gp130<sup>-/-</sup>* cultures, the number of neurons responding to NPPB was significantly decreased, and NPPB-responsive cells showed a significantly lower ratio increase to the stimulus. \* $p < 0.05$ . \*\* $p < 0.01$ . \*\*\* $p < 0.001$ .



**Figure 8.** Attenuated CA-induced nocifensive behavior in *SNS-gp130<sup>-/-</sup>* mice. *SNS-gp130<sup>-/-</sup>* mice displayed a significantly shorter licking duration and reduced number of licks of their hindpaws in response to an injection of 20  $\mu$ l 1% CA during the 5 min observation period. \* $p < 0.05$ . \*\* $p < 0.01$ .

2010). Here, we present strong evidence that regulation of TRPA1 by gp130 primarily takes place on the transcriptional level because (1) *Trpa1* mRNA expression was reduced in *SNS-gp130<sup>-/-</sup>* compared with control mice (Figs. 2 and 3); (2) the percentage of *Trpa1* mRNA-expressing neurons was lower in *SNS-gp130<sup>-/-</sup>* than in control mice (Figs. 4 and 5); and (3) *Trpa1* mRNA expression was significantly downregulated by experimental inhibition of the JAK-STAT pathway (Fig. 6).

Regulation of TRPA1 so far has been predominantly studied on the level of ion-channel modification (for review, see Nilius et al., 2012); however, only a few factors are known to influence *Trpa1* mRNA expression. The proinflammatory cytokines TNF- $\alpha$  and IL-1 activate the transcription factors NF- $\kappa$ B and HIF-1 $\alpha$ , which increase *Trpa1* mRNA expression (Hatano et al., 2012). Further support for a cytokine-based regulation of *Trpa1* comes from a transgenic IL-13 mouse model for atopic dermatitis, in which *Trpa1* expression is increased in mast cells, DRG neurons, and nerve fibers innervating the skin (Oh et al., 2013). In DRG neurons, previous studies mainly investigated differential *Trpa1* expression after inflammation (Obata et al., 2005; Schwartz et al., 2011; Merrill et al., 2012) and during late embryonic and postnatal development (Hjerling-Lefler et al., 2007; Abdel Samad et al., 2010; Gascon et al., 2010; Franck et al., 2011). After inflammation and spinal nerve ligation, NGF increases *Trpa1* mRNA expression in DRG neurons via p38 (Obata et al., 2005; Diogenes et al., 2007). During development, the signaling receptor for GDNF family ligand Ret is required for *Trpa1* expression and the development of nonpeptidergic DRG neurons (Luo et al., 2007; Abdel Samad et al., 2010; Franck et al., 2011). However, Ret also regulates *Trpa1* expression in adulthood, and *Trpa1* mRNA levels are increased by overexpression of the GDNF family ligand artemin, but not by injection of GDNF (Obata et al., 2005; Elitt et al., 2006; Franck et al., 2011).

Apart from their crucial role in inflammation and regeneration, cytokines of the IL-6 family are important survival and differentiation factors for neurons (Murphy et al., 1991, 1993; Thompson et al., 1996; Banner et al., 1998; Nakashima et al., 1999; Zhong et al., 1999). Our results indicate a new role for IL-6-related cytokines, in which gp130 determines the expression of TRPA1, most likely via the JAK-STAT pathway. Our model of gp130-dependent ion channel expression in nociceptive neurons is backed up by studies investigating the role of OSM in DRG neurons where OSM is essential for the development of a subpopulation of TRPV1/P2X3-expressing neurons (Morikawa et al., 2004). The OSM receptor  $\beta$  is predominantly expressed in nonpeptidergic neurons, and its expression starts around postnatal day 0 before it reaches adult expression levels at P14 (Tamura et al., 2003; Morikawa et al., 2004). *OSM<sup>-/-</sup>* mice do not only reveal attenuated reactions to the injection of TRPV1 and P2X3 agonists, but also to formalin injection and in the tail-clip test for mechanical sensitivity. Because formalin is a direct agonist for the mechanosensitive channel TRPA1 (McNamara et al., 2007), this test can also be interpreted as an indicator for a differential expression of TRPA1 in *OSM<sup>-/-</sup>* mice, which so far has not been investigated. In line with these results, we observed that gp130 was especially necessary for *Trpa1* expression in nonpeptidergic IB4-binding neurons (Fig. 5) and that the ablation of gp130 affected *Trpa1* expression already during early postnatal development (Fig. 3). In contrast to the study of Morikawa et al. (2004), we detected neither a change in *Trpv1* mRNA expression levels (Fig. 3) nor in the percentage of TRPV1-expressing neurons in *SNS-gp130<sup>-/-</sup>* mice (Fig. 7). This difference can easily be explained by the nature of the two mouse strains. In *SNS-gp130<sup>-/-</sup>*

mice, gp130 ablation is effective in 90% of the Na<sub>v</sub>1.8-expressing DRG neuron population (Andratsch et al., 2009). Hence, a residual population of nociceptive neurons still maintains gp130 signaling. Furthermore, functional TRPV1 expression emerges on embryonic day E12.5 and thus precedes Na<sub>v</sub>1.8 and Cre-recombinase expression, which becomes detectable around P0 and coincides with TRPA1 expression (Agarwal et al., 2004; Hjerling-Leffler et al., 2007). However, in *OSM*<sup>-/-</sup> mice with a global null mutation of OSM, it cannot be excluded that non-neuronal cell types contribute to the phenotype observed. More importantly, OSM effects can be conveyed via both OSMR-β-gp130 and LIFR-gp130 heteromeric receptor complexes. The latter can regulate specific populations of sensory neurons during embryonic development (Murphy et al., 1997), whereas TRPV1-expressing populations are very likely established before gp130 depletion in *SNS-gp130*<sup>-/-</sup> mice.

Although different mechanisms, including mechanically activated ion channels, have been proposed for force transduction in nociceptive neurons, the molecular mechanisms of mechanonociception are still largely enigmatic (Lumpkin and Caterina, 2007; Delmas et al., 2011; Delmas and Coste, 2013). We have previously shown that cultured *SNS-gp130*<sup>-/-</sup> DRG neurons demonstrate reduced excitability compared with control mice, which is based on increased delayed rectifier and A-type potassium currents with no observable changes in voltage-gated sodium and calcium currents. Moreover, this is accompanied by increased expression of A-type channel *Kcna4*, but not *Kcnh1* (Langeslag et al., 2014). Although *Kcna4* has not been associated with mechanonociception, reduced excitability of nociceptive fibers could lead to a hyponociceptive phenotype. However, our previous studies do not reveal reduced sensitivity of *SNS-gp130*<sup>-/-</sup> mice to heat stimulation under basal conditions (Andratsch et al., 2009; Langeslag et al., 2011). Therefore, the reduced excitability would have to be restricted to mechanosensitive C-fibers, a theory for which we do not have any indication. Alternatively to a reduced neuronal excitability, we present here with the decreased expression of TRPA1 a further possible mechanism for the deficit in mechanonociception of *SNS-gp130*<sup>-/-</sup> mice. Among several other candidate ion channels, TRPA1 has evolved as an important transducer for noxious mechanical stimuli. TRPA1 can be activated by mechanical stimulation of DRG neurons, and a null mutation of TRPA1 affects intermediate or slowly adapting mechanosensitive ionic currents in small-diameter DRG neurons (Vilceanu and Stucky, 2010; Brierley et al., 2011). Furthermore, this effect can be rescued by reintroduction of TRPA1 into DRG neurons of *TRPA1* knock-out mice (Brierley et al., 2011). Although controversial results have been published, the majority of studies show that deletion and pharmacological inhibition of TRPA1 reduce the responsiveness of nociceptors to mechanical forces in electrophysiological recordings of single-nerve fibers and in assays of mechanosensitivity *in vivo* (Bautista et al., 2006; Kwan et al., 2006, 2009; Petrus et al., 2007). The reduced mechanosensitivity of *SNS-gp130*<sup>-/-</sup> mice in the present study bears great resemblance to the previously published phenotype observed in *TRPA1*<sup>-/-</sup> mice. In particular, both transgenic mouse strains reveal deficits in the detection of strong mechanical forces (Kwan et al., 2006).

Together, we propose a model in which the presence of gp130 in small DRG neurons is critically important for mechanonociception and control of TRPA1 expression in nonpeptidergic neurons. Our data shed new light on the importance of IL-6-related cytokines for the development and functional properties of the nociceptive system. Despite conflicting results, numerous studies

indicate that pharmacological inhibition of TRPA1 attenuates mechanical hypersensitivity originating from inflammation, nerve injury, or chemotherapy-induced neuropathy (Obata et al., 2005; Bautista et al., 2006; Dai et al., 2007; Petrus et al., 2007; Eid et al., 2008; Brierley et al., 2011; Nassini et al., 2011). Neuropoietic IL-6 is a key mediator in innate immunity, inflammation, and mechanical hypersensitivity in different models for chronic pain (Brenn et al., 2007; Quarta et al., 2011). The present study stresses the importance of the IL-6 signal transducer gp130 in nociceptor responsiveness. Rather than gp130 causing direct sensitization, we suggest that it acts in nociceptors as general switch for the expression of TRPA1, as one critically important ion channel for mechanonociception.

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