

PUBLISHED VERSION

Antony Leonard, Adrienne W. Paton, Monaliza El-Quadi, James C. Paton, Fabeha Fazal
Preconditioning with endoplasmic reticulum stress ameliorates endothelial cell inflammation
PLoS One, 2014; 9(10): e110949-1-e110949-18

© 2014 Leonard et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

PERMISSIONS

<http://creativecommons.org/licenses/by/4.0/>



This is a human-readable summary of (and not a substitute for) the [license](#).

[Disclaimer](#)



You are free to:

Share — copy and redistribute the material in any medium or format

Adapt — remix, transform, and build upon the material

for any purpose, even commercially.

The licensor cannot revoke these freedoms as long as you follow the license terms.

Under the following terms:



Attribution — You must give [appropriate credit](#), provide a link to the license, and [indicate if changes were made](#). You may do so in any reasonable manner, but not in any way that suggests the licensor endorses you or your use.

No additional restrictions — You may not apply legal terms or [technological measures](#) that legally restrict others from doing anything the license permits.

<http://hdl.handle.net/2440/95048>



Preconditioning with Endoplasmic Reticulum Stress Ameliorates Endothelial Cell Inflammation

Antony Leonard¹, Adrienne W. Paton², Monaliza El-Quadi¹, James C. Paton², Fabeha Fazal^{1*}

1 Department of Pediatrics, Lung Biology and Disease Program, University of Rochester School of Medicine and Dentistry, Rochester, New York, United States of America, **2** Research Centre for Infectious Diseases, School of Molecular and Biomedical Science, University of Adelaide, Adelaide, South Australia, Australia

Abstract

Endoplasmic Reticulum (ER) stress, caused by disturbance in ER homeostasis, has been implicated in several pathological conditions such as ischemic injury, neurodegenerative disorders, metabolic diseases and more recently in inflammatory conditions. Our present study aims at understanding the role of ER stress in endothelial cell (EC) inflammation, a critical event in the pathogenesis of acute lung injury (ALI). We found that preconditioning human pulmonary artery endothelial cells (HPAEC) to ER stress either by depleting ER chaperone and signaling regulator BiP using siRNA, or specifically cleaving (inactivating) BiP using subtilase cytotoxin (SubAB), alleviates EC inflammation. The two approaches adopted to abrogate BiP function induced ATF4 protein expression and the phosphorylation of eIF2 α , both markers of ER stress, which in turn resulted in blunting the activation of NF- κ B, and restoring endothelial barrier integrity. Pretreatment of HPAEC with BiP siRNA inhibited thrombin-induced I κ B α degradation and its resulting downstream signaling pathway involving NF- κ B nuclear translocation, DNA binding, phosphorylation at serine536, transcriptional activation and subsequent expression of adhesion molecules. However, TNF α -mediated NF- κ B signaling was unaffected upon BiP knockdown. In an alternative approach, SubAB-mediated inactivation of NF- κ B was independent of I κ B α degradation. Mechanistic analysis revealed that pretreatment of EC with SubAB interfered with the binding of the liberated NF- κ B to the DNA, thereby resulting in reduced expression of adhesion molecules, cytokines and chemokines. In addition, both knockdown and inactivation of BiP stimulated actin cytoskeletal reorganization resulting in restoration of endothelial permeability. Together our studies indicate that BiP plays a central role in EC inflammation and injury via its action on NF- κ B activation and regulation of vascular permeability.

Citation: Leonard A, Paton AW, El-Quadi M, Paton JC, Fazal F (2014) Preconditioning with Endoplasmic Reticulum Stress Ameliorates Endothelial Cell Inflammation. PLoS ONE 9(10): e110949. doi:10.1371/journal.pone.0110949

Editor: Yulia Komarova, University of Illinois at Chicago, United States of America

Received: June 10, 2014; **Accepted:** September 17, 2014; **Published:** October 30, 2014

Copyright: © 2014 Leonard et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability: The authors confirm that all data underlying the findings are fully available without restriction. All relevant data are within the paper and its Supporting Information files.

Funding: This work was supported by National Heart, Lung and Blood Institute Grant HL096907. The funder had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* Email: Fabeha_Fazal@URMC.Rochester.edu

Introduction

Endoplasmic reticulum, an intricate cellular organelle present in eukaryotic cells, is a major site for the synthesis and maturation of secretory and membrane proteins [1–3]. Protein synthesis in ER is dynamically regulated as per the physiological need of the cell. However, a wide variety of disturbances such as glucose deprivation, changes in redox status, disruption of calcium homeostasis and viral and bacterial infections can cause imbalance in the luminal flux of the newly synthesized unfolded or misfolded peptides resulting in a condition known as ER stress. To combat ER stress, an evolutionarily conserved adaptive mechanism, termed the unfolded protein response (UPR) is activated and assists in cell survival. However, if the ER dysfunction is prolonged and severe, the UPR initiates cell death via apoptosis or autophagy. A key component involved in the regulation and activation of the UPR is the ER chaperone BiP (Binding Immunoglobulin Protein), a 78-kDa glucose-regulated protein (GRP78), also referred to as heat-shock protein A5 (HSPA5). In the unstressed state, BiP is found associated with the luminal domains of three ER stress sensors, protein kinase RNA-like ER

kinase (PERK), inositol-requiring enzyme (IRE)1- α/β and activating transcription factor (ATF)6- α/β . However, upon induction of ER stress, BiP dissociates from the ER signal sensors, causing their phosphorylation, activation and translocation. Together, these three branches of the UPR restore ER homeostasis [4,5]. ER stress is mechanistically linked to inflammation at several levels [6] as evidenced by the fact that it is an underlying factor in the pathogenesis of several metabolic and immunological diseases, with inflammatory underpinning, such as obesity, diabetes, inflammatory bowel disease (IBD) and glomerular disease [7–10].

Inflammation is the body's protective mechanism against infection or injury and it tends to resolve once the source has been cleared from the system. However, when the inflammatory response becomes severe or prolonged it results in a diseased state such as acute lung injury (ALI). This is characterized by massive infiltration of polymorphonuclear leukocytes (PMN) from the blood into the lung that leads to disruption of vascular endothelial permeability and development of pulmonary edema, with severe consequences for pulmonary gas exchange [11–17]. The movement of PMN from the blood to the inflammatory site involves the interaction between intercellular cell adhesion molecule-1 (ICAM-1) on EC surfaces and

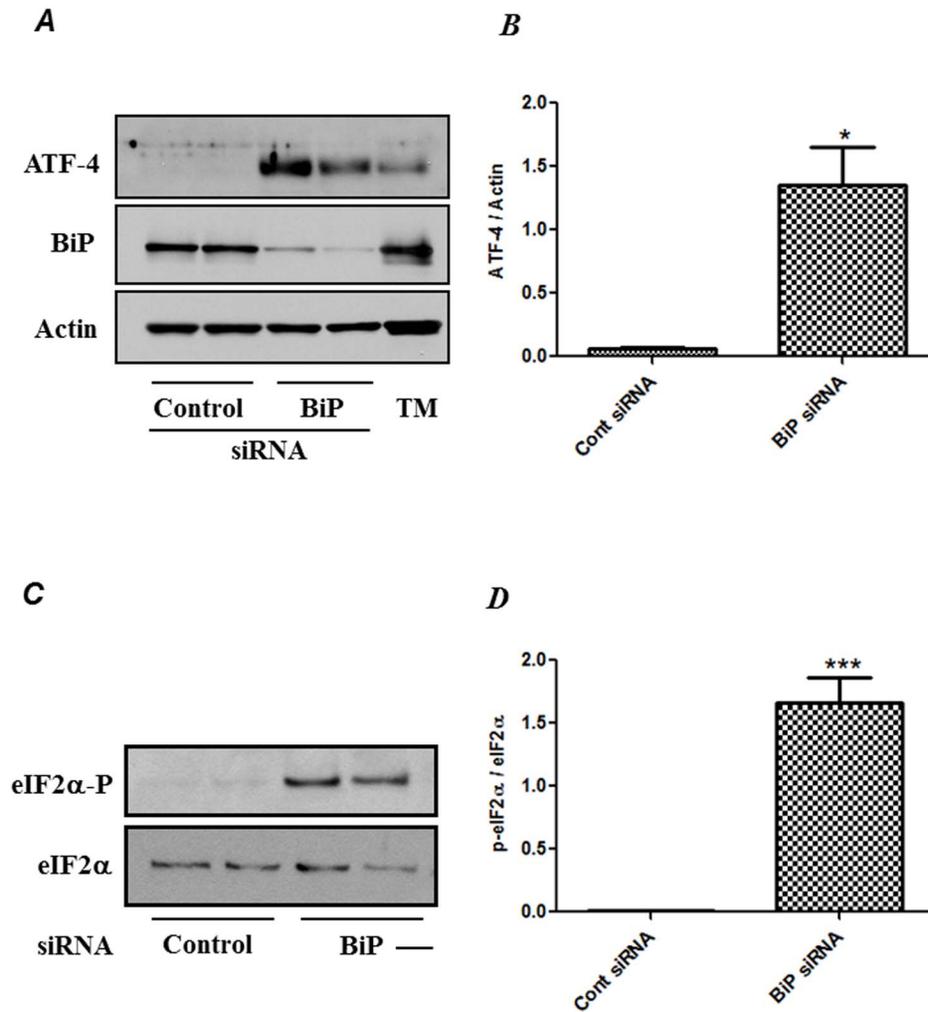


Figure 1. BiP knockdown induces ER stress. HPAEC were transfected with control siRNA or BiP siRNA for 48 h using DharmaFect1. Total cell lysates were separated by SDS-PAGE and immunoblotted with (A) anti-ATF4 antibody, and anti-actin antibody were used to monitor loading and anti-BiP antibody was used to monitor depletion and with (C) anti-phospho-eIF2 α and anti-eIF2 α antibody. TM, tunicamycin was used as a positive control. The data are the means \pm S.E. (n = 3–6 for each condition). The bar graph represents the effect of BiP depletion on (B) ATF-4 expression and (D) eIF2- α phosphorylation, normalized to actin level. The data are the means \pm S.E. (n = 3–6 for each condition). * p <0.01 or *** p <0.001 difference from controls.

doi:10.1371/journal.pone.0110949.g001

its counter receptor β 2 integrins (CD11/CD18) on the surface of PMNs (18). Chemokines Interleukin-8 (IL-8) and monocyte chemoattractant protein-1 (MCP-1) are immune mediators involved in targeting leukocytes and monocytes to sites of inflammation. The expression of ICAM-1, VCAM-1, IL-8 and MCP-1 is under the tight control of the inflammatory transcription factor NF- κ B [18,19]. NF- κ B is activated upon phosphorylation of its cytoplasmic inhibitor I κ B α on Serine³² and Serine³⁶ by I κ B kinase (IKK) complex. Phosphorylation triggers the ubiquitination-mediated degradation of I κ B α , which results in nuclear translocation of NF- κ B and subsequent transcription of inflammatory genes.

Interestingly, recent studies have also shown that ER stress regulates NF- κ B activity in a biphasic and bidirectional manner [7] in different cell types, contributing to the pathogenesis of diseases such as cancer, amyotrophic lateral sclerosis (ALS) and diabetic retinopathy [20–23]. In contrast, studies have also shown that ER stress preconditioning protects the cells against a number of inflammatory stimuli [24–30]. Deregulated NF κ B activity has

been implicated in a wide range of human diseases including cancer, ALI, diabetes, arthritis, and infection [31–35].

In the present study we adopted a dual approach to unravel the role of the ER stress regulatory protein BiP in EC inflammation and injury associated with ALI. Our data show that preconditioning the endothelial cells with ER stress by depleting BiP using siRNA, or by inactivating BiP using SubAB [36], mitigates inflammation by blunting the NF- κ B transcriptional activity and by restoring vascular endothelial permeability.

Materials and Methods

Reagents

Human thrombin was obtained from Enzyme Research Laboratories (South Bend, IN). Tunicamycin was purchased from Sigma. Polyclonal antibodies to ICAM-1, VCAM-1, RelA/p65, β -actin, I κ B α , and Lamin B were from Santa Cruz Biotechnology (Santa Cruz, CA). Antibodies to phospho-eIF2 α , eIF2 α , BiP/GRP78, ATF-4, phospho-AKT, phospho-IKK β , IKK β , phospho-

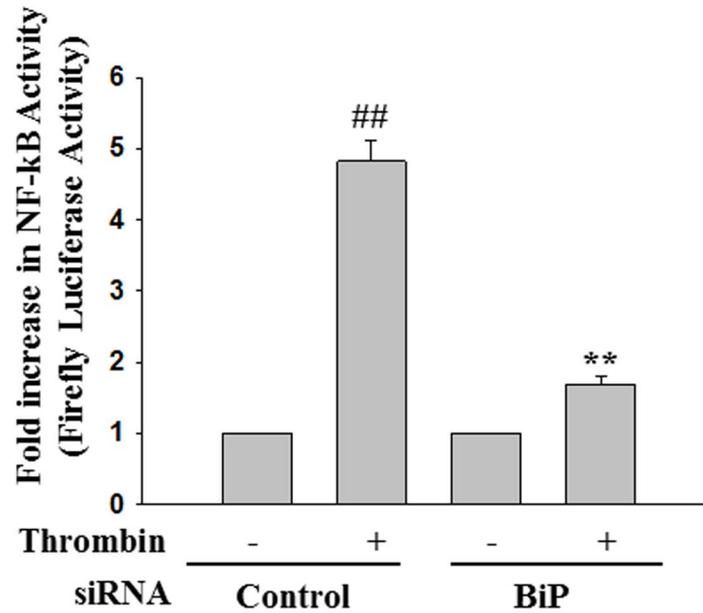
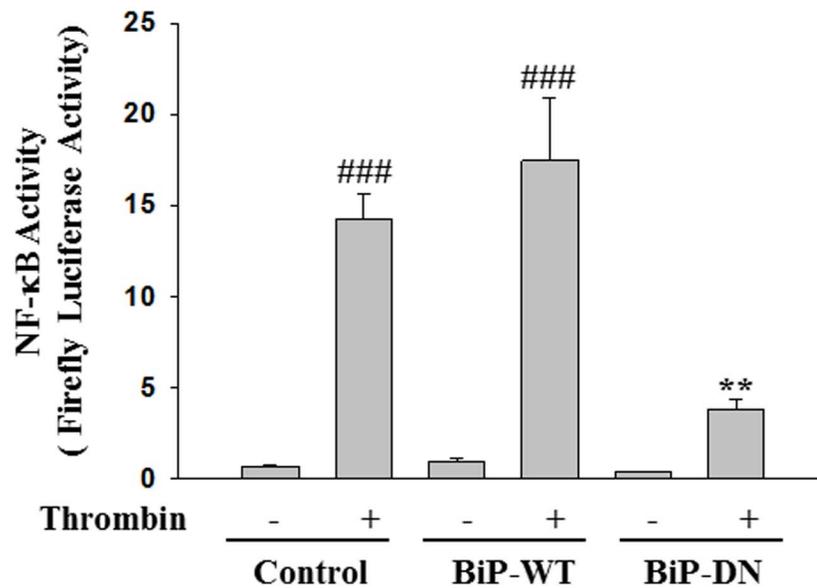
A**B**

Figure 2. BiP knockdown attenuates thrombin-induced NF-κB reporter activity. HPAEC were transfected with control siRNA or BiP siRNA by use of DharmaFect1. Twenty-four hours later, cells were again transfected with NF-κBLUC construct by using DEAE-dextran as described in Materials and Methods. Cells were then challenged with (A) thrombin (5 U/ml) for 6 h, and the cell extracts were prepared and assayed for firefly and Renilla luciferase activities. The data were expressed as a fold increase in firefly to Renilla luciferase activities. Data are means \pm SE (n=4–6 for each condition). ## p <0.01 difference from controls; ** p <0.01 difference from thrombin stimulated controls. (B) HPAEC were transfected with NF-κBLUC in combination with BiP WT (pCMV-BiP-Myc-KDEL-WT) or BiP dominant negative (pCMV-BiP-Myc-KDEL-T37G) as described in Materials and Methods. Sixteen hours after transfection cells were treated with thrombin (5 U/ml) for 6 hrs. Cell extracts were prepared and assayed for firefly and Renilla luciferase activities. The data were expressed as a ratio of firefly to Renilla luciferase activities. Data are means \pm SE (n=4–6 for each condition). ### p <0.001 difference from controls; ** p <0.01 difference from thrombin stimulated controls. doi:10.1371/journal.pone.0110949.g002

(Ser⁵³⁶)-RelA/p65, were obtained from Cell Signaling (Beverly, MA). Tata Binding Protein (TBP) antibody was purchased from Abcam. RelA/p65 transcription factor assay kit was purchased

from Cayman Chemical (Ann Arbor, MI) and plasmid maxi kit was from QIAGEN Inc. (Valencia, CA). Protein assay kit and nitrocellulose membrane were from Bio-Rad. Alexa Fluor

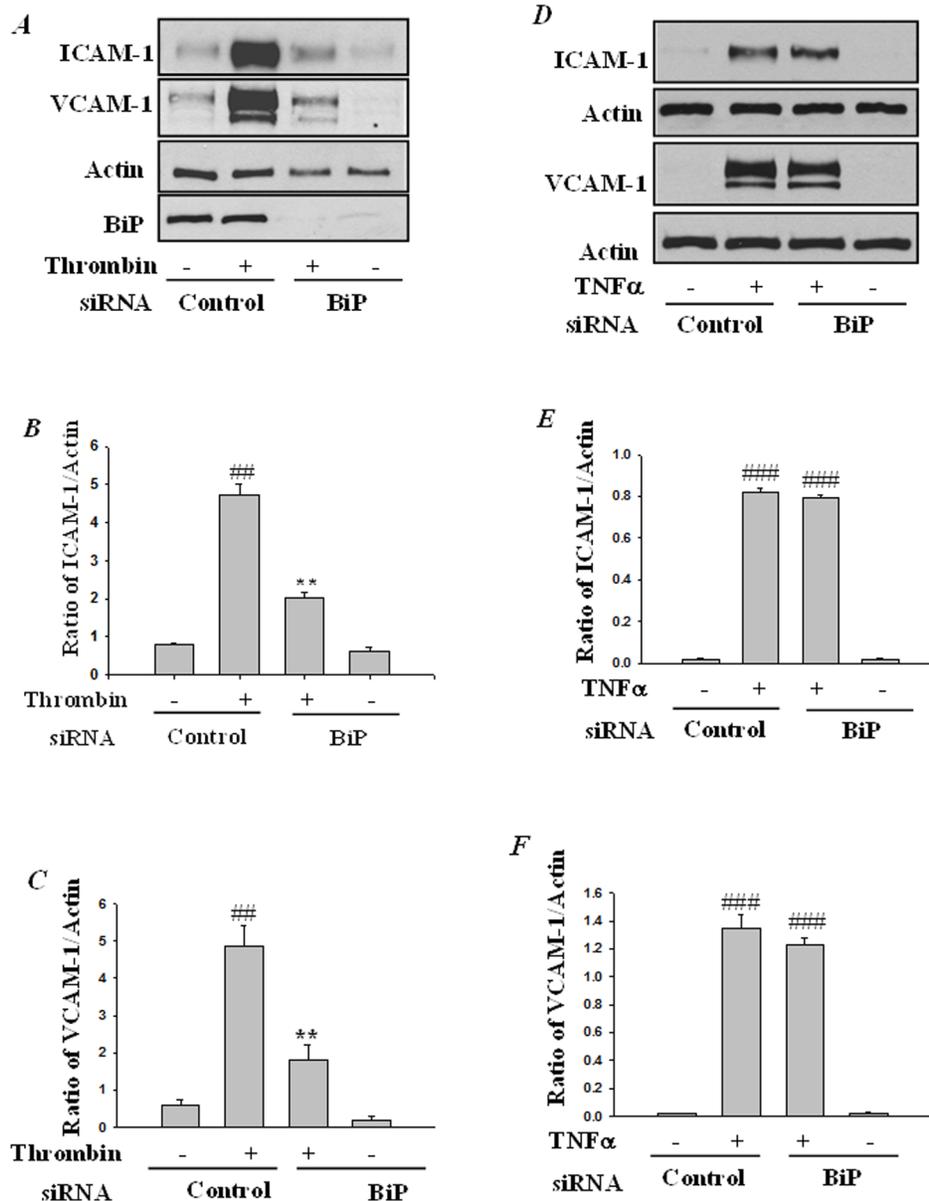


Figure 3. BiP knockdown mitigates thrombin-induced adhesion molecule expression. HPAEC were transfected with control siRNA or BiP-siRNA using DharmaFect1. After 24–36 h, the cells were challenged with (A) thrombin (5 U/ml) or (D) TNF α (100 U/ml), for 6 h. Total cell lysates were immunoblotted with an anti-ICAM-1, anti VCAM-1, and anti-BiP antibody. Actin was used to monitor loading. The bar graph represents the effect of BiP depletion on (B & C) thrombin and (E & F) TNF α -induced ICAM-1 and VCAM-1 expression normalized to actin level. The data are the means \pm S.E. (n=6 for each condition). ^{##} $p < 0.01$ or ^{###} $p < 0.001$ difference from controls; ^{**} $p < 0.01$ difference from thrombin stimulated controls. doi:10.1371/journal.pone.0110949.g003

488-phalloidin was purchased from Invitrogen. Expression vector encoding Wild type BiP and dominant negative BiP were from Addgene. All other materials were from VWR Scientific Products Corporation (Gaithersburg, MD) and Fisher Scientific (Pittsburgh, PA). SubAB and its non-toxic derivative SubA_{A272}B were purified as previously described [37].

Cell Culture

Human pulmonary artery endothelial cells (HPAEC) were purchased from Lonza (Walkersville, MD). Cells were cultured as described previously [38,39] in endothelial basal medium 2 (EBM2) supplemented with bullet kit additives (Lonza, Walkersville, MD) and were used between passages 3 and 7.

RNAi knockdown

SMARTpool siRNA specific for human BiP and a non-targeting siRNA control were purchased from Dharmacon (Lafayette, CO). EC were transfected with BiP siRNA or control siRNA using DharmaFect1 siRNA Transfection Reagent (Dharmacon) essentially as described [40].

NF- κ B Transcriptional Activity

The construct pNF- κ B-LUC containing five copies of consensus NF- κ B sequences linked to a minimal E1B promoter-luciferase gene was purchased from Stratagene (La Jolla, CA). Transfections were performed using the DEAE-dextran method essentially as described [41]. Briefly, 5 μ g of DNA was mixed with 50 μ g/ml

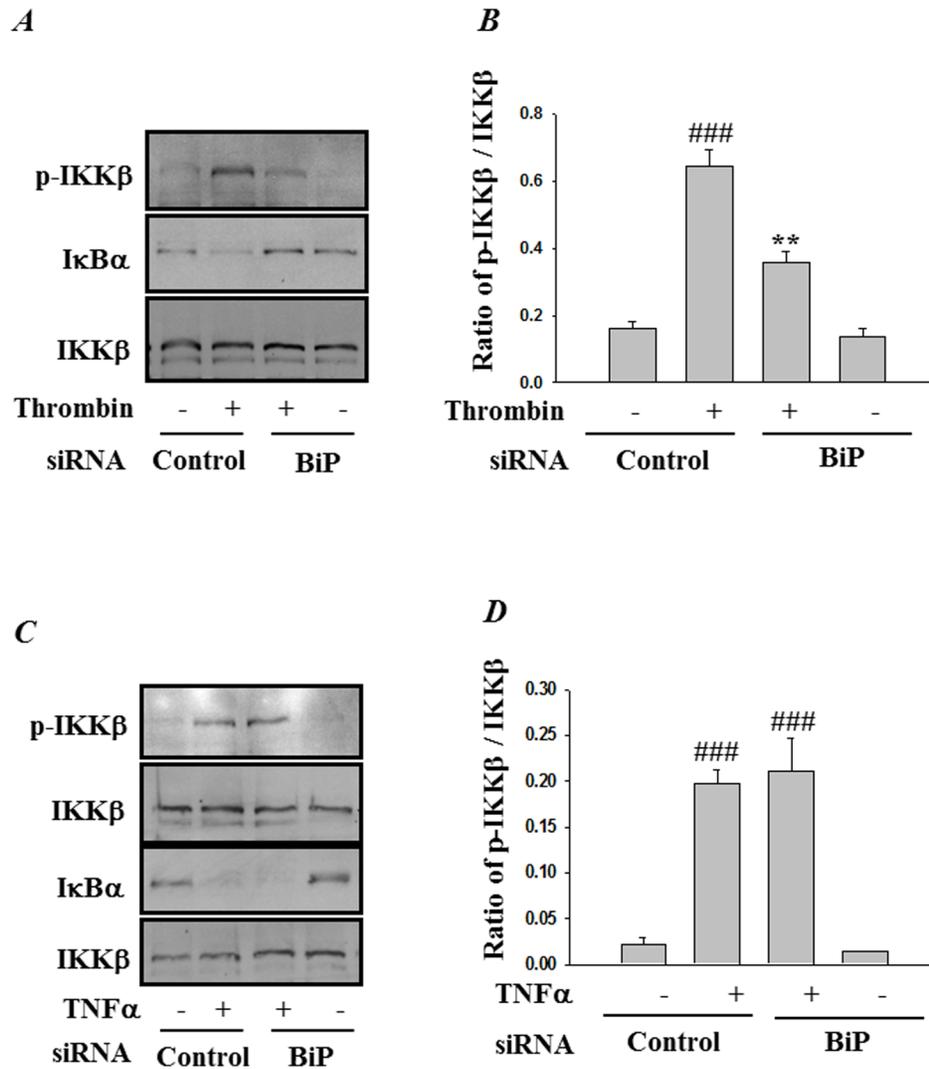


Figure 4. BiP knockdown blocks thrombin-induced IKK β phosphorylation and I κ B α degradation. HPAEC were transfected with control siRNA or BiP siRNA using DharmaFect1. After 24–36 h, the cells were challenged for 1 h with (A) thrombin (5 U/ml) or 0.5 h with (C) TNF α (100 U/ml). Total cell lysates were prepared and immunoblotted with anti-phospho-IKK β and anti-I κ B α to determine the phosphorylation of IKK β and degradation of I κ B α respectively. Total levels of IKK β were used to monitor loading. The bar graphs represent the effect of BiP depletion on (B) thrombin-induced or (D) TNF α -induced IKK β phosphorylation normalized to total IKK β levels. The data are the means \pm S.E. (n=3–6 for each condition). ^{###} p <0.001 difference from controls; ^{**} p <0.01 difference from thrombin stimulated controls. doi:10.1371/journal.pone.0110949.g004

DEAE-dextran in serum-free endothelial basal medium 2, and the mixture was added onto plasmid (Promega, Madison, WI) containing *Renilla* luciferase gene driven by the constitutively active thymidine kinase promoter to normalize transfection efficiencies. After 1 h, the cells were incubated for 4 min with 10% dimethyl sulfoxide in serum-free endothelial basal medium 2. The cells were then washed two times with endothelial basal medium 2, 10% fetal bovine serum and grown to confluence. We achieved transfection efficiency of 16 ± 3 (mean \pm S.D.; $n = 3$) in these cells. Cell extracts were prepared and assayed for firefly luciferase activity using the Promega Biotech dual luciferase reporter assay system. The data were expressed as a ratio of firefly luciferase activity. For experiments examining the effect of BiP knockdown on NF- κ B activity, the cells were first transfected with siRNA using DharmaFect1. After 12–16 h, the cells were again transfected with pNF- κ B-LUC using the DEAE-dextran method,

and luciferase activity was determined as described above. cells that were 60–80% confluent. We used 0.125 μ g of pTKRLUC.

Immunoblot Analysis

EC were lysed in radioimmune precipitation (RIPA) buffer containing 50 mM Tris-HCl, pH 7.4, 150 mM NaCl, 0.25 mM EDTA, pH 8.0, 1% deoxycholic acid, 1% Triton X-100, 5 mM NaF, 1 mM sodium orthovanadate supplemented with complete protease inhibitors (Sigma). The residual binding sites on the filters were blocked by incubating with 5% (w/v) nonfat dry milk in TBST (10 mM Tris, pH 8.0, 150 mM NaCl, 0.05% Tween 20) or 5% BSA in TBST for 1 h at room temperature. The membranes were subsequently incubated with the indicated antibodies and developed using an enhanced chemiluminescence (ECL) method, as described [42]. Representative blots shown in the result section come from the same membrane which may have more samples in various groups.

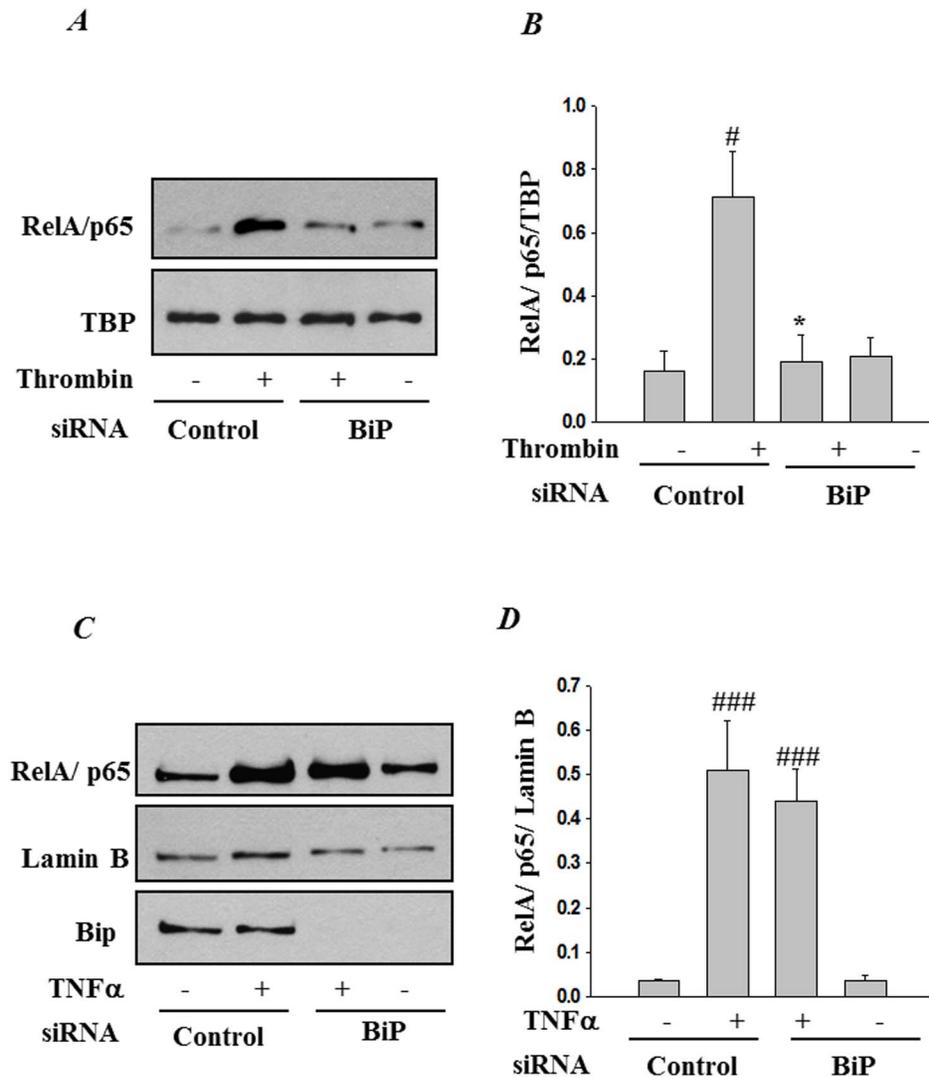


Figure 5. BiP knockdown prevents thrombin-induced RelA/p65 translocation to the nucleus. HPAEC were transfected with control-siRNA or BiP siRNA using DharmaFect1. After 24–36 h, the cells were challenged with (A) thrombin (5 U/ml) for 1 h or (C) TNF α for 0.5 h. Nuclear extracts (NE) were separated by SDS-PAGE and immunoblotted with anti-RelA/p65 antibody. Tata Binding Protein (TBP) and Lamin B, both nuclear proteins, were used as loading control for nuclear extracts. Anti-BiP antibody was used to monitor BiP depletion. The bar graph represents the effect of BiP knockdown on (B) thrombin or (D) TNF α -induced nuclear translocation of RelA/p65 normalized to TBP and Lamin B levels respectively. The data are the means \pm S.E. (n = 6 for each condition). [#] $p < 0.05$ or ^{###} $p < 0.001$ difference from controls; ^{*} $p < 0.05$ difference from thrombin stimulated controls. doi:10.1371/journal.pone.0110949.g005

ELISA

The levels of IL-8 and MCP-1 in HPAEC culture supernatants were determined using ELISA kits from R&D Systems (Minneapolis, MN) according to the manufacturer's recommendations.

Nuclear Extract Preparation and Assessment of RelA/p65 DNA Binding

After treatment, cells were washed twice with ice-cold phosphate-buffered saline and resuspended in 400 μ l of buffer A (10 mM HEPES [pH 7.9], 10 mM KCl, 0.1 mM EDTA, 0.1 mM EGTA, 1 mM [DTT], and 0.5 mM PMSF). Fifteen minutes later, NP-40 was added to a final concentration of 0.6%, and the samples were centrifuged to collect the supernatants containing the cytoplasmic proteins. The pelleted nuclei were resuspended in 50 μ l of buffer B (20 mM HEPES [pH 7.9], 0.4 M NaCl, 1 mM EDTA, 1 mM EGTA, 1 mM DTT, and 1 mM PMSF). After 0.5 h at 4°C, lysates were centrifuged and supernatants containing

the nuclear proteins were collected. The DNA binding activity of RelA/p65 was determined using an ELISA-based DNA binding assay kit (Cayman Chemical, Ann Arbor, MI) in accordance with the manufacturer's recommendations.

Immunofluorescence

Cells grown on Collagen I coated coverslips were fixed in 3.7% paraformaldehyde/PBS for 10 min at room temperature (RT) and then permeabilized with 0.1% Triton X-100 for 5 min at room temperature as described [38,43]. Permeabilized cells were rinsed 3 times with 1XPBS and incubated in blocking solution (5% normal goat serum or 1% bovine serum albumin/PBS) for 1 h at room temperature to prevent nonspecific binding of the antibody. All subsequent steps were carried out at room temperature and cells were rinsed 3 times with 1xPBS between each of the steps. To localize Filamentous-actin (F-actin), cells were incubated with Alexa Fluor 488-labeled phalloidin for 20 min at room temperature. The

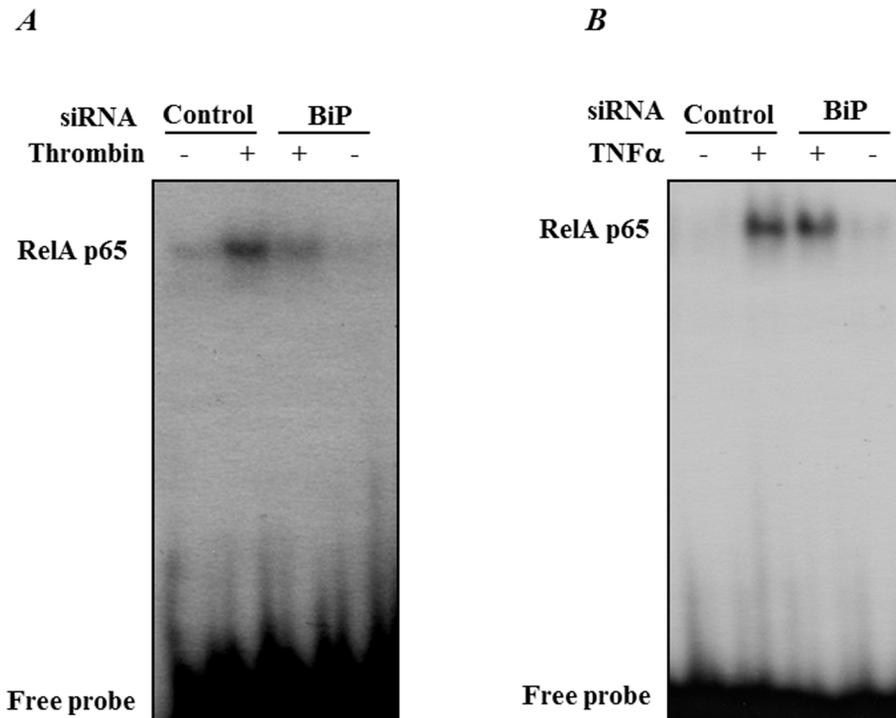


Figure 6. BiP knockdown blocks thrombin-induced RelA/p65 binding to DNA. HPAEC were transfected with control-siRNA or BiP-siRNA using DharmaFect1. After 24–36 h, the cells were challenged for 1 h with (A) thrombin or 0.5 h with (B) TNF α . Nuclear extracts were prepared and assayed for DNA binding of RelA/p65 by EMSA as described in the Materials and Methods. Results are representatives of two experiments. doi:10.1371/journal.pone.0110949.g006

coverslips were rinsed in PBS and mounted on the slide using Vectashield mounting media (Vector Laboratories, Lincolnshire, IL). Images were obtained using Nikon Eclipse - TE2000-E Fluorescent microscope and Olympus FV1000 confocal microscope.

Electrophoretic Mobility Shift Assay (EMSA)

The EMSA was performed basically as described [44]. Briefly, 10 μ g of nuclear extract was mixed with 1 μ g of poly (dI-dC) in a binding buffer (10 mM Tris-HCl [pH 7.5], 50 mM NaCl, 0.5 mM DTT, 10% glycerol (20 μ l final volume) for 15 min at room temperature. The double stranded oligonucleotides were end labeled using gamma³²P ATP. The reaction mixture was incubated with the end-labeled double-stranded oligonucleotides containing an NF- κ B site (30,000 cpm each) for 15 min at room temperature. The DNA-protein complexes were resolved on a 5% native polyacrylamide gel in low ionic strength buffer (0.25x Tris-borate-EDTA). The oligonucleotide used for the gel shift analysis was Ig- κ B 5'-GTTGAGGGGACTTTCCAGGC-3' and contains the consensus NF- κ B binding site sequence (underlined) present in mouse Ig kappa light chain gene [45].

Endothelial permeability assay

Endothelial permeability was measured using Millipore's *In vitro* Vascular Permeability Assay Kit. HPAEC transfected with control-siRNA or BiP-siRNA for 48 hours or treated with SubAB or SubA_{A272}B for 6 hours were seeded at 20,000 cells per transwell insert and cultured for 48 hours. Following this, the confluent monolayer was treated with thrombin (5 U/ml) for 30 minutes. FITC-Dextran permeability testing was done to check monolayer integrity. Permeation was stopped by removing the inserts from the wells. Media from the receiver tray was

transferred to a 96 well opaque plate to measure fluorescence. Fluorescent intensities were quantified using a fluorescent plate reader with filters appropriate for 485 nm and 535 nm excitation and emission. The endothelial monolayer was stained to check for monolayer integrity using Cell Stain provided in the kit. Cells were visualized using the Leica DMI 3000B fluorescent microscope.

Statistical Analysis

Results are presented as mean \pm SE and were analyzed by using standard one-way ANOVA. The significance between the groups was determined using Tukey's test (Prism 5.0, GraphPad Software, San Diego). A p value < 0.05 between two groups was considered statistically significant.

Results

RNAi knockdown of BiP induces eIF2 α phosphorylation and ATF4 protein expression

To understand the significance of ER stress in EC inflammation, we first determined whether siRNA-mediated knockdown of BiP induces ER stress. ER stress activates a signaling network called the unfolded protein response (UPR) to alleviate the stress and promote cell survival. Of the three distinct signaling pathways that are activated by UPR, one of the pathways involves PERK autophosphorylation and dimerization, leading to phosphorylation of eIF2 α , and concurrent increase in the expression of ATF4 that regulates the transcription of several UPR target genes that mitigate ER stress. Therefore, we investigated whether BiP knockdown induces the phosphorylation of eIF2 α and ATF4 protein levels, the two markers of ER stress. HPAEC were transfected with control-siRNA or BiP-siRNA for 48 hrs and were then subjected to Western blot analysis. Results showed that BiP

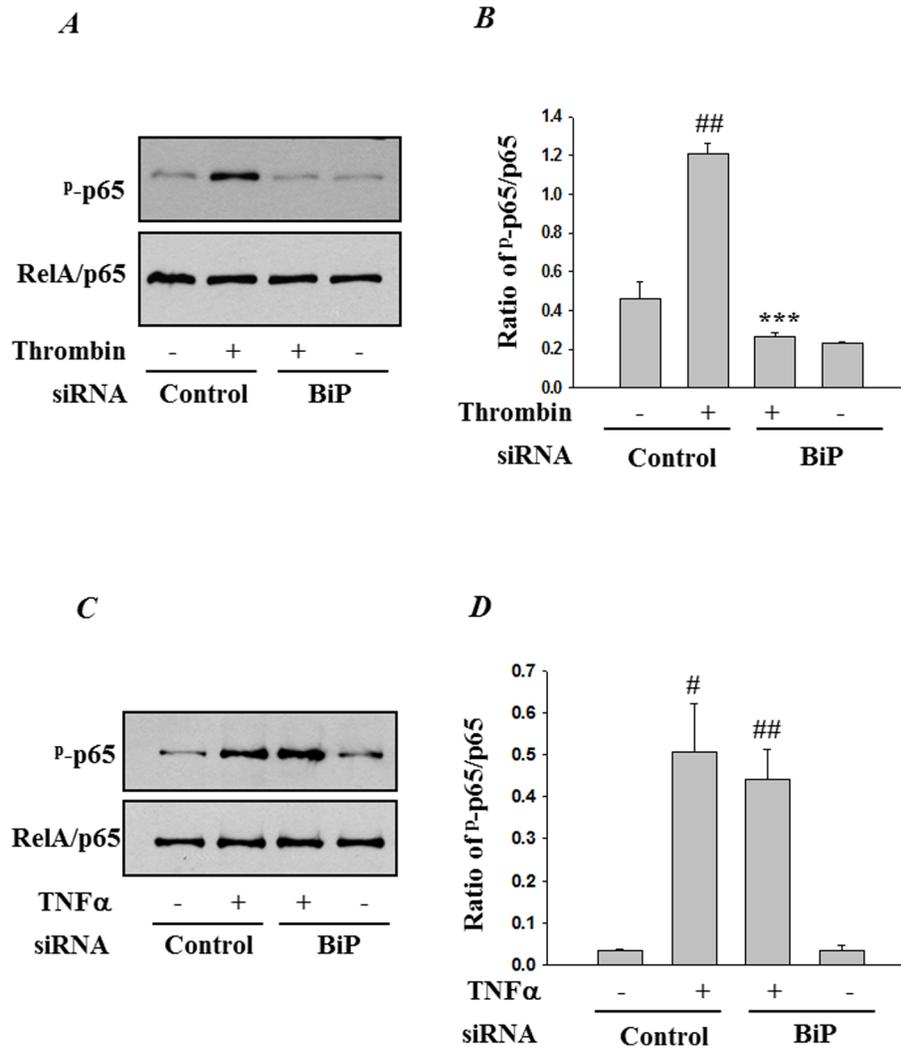


Figure 7. BiP knockdown markedly ameliorates thrombin-induced RelA/p65 phosphorylation at Ser⁵³⁶. HPAEC were transfected with control-siRNA or BiP-siRNA using DharmaFect1. After 24–36 h, the cells were challenged for 1 h with (A) thrombin or 0.5 h with (C) TNF α . Cell lysates were immunoblotted with an anti-phospho RelA/p65 (Ser⁵³⁶) to determine the phosphorylation status of RelA/p65. The levels of total RelA/p65 were used to monitor loading. The bar graph represents the effect of BiP depletion on (B) thrombin or (D) TNF α -induced phosphorylation of RelA/p65 at Ser⁵³⁶ normalized to total RelA/p65 levels. The data are the means \pm S.E. (n = 3 for each condition). [#] $p < 0.05$ or ^{##} $p < 0.01$ difference from controls; ^{***} $p < 0.001$ difference from thrombin stimulated controls. doi:10.1371/journal.pone.0110949.g007

expression was significantly inhibited in cells transfected with BiP siRNA compared to the control siRNA and that BiP-depleted cells showed a robust increase in ATF4 protein expression (Fig. 1A & B). Also, cells transfected with BiP siRNA showed a marked induction in the phosphorylation of eIF2 α (Fig. 1C & D). Tunicamycin (TM) was used as a positive control to monitor ER stress (Fig. 1A). These results indicate the induction of ER stress upon BiP knockdown.

RNAi knockdown of BiP inhibits thrombin-induced NF- κ B activity and adhesion molecules expression

Next, to determine the role of BiP-mediated ER stress in EC inflammation, we monitored the effect of BiP knockdown on the activity of NF- κ B, the master regulator of inflammation. Cells were transfected with pNF- κ B-LUC in combination with either control-siRNA or BiP-siRNA. Results showed that thrombin challenge of cells transfected with control-siRNA resulted in increased NF- κ B reporter activity and this response was inhibited

in cells transfected with BiP-siRNA (Fig. 2A). In a reciprocal approach we addressed the effect of overexpression of BiP wild-type and a BiP dominant negative mutant on thrombin-induced NF- κ B activity. Transfection of cells with a construct encoding wild-type BiP (pCMV-BiP-Myc-KDEL-WT) showed no effect on thrombin-induced NF- κ B activity. However, expression of a construct encoding a dominant negative BiP (pCMV-BiP-Myc-KDEL-T37G) significantly inhibited thrombin-induced NF- κ B activity (Fig. 2B). In view of the essential role of NF- κ B in adhesion molecule expression, we determined the effect of BiP knockdown on thrombin-induced expression of ICAM-1 and VCAM-1. We found that depletion of BiP attenuated thrombin-induced ICAM-1 and VCAM-1 expression consistent with its effect on NF- κ B activity (Fig. 3A–C). To further ascertain our observation that ER stress inhibits NF- κ B activity, cells were treated with a known inducer of ER stress, tunicamycin. Results (Figure S1) show that tunicamycin significantly inhibited thrombin-induced NF- κ B activity and adhesion molecule expression. Additionally, to

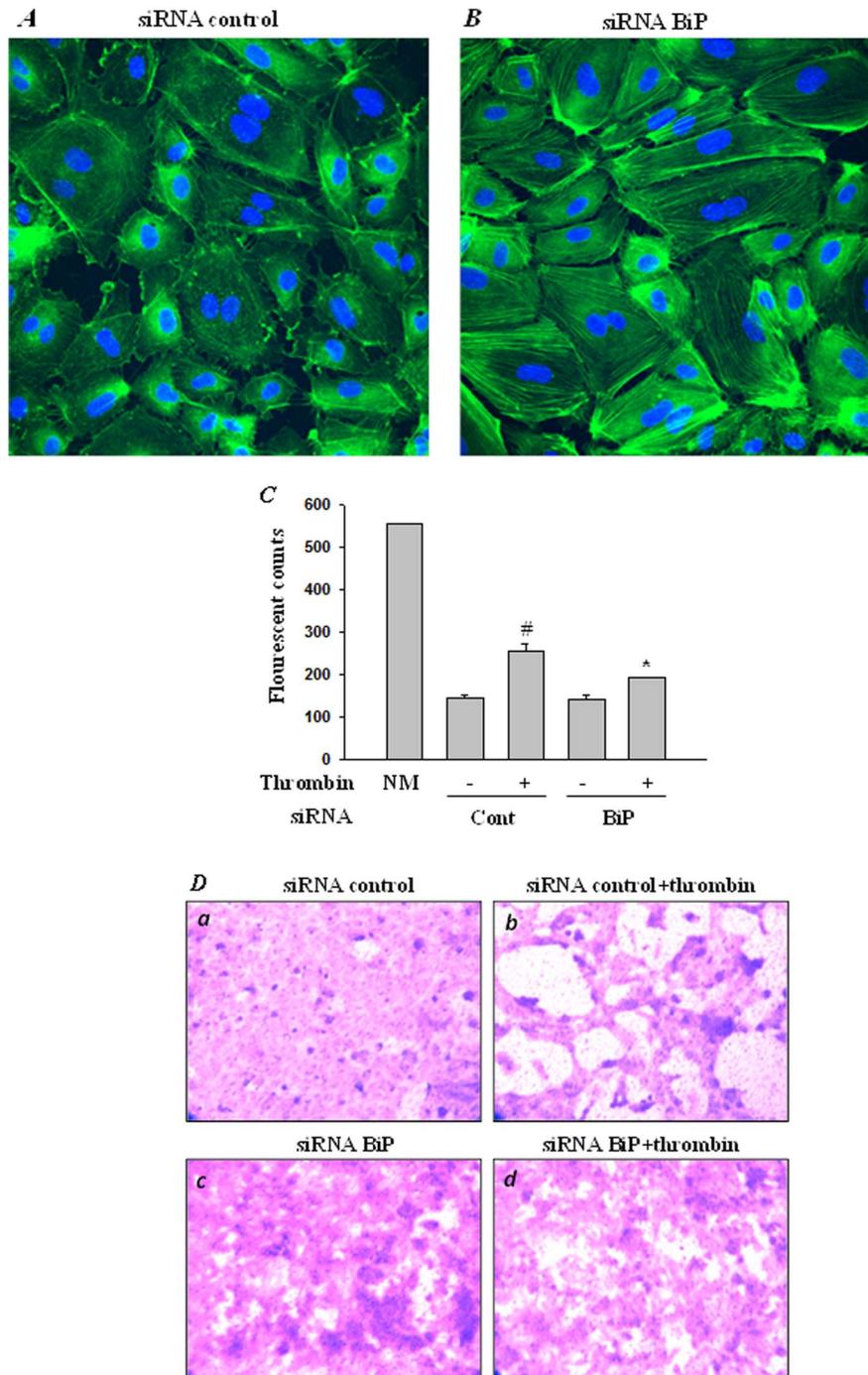
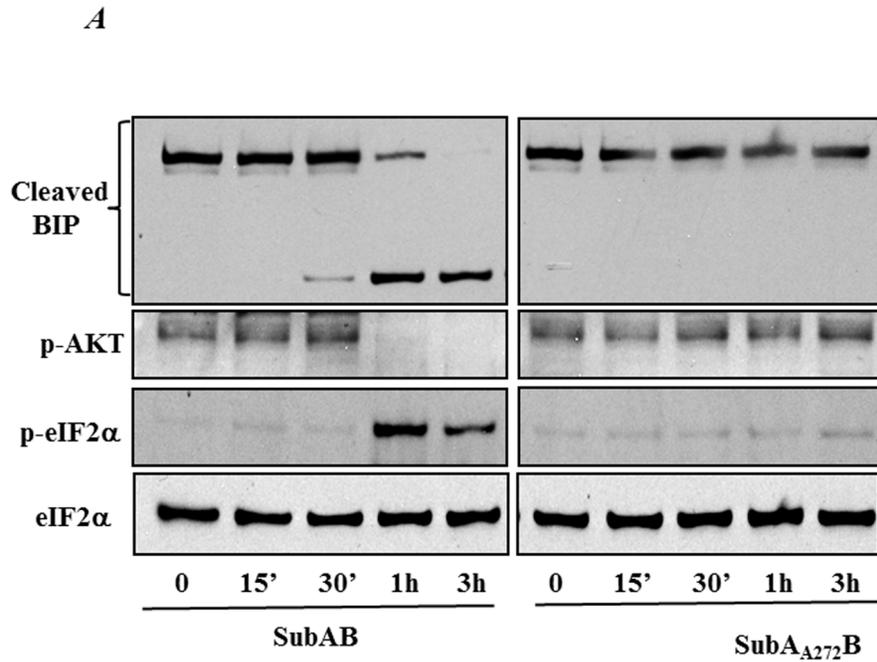


Figure 8. BiP knockdown potentiates actin filament formation and regulates thrombin-induced endothelial permeability. HPAEC were transfected with (A) control-siRNA or (B) BiP-siRNA using DharmaFect1. After 24–36 h cells were fixed, permeabilized, and stained with Alexa Fluor 488 labeled phalloidin to visualize the actin filaments. Images were analyzed by Fluorescence microscopy. Results are representative of three experiments. (C) HPAEC transfected with control-siRNA or BiP- siRNA were seeded at 20,000 cells per transwell insert and cultured for 48 hours. Following this, the confluent monolayer was treated with thrombin (5 U/ml) for 30 minutes. FITC-Dextran permeability testing was done to check monolayer integrity. Permeation was stopped by removing the inserts from the wells. Media from the receiver tray was transferred to a 96 well opaque plate to measure fluorescence. Fluorescent intensities were quantified using a fluorescent plate reader with filters appropriate for 485 nm and 535 nm excitation and emission. The data are the means \pm S.E. (n=3–6 for each condition). [#] $p < 0.05$ difference from controls; ^{*} $p < 0.05$ difference from thrombin-stimulated controls. (D) Following permeability testing the endothelial monolayer transfected with control siRNA (a & b) or with BiP siRNA (c & d), followed by treatment with thrombin (b & d) or left untreated (a & c) were stained for bright field imaging. doi:10.1371/journal.pone.0110949.g008

investigate the specificity of the above response, we determined whether knockdown of BiP affected TNF α activation of NF- κ B-induced ICAM-1 and VCAM-1 expression. Results showed that depletion of BiP (Fig. 3D–F) failed to inhibit TNF α -induced

ICAM-1 and VCAM-1 expression. Together, these results indicate that in HPAEC, BiP regulates adhesion molecules expression in a stimulus specific manner.



B

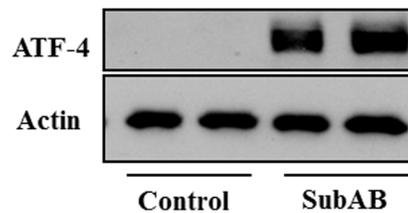


Figure 9. SubAB induces BiP cleavage and ER stress. (A) HPAEC were treated with 0.1 μ g/ml of SubAB or mutant SubA_{A272}B for the indicated time points. Cell lysates were immunoblotted with an anti-phospho eIF2 α antibody and anti-phospho-AKT antibody to determine the induction of ER stress. The levels of total eIF2 α were used to monitor loading. BiP antibody was used to monitor the cleavage of BiP by SubAB. (B) HPAECs were treated with 0.1 μ g SubAB for 3 hours and cell lysates were immunoblotted for ATF-4 antibody. Actin was used as a loading control. doi:10.1371/journal.pone.0110949.g009

RNAi knockdown of BiP interferes with IKK β activation and subsequent degradation of I κ B α

Next we analyzed the mechanism by which BiP depletion inhibits the NF- κ B signaling cascade in endothelial cells. Phosphorylation of I κ B α and its subsequent degradation is a requirement for the release and translocation of NF- κ B to the nucleus. Since phosphorylation of I κ B α is mediated by the IKK complex, we first evaluated the role BiP in IKK activation. Depletion of BiP inhibited activation of IKK upon thrombin challenge, as determined by decreased phosphorylation of IKK β at Ser^{176/181} (Fig. 4A & B). We next examined whether knockdown of BiP affects IKK β -mediated degradation of I κ B α . Results show that depletion of BiP was effective in inhibiting I κ B α degradation. In contrast, BiP depletion had no significant effect on TNF α -induced IKK β phosphorylation and consequently on I κ B α degradation (Fig. 4C & D).

RNAi knockdown of BiP inhibits thrombin-induced NF- κ B nuclear translocation and subsequent DNA binding

Since I κ B α degradation is a prerequisite for the release of NF- κ B for its nuclear translocation, we next assessed the effect of BiP depletion on NF- κ B nuclear translocation and subsequent DNA binding. Nuclear extracts from control and treated cells were analyzed by immunoblotting and electrophoretic mobility shift assay. Results showed a marked decrease in nuclear translocation (Fig. 5A & B) and DNA binding of RelA/p65 (Fig. 6A) in thrombin-treated cells transfected with BiP-siRNA, as compared to cells transfected with control-siRNA. Together these results are consistent with our previous data showing a block in I κ B α degradation upon BiP depletion. Interestingly, results show that TNF α -induced RelA/p65 nuclear translocation (Fig. 5C & D) and subsequent DNA binding (Fig. 6B) were unchanged upon knockdown of BiP.

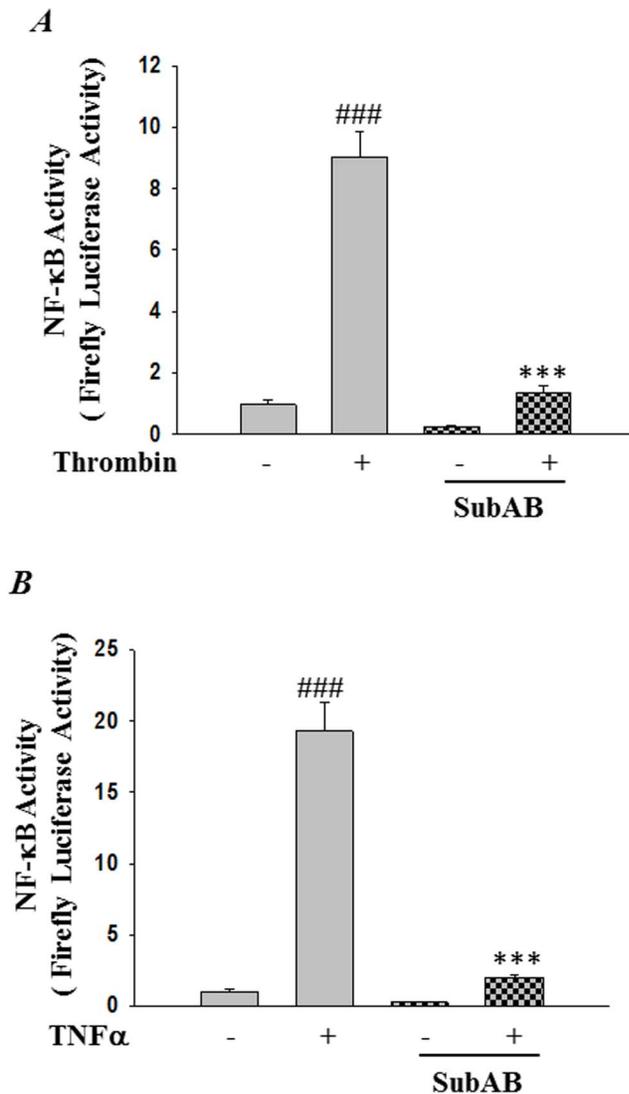


Figure 10. SubAB inhibits NF- κ B transcriptional activity. HPAEC were transfected with NF- κ BLUC construct by using DEAE-dextran as described in Materials and Methods. After 24 hours cells were treated with 0.1 μ g/ml of SubAB or mutant SubA_{A272}B for 6 h, followed by treatment with (A) thrombin (5 U/ml) or (B) TNF α (100 U/ml) for 6 h. Cell extracts were prepared and assayed for Firefly and Renilla luciferase activities. The data were expressed as a ratio of Firefly to Renilla luciferase activities. Data are means \pm SE (n=4–6 for each condition). ###*p*<0.001 difference from controls; ****p*<0.001 difference from thrombin and TNF- α stimulated controls. doi:10.1371/journal.pone.0110949.g010

RNAi knockdown of BiP regulates thrombin-induced NF- κ B phosphorylation at Ser⁵³⁶

We also determined whether BiP contributes to the transactivating potential of RelA/p65, in addition to promoting I κ B α degradation, nuclear translocation and DNA binding function. To this end, BiP-depleted cells were challenged with thrombin and total lysates were analyzed for Ser⁵³⁶ phosphorylation of RelA/p65. Depletion of BiP caused a significant inhibition in RelA/p65 phosphorylation caused by thrombin (Fig. 7A & B). As expected, TNF α -induced RelA/p65 phosphorylation remained unaffected (Fig. 7C & D), further establishing the notion that BiP mediates EC inflammation in a stimulus-specific manner.

RNAi knockdown of BiP promotes filamentous actin formation and abrogates endothelial permeability

Previous work in our lab has shown that the actin cytoskeleton is a dynamic structure that undergoes rearrangement upon exposure to thrombin. Thrombin engages actin depolymerizing protein cofilin, as well as cofilin kinase LIMK1 and cofilin phosphatase SSH-1L to regulate actin dynamics [38,40] in endothelial cells. The endothelial cell cytoskeleton is primarily composed of three structures: actin microfilaments, microtubules and intermediate filaments. It is the actin filaments that are of critical importance to EC permeability and their role is much more defined as compared to microtubules and intermediate filaments [46]. These studies prompted us to assess the role of BiP in regulating actin cytoskeletal dynamics and endothelial barrier integrity leading to vascular permeability. Controls as well as BiP-depleted cells were stained with Alexa Flour 488-labeled phalloidin to visualize actin filaments. Results show an increase in filamentous actin formation in cells depleted of BiP as compared to control cells (Fig. 8A & B). Furthermore the effect of actin cytoskeleton rearrangement on endothelial permeability was analyzed using an *in vitro* permeability assay. EC permeability is mostly regulated by intercellular junction organization. The intercellular boundaries in confluent resting cells are maintained by adherence-junction proteins such as VE-cadherin and catenin. Thrombin a serine protease, acts by cleaving its receptor PAR1 causing an increase in intracellular calcium and PKC activation, this in turn stimulates the endothelial contractile apparatus. The endothelial contraction results in disruption of VE-cadherin/catenin complex leading to intercellular gap formation. This effect of thrombin is reversible and is responsible for increased endothelial permeability [47]. To, this end HPAEC transfected with BiP-siRNA or control-siRNA were seeded at 20,000 cells per well into transwell inserts containing 1 μ m pores within a transparent polyethylene terephthalate (PET) membrane coated with type 1 rat-tail collagen, and cultured for 48 hours. The confluent monolayer was then treated with thrombin (5 U/ml) for 30 minutes, followed by addition of a high molecular weight FITC-Dextran on top of the cells. The movement of the fluorescent molecule across the endothelial monolayer is a direct determinant of monolayer permeability. Our results show that knockdown of BiP significantly reduced thrombin-induced permeability, as indicated by a marked decrease in the fluorescent counts measured in the receiver tray using a fluorescent plate reader (Fig. 8C). Next the endothelial monolayer was stained to monitor monolayer integrity using Cell stain provided in the kit. Data show that knockdown of BiP significantly reduced the gaps between untreated and thrombin treated cells, implying the role of BiP in EC integrity (Fig. 8D).

Inactivation of BiP by Subtilase Cytotoxin (SubAB) induces ER stress

In order to further ascertain the role of BiP in EC inflammation and injury we used an alternative approach. Cells were treated with SubAB, the prototype of a family of AB₅ cytotoxins produced by Shiga toxigenic *Escherichia coli* that specifically cleaves and inactivates BiP, resulting in the activation ER stress and therefore UPR [48]. A time course experiment showed that treatment of HPAEC with SubAB induced BiP cleavage at 1 hour, which coincided with the phosphorylation of eIF2 α . We also observed a decrease in AKT-phosphorylation at 1 hour of SubAB treatment. In contrast, treatment of cells with SubA_{A272}B, a nontoxic variant of SubAB, failed to cleave BiP, induce eIF2 α phosphorylation and decrease AKT-phosphorylation (Fig. 9A). As expected the level of ATF4 was also induced upon SubAB treatment (Fig. 9B). These

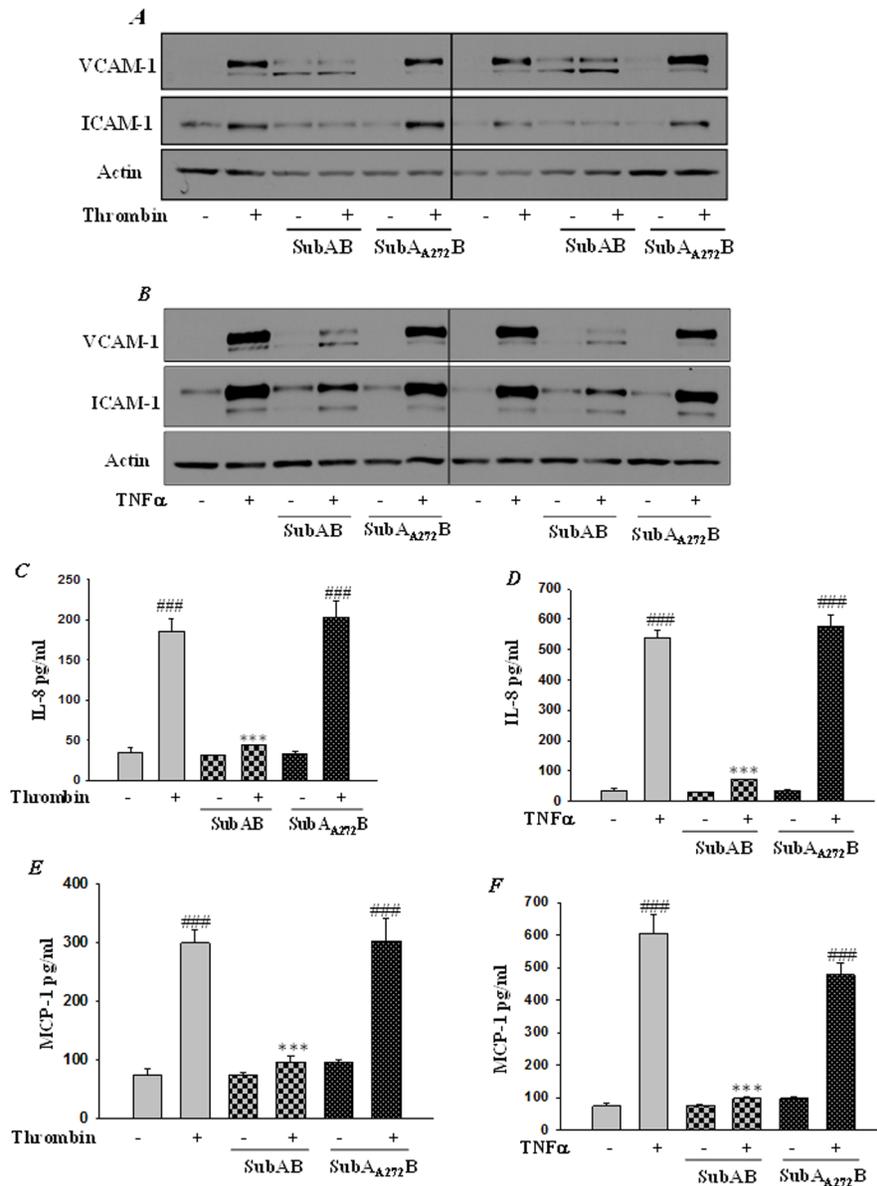


Figure 11. SubAB inhibits NF- κ B mediated proinflammatory gene expression. HPAEC were treated with 0.1 μ g/ml of SubAB or mutant SubA₂₇₂B for 6 h, followed by treatment with (A & C & E) thrombin (5 U/ml) or (B & D & F) TNF α (100 U/ml) for 6 h. Total cell lysates were immunoblotted with (A & B) anti-ICAM-1 antibody and anti-VCAM-1 antibody. Actin was used to monitor loading. The conditioned media were subjected to ELISA to determine the levels of (C & D) IL-8 or (E & F) MCP-1. Data are means \pm S.E. (n=6-9 for each condition). ###p<0.001 difference from control; ***p<0.001 difference from thrombin and TNF α -stimulated controls. doi:10.1371/journal.pone.0110949.g011

data indicate that SubAB specifically cleaves BiP and thereby induce ER stress.

Preconditioning HPAEC with SubAB-induced ER stress suppressed NF- κ B transcriptional activity and proinflammatory gene expression

Expression of adhesion molecules, cytokines and chemokines in endothelial cells by inflammatory mediators such as thrombin and TNF α is a central step in the pathogenesis of ALI and is under the tight regulation of the transcription factor NF- κ B. We determined whether SubAB-mediated ER stress preconditioning affects thrombin and TNF α -induced NF- κ B activation and subsequent expression of adhesion molecules ICAM-1, VCAM-1, and chemo-

attractant proteins IL-8 and MCP-1. Cells were transfected with pNF- κ B-LUC for 24 hours, followed by treatment with SubAB for 6 hours. Results showed that thrombin or TNF α challenge of cells resulted in increased NF- κ B reporter activity and this response was inhibited in cells pretreated with 0.1 μ g/ml of SubAB (Fig. 10A & B). Next we determined the effect of SubAB on NF- κ B target genes, ICAM-1 and VCAM-1. HPAEC were exposed to 0.1 μ g/ml SubAB and SubA₂₇₂B for 6 hours and then treated with 5 U/ml thrombin and 100 U/ml of TNF α for 6 hours. Western blot analysis using total cell lysates showed a reduced expression of both thrombin- and TNF α -induced ICAM-1 and VCAM-1 expression, in cells treated with SubAB (Fig. 11A & B). ELISA using culture supernatants showed a marked reduction in the levels of thrombin- and TNF α -induced for IL-8 and MCP-1, in cells

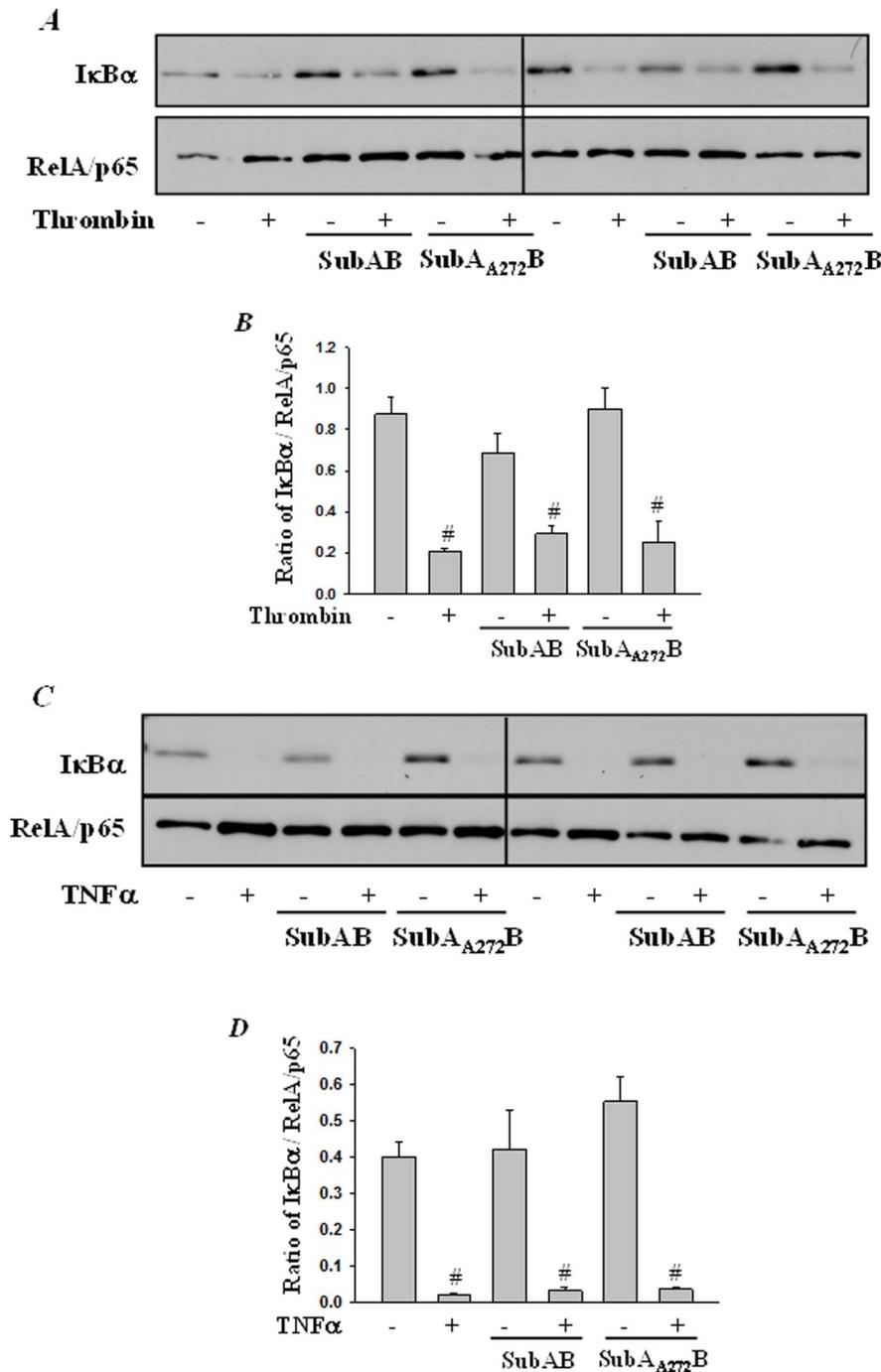


Figure 12. SubAB failed to block thrombin-induced IκBα degradation. HPAEC were treated with 0.1 μg/ml of SubAB or mutant SubA_{A272}B for 6 h, followed by treatment with (A) thrombin (5 U/ml) for 1 h or with (C) TNFα (100 U/ml) for 30 min. Total cell lysates were prepared and immunoblotted with anti-IκBα antibody to determine degradation of IκBα. Levels of RelA/p65 was used to monitor loading. The bar graphs represent the effect SubAB and SubA_{A272}B on (B) thrombin-induced or (D) TNFα-induced IκBα degradation normalized to total RelA/p65 levels. The data are the means ± S.E. (n=3–6 for each condition). #p<0.05 difference from controls.
doi:10.1371/journal.pone.0110949.g012

preconditioned with SubAB (Fig. 11C–F). In contrast, preconditioning of HPAECs to SubA_{A272}B, did not affect any of the above responses (Fig. 11C–F).

SubAB attenuates EC inflammation independent of IκBα degradation and NF-κB nuclear translocation

Next we determined whether the reduced NF-κB activity and subsequent adhesion molecule and chemokine expression following preconditioning of HPAEC with SubAB is due to reduced

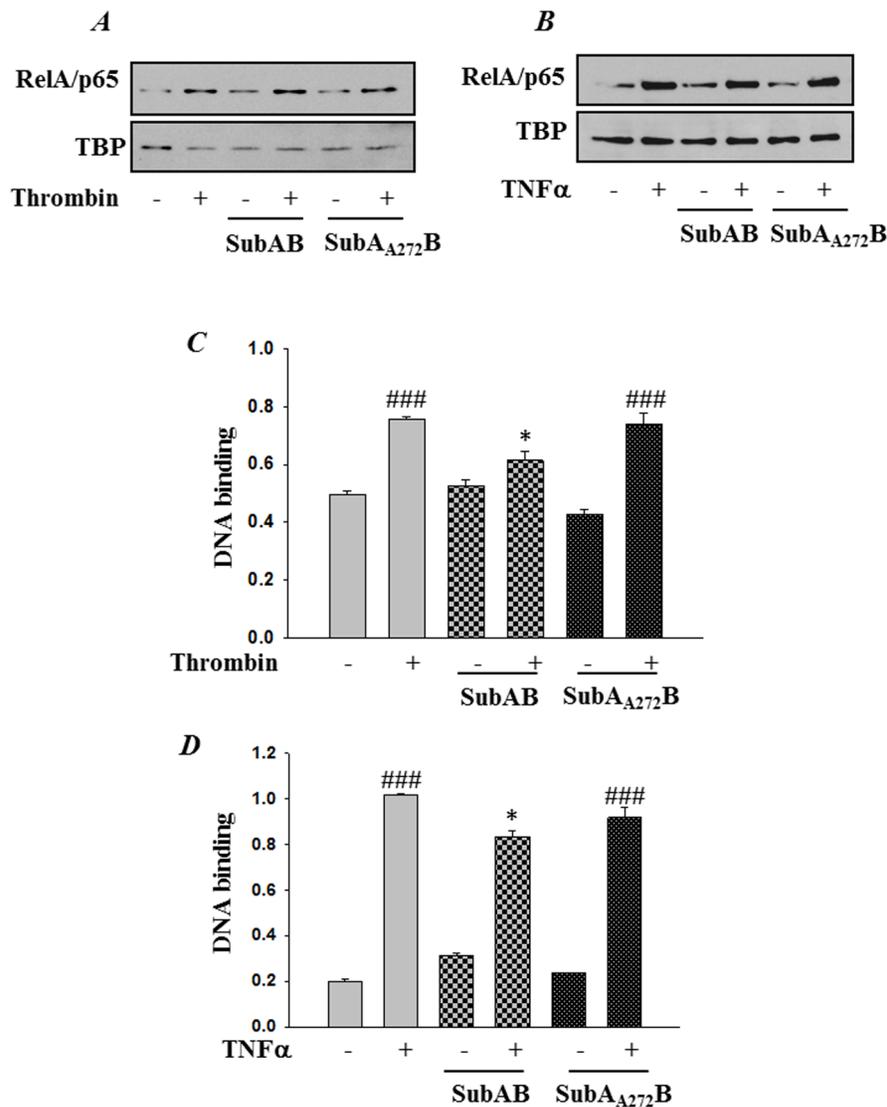


Figure 13. SubAB attenuates RelA/p65 DNA binding independent of its nuclear translocation. HPAEC were treated with 0.1 μ g/ml of SubAB or mutant SubA_{A272}B for 6 h, followed by treatment with (A & C) thrombin (5 U/ml) for 1 h or with (B & D) TNF α (100 U/ml) for 30 min. (A & B) Nuclear extracts (NE) were separated by SDS-PAGE and immunoblotted with anti-RelA/p65 antibody. Tata Binding Protein (TBP) was used as a loading control for nuclear extract. (C & D) Nuclear extracts were assayed for DNA binding of RelA/p65 by Cayman's NF- κ B (RelA/p65) Transcription Factor Assay Kit as described in Materials and Methods. The data are the means \pm S.E. (n = 6 for each condition). ###*p* < 0.001 difference from controls; **p* < 0.05 difference from thrombin and TNF- α stimulated controls. doi:10.1371/journal.pone.0110949.g013

degradation of I κ B α , the cytoplasmic inhibitor that sequesters NF- κ B in the cytosol and thereby blocks its translocation to the nucleus and consequently gene transcription. Interestingly, analysis of the cytoplasmic extracts showed that preconditioning of cells with SubAB did not inhibit thrombin or TNF α -induced I κ B α degradation (Fig. 12A–D). Furthermore, our data showed that the nuclear translocation of the liberated NF- κ B also remained unaffected upon preconditioning with ER stress (Fig. 13A & B).

Preconditioning cells with SubAB-induced ER stress suppressed NF- κ B binding to DNA

Next we analyzed whether the reduced NF- κ B activity and gene expression is due to inhibition of NF- κ B DNA binding activity. To this end we used a nonradioactive, ELISA-based assay to monitor RelA/p65 binding to DNA in nuclear extracts. Results show that both thrombin and TNF α -induced binding of NF- κ B to the DNA

was significantly inhibited in cells pretreated with the ER stress inducer SubAB (Fig. 13C & D). However the treatment of cells with SubA_{A272}B did not result in attenuation of RelA/p65 binding to DNA (Fig. 13C & D). Together these observations indicate that different ER stress inducers, RNAi-mediated knockdown of BiP versus SubAB-mediated cleavage of BiP, adopt different mechanisms to inhibit EC inflammation.

Preconditioning cells with SubAB-induced ER stress potentiated F-actin formation and abrogated endothelial permeability

Since SubAB-mediated preconditioning to ER stress mitigates thrombin- and TNF α -induced NF- κ B activation, we next analyzed the effect of SubAB on actin dynamics and endothelial permeability, another salient feature contributing to the pathogenesis of ALI and ARDS. Our results show that similar to siRNA-

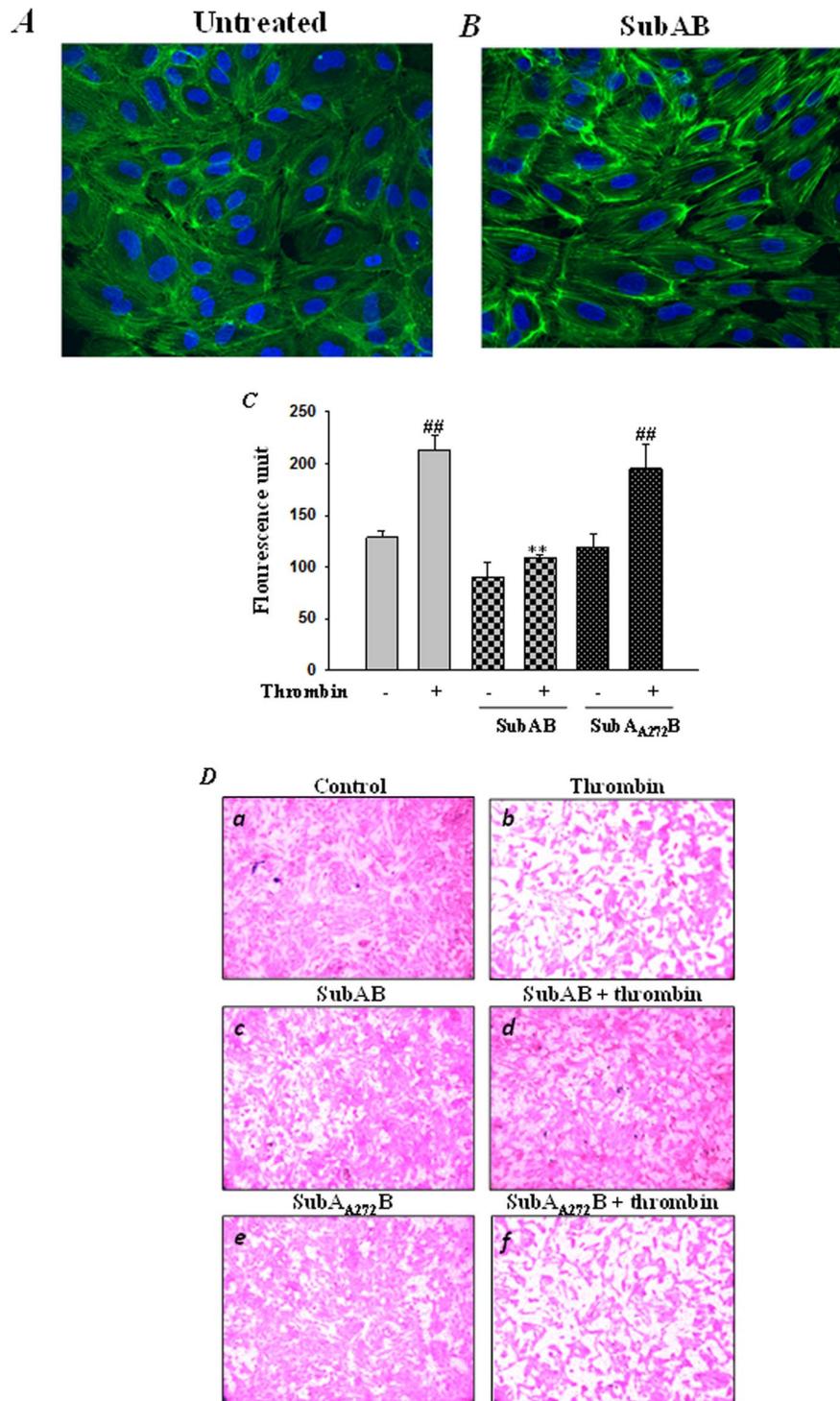


Figure 14. SubAB potentiates actin filament formation and regulates thrombin-induced endothelial permeability. HPAEC were left untreated (*A*) or (*B*) treated with 0.1 $\mu\text{g/ml}$ of SubAB for 6 hours. The cells were then fixed, permeabilized, and stained with Alexa Fluor 488 labeled phalloidin to visualize the actin filaments. Images were analyzed by Fluorescence microscopy. Results are representative of three experiments. (*C*) HPAEC treated with 0.1 $\mu\text{g/ml}$ of SubAB or mutant SubA_{A272}B for 6 h were seeded at 20,000 cells per transwell insert and cultured for 48 hours. Following this, the confluent monolayer was treated with thrombin (5 U/ml) for 30 minutes. FITC-Dextran permeability testing was done to check monolayer integrity. Permeation was stopped by removing the inserts from the wells. Media from the receiver tray was transferred to a 96 well opaque plate to measure fluorescence. Fluorescent intensities were quantified using a fluorescent plate reader with filters appropriate for 485 nm and 535 nm excitation and emission. The data are the means \pm S.E. ($n=3-6$ for each condition). ^{##} $p<0.01$ difference from controls; ^{**} $p<0.01$ difference from thrombin stimulated controls. (*D*) Following permeability testing the endothelial monolayer representing various experimental conditions (as indicated in the Figure) was stained for bright field imaging. doi:10.1371/journal.pone.0110949.g014

mediated knockdown of BiP, SubAB-mediated inactivation of BiP also resulted in alterations in actin dynamics, as evidenced by an increase in Alexa Fluor 488-labeled phalloidin staining of actin stress fibers in SubAB treated cells compared to untreated cells (Fig. 14A & B). Next, in order to analyze whether SubAB-mediated alteration in actin dynamics affects endothelial permeability, we used an in vitro vascular permeability assay. Our data showed a significant inhibition in thrombin-induced permeability in cells pretreated with SubAB, as indicated by a decrease in fluorescence units. However, cells pretreated with the inactive mutant SubA_{A272B} were unable to restore thrombin-induced endothelial permeability (Fig. 14C). In addition, monolayer integrity was visualized using Cell stain provided in the kit, under bright field microscopy. Cells pretreated with SubAB showed tightening of thrombin-induced intercellular gaps as compared to cells pretreated with the inactive SubAB mutant (Fig. 14D). Together these results imply that BiP is a central regulator of EC inflammation and injury.

Discussion

Studies have shown that BiP, a central regulator of ER stress, is critically involved in the pathogenesis of cancer [49], and inflammation associated with rheumatoid arthritis [50] and diabetic retinopathy [51]. The focus of this study is to understand the relevance of ER stress in endothelial cell inflammation and injury associated with pulmonary inflammatory diseases such as ALI and ARDS. To begin with, the present study demonstrates that depletion or inactivation of BiP per se induced ER stress, as evidenced by increase in phosphorylation of the eukaryotic initiation factor alpha (eIF2 α), and an increase in the expression of activated transcription factor 4 (ATF4), the two markers of ER stress. Next, our results indicate that preconditioning of endothelial cells with ER stress is protective against EC inflammation and injury. Mechanistic analysis revealed that the protective effect of ER stress is mediated via dampening of NF- κ B activation and restoration of endothelial barrier dysfunction.

Recent studies have shown a protective effect of ER stress preconditioning against cytokine-induced inflammation in retinal endothelial and renal mesangial cells by suppressing NF- κ B-mediated adhesion molecule expression [25,28]. Also, pretreatment of rats with subnephritogenic doses of ER stress inducers tunicamycin or thapsigargin ameliorated mesangioproliferative glomerulonephritis [25]. These studies suggest a protective role of signaling pathways activated by ER stress, against inflammatory conditions, but the mechanisms are largely unknown. Our data show that siRNA mediated knockdown of BiP inhibited EC inflammation in a stimulus specific manner. Endothelial inflammation induced by thrombin, whose levels are elevated in the BALF of patients suffering from ALI [52,53], is abolished upon BiP depletion as noted by inhibition of IKK β -mediated phosphorylation and degradation of I κ B α , an event responsible for the release and subsequent activation of NF- κ B. Downstream NF- κ B signaling events involving its cytoplasmic trafficking, phosphorylation at Ser⁵³⁶, DNA binding and resulting gene expression were consequently abrogated. Surprisingly, TNF α -induced activation of NF- κ B and EC inflammation was not hindered upon BiP knockdown in HPAEC, indicating a stimulus-specific regulation of endothelial NF- κ B by BiP. Interestingly, Zhang *et al.* show that tunicamycin-induced ER stress preconditioning abolished TNF α -elicited NF- κ B activation and adhesion molecule expression via activation of XBP-1 in primary retinal microvascular endothelial cells (HREC) [28]. To understand the discrepancy we evaluated the role of XBP-1 in TNF α induced signaling in HPAEC by knocking down XBP-1 expression

using siRNA. Results show that depletion of XBP-1 did not inhibit TNF α -induced adhesion molecule expression in HPAEC (data not shown). This disparity in cellular response can either be attributed to different ER stress inducers i.e., BiP-siRNA versus tunicamycin, or to endothelial heterogeneity i.e., HPAEC versus HREC. Studies have shown that endothelial cells from different size vessels or from same vessel but different sites show significant heterogeneity in structure and function [54].

In order to further substantiate our data we used an unrelated methodological approach. HPAEC were pretreated with sub toxic dose of SubAB, a serine protease characterized by a conserved catalytic triad Asp-His-Ser₂₇₂, which selectively cleaves BiP between a dileucine motif (Leu_{416/417}), resulting in disruption of BiP function and thereby induction of ER stress [55]. An inactive mutant SubA_{A272B}, in which Ser₂₇₂ was mutated to an alanine, was used as a negative control. Our results show that unlike BiP knockdown-mediated abrogation of EC inflammation via inhibition of I κ B α degradation, SubAB repressed EC inflammation by blocking the binding of the released NF- κ B to the DNA and subsequent adhesion molecule, chemokine and cytokine expression. The events upstream of DNA binding such as I κ B α degradation and nuclear translocation remain unaffected. As expected, the inactive SubA_{A272B} failed to inhibit the above responses. These observations indicate that the two approaches resulted in a similar outcome; inhibition of EC inflammation, however via different mechanism, which can be biologically informative and therapeutically relevant. Earlier studies have shown that the inhibition of Aurora Kinases and PI3K using siRNA-mediated gene silencing or pharmacological inhibitor-mediated inactivation resulted in phenotypes that were not consistent [56]. Kitamura *et al.*, have also shown a protective effect of SubAB on TNF α -induced inflammation in rat epithelial cells via induction of C/EBP β [27]. SubAB has also been shown to protect mice against LPS-induced experimental arthritis [57]. Furthermore, our studies show that BiP plays a central role in EC injury via regulation of endothelial barrier integrity. Preconditioning EC with ER stress by knockdown or inactivation of BiP significantly blocked thrombin-induced endothelial permeability.

In summary our studies demonstrate that the dual mechanism by which BiP regulates EC inflammation involves the activation of the NF- κ B pathway and disruption of endothelial barrier integrity. Thus, the specific targeting of BiP may be a strategy for dampening inflammatory responses associated with intravascular coagulation and sepsis in mice.

Supporting Information

Figure S1 (A) Tunicamycin attenuates thrombin-induced NF- κ B reporter activity. HPAEC were transfected with NF- κ B-LUC and Renilla luciferase construct by using DEAE-dextran as described in Materials and Methods. Cells were then treated with 0.5 μ g/ml tunicamycin for 30 minutes followed by challenge with thrombin (5 U/ml) for 6 hours. Cell extracts were prepared and assayed for firefly and Renilla luciferase activities. The data were expressed as a ratio of firefly to Renilla luciferase activities. Data are means \pm SE (n=4-6 for each condition). ###*p*<0.001 difference from controls; ****p*<0.001 difference from thrombin stimulated controls. **(B) Tunicamycin inhibits thrombin-induced adhesion molecule expression.** HPAEC were treated with 0.5 μ g/ml tunicamycin for 30 minutes followed by challenge with thrombin (5 U/ml) for 6 hours. Total cell lysates were immunoblotted with an anti-ICAM-1, anti VCAM-1, and anti-BiP antibody. Actin was used to monitor loading. The bar graphs represent the effect of tunicamycin on

thrombin-induced (**C**) ICAM-1 and (**D**) VCAM-1 expression normalized to actin level. The data are the means \pm S.E. (n = 6 for each condition). ## $p < 0.01$ or ### $p < 0.001$ difference from controls; *** $p < 0.001$ difference from thrombin stimulated controls. (TIF)

References

- Xu C, Bailly-Maitre B, Reed JC (2005) Endoplasmic reticulum stress: cell life and death decisions. *J Clin Invest* 115: 2656–2664.
- Lin JH, Walter P, Yen TS (2008) Endoplasmic reticulum stress in disease pathogenesis. *Annu Rev Pathol* 3: 399–425.
- Kim I, Xu W, Reed JC (2008) Cell death and endoplasmic reticulum stress: disease relevance and therapeutic opportunities. *Nat Rev Drug Discov* 7: 1013–1030.
- Walter P, Ron D (2011) The unfolded protein response: from stress pathway to homeostatic regulation. *Science* 334: 1081–1086.
- Kaufman RJ (1999) Stress signaling from the lumen of the endoplasmic reticulum: coordination of gene transcriptional and translational controls. *Genes Dev* 13: 1211–1233.
- Rath E, Haller D (2011) Inflammation and cellular stress: a mechanistic link between immune-mediated and metabolically driven pathologies. *Eur J Nutr* 50: 219–233.
- Kitamura M (2009) Biphasic, bidirectional regulation of NF-kappaB by endoplasmic reticulum stress. *Antioxid Redox Signal* 11: 2353–2364.
- Dudek J, Benedix J, Cappel S, Greiner M, Jalal C, et al. (2009) Functions and pathologies of BiP and its interaction partners. *Cell Mol Life Sci* 66: 1556–1569.
- Hotamisligil GS (2006) Inflammation and metabolic disorders. *Nature* 444: 860–867.
- Hotamisligil GS (2010) Endoplasmic reticulum stress and the inflammatory basis of metabolic disease. *Cell* 140: 900–917.
- Ware LB, Matthay MA (2000) The acute respiratory distress syndrome. *N Engl J Med* 342: 1334–1349.
- Levitt JE, Matthay MA (2006) Treatment of acute lung injury: historical perspective and potential future therapies. *Semin Respir Crit Care Med* 27: 426–437.
- Idell S (2003) Coagulation, fibrinolysis, and fibrin deposition in acute lung injury. *Crit Care Med* 31: S213–220.
- van der Poll T (2008) Tissue factor as an initiator of coagulation and inflammation in the lung. *Crit Care* 12 Suppl 6: S3.
- Finigan JH (2009) The coagulation system and pulmonary endothelial function in acute lung injury. *Microvasc Res* 77: 35–38.
- Ben-Neriah Y, Karin M (2011) Inflammation meets cancer, with NF-kappaB as the matchmaker. *Nat Immunol* 12: 715–723.
- Medzhitov R (2010) Inflammation 2010: new adventures of an old flame. *Cell* 140: 771–776.
- Bair AM, Thippogowda PB, Freichel M, Cheng N, Ye RD, et al. (2009) Ca²⁺ entry via TRPC channels is necessary for thrombin-induced NF-kappaB activation in endothelial cells through AMP-activated protein kinase and protein kinase Cdelta. *J Biol Chem* 284: 563–574.
- Rahman A, Anwar KN, True AL, Malik AB (1999) Thrombin-induced p65 homodimer binding to downstream NF-kappa B site of the promoter mediates endothelial ICAM-1 expression and neutrophil adhesion. *J Immunol* 162: 5466–5476.
- Shenderov K, Riteau N, Yip R, Mayer-Barber KD, Oland S, et al. (2014) Cutting edge: Endoplasmic reticulum stress licenses macrophages to produce mature IL-1beta in response to TLR4 stimulation through a caspase-8- and TRIF-dependent pathway. *J Immunol* 192: 2029–2033.
- Prell T, Lautenschlager J, Weidemann L, Ruhmer J, Witte OW, et al. (2014) Endoplasmic reticulum stress is accompanied by activation of NF-kappaB in amyotrophic lateral sclerosis. *J Neuroimmunol* 270: 29–36.
- Tam AB, Mercado EL, Hoffmann A, Niwa M (2012) ER stress activates NF-kappaB by integrating functions of basal IKK activity, IRE1 and PERK. *PLoS One* 7: e45078.
- Adachi T, Teramachi M, Yasuda H, Kamiya T, Hara H (2012) Contribution of p38 MAPK, NF-kappaB and glucocorticoid signaling pathways to ER stress-induced increase in retinal endothelial permeability. *Arch Biochem Biophys* 520: 30–35.
- Hayakawa K, Nakajima S, Hiramatsu N, Okamura M, Huang T, et al. (2010) ER stress depresses NF-kappaB activation in mesangial cells through preferential induction of C/EBP beta. *J Am Soc Nephrol* 21: 73–81.
- Inagi R, Kumagai T, Nishi H, Kawakami T, Miyata T, et al. (2008) Preconditioning with endoplasmic reticulum stress ameliorates mesangioproliferative glomerulonephritis. *J Am Soc Nephrol* 19: 915–922.
- Usuki F, Fujimura M, Yamashita A (2013) Endoplasmic reticulum stress preconditioning attenuates methylmercury-induced cellular damage by inducing favorable stress responses. *Sci Rep* 3: 2346.
- Nakajima S, Hiramatsu N, Hayakawa K, Saito Y, Kato H, et al. (2011) Selective abrogation of BiP/GRP78 blunts activation of NF-kappaB through the ATF6 branch of the UPR: involvement of C/EBPbeta and mTOR-dependent dephosphorylation of Akt. *Mol Cell Biol* 31: 1710–1718.
- Li J, Wang JJ, Zhang SX (2011) Preconditioning with endoplasmic reticulum stress mitigates retinal endothelial inflammation via activation of X-box binding protein 1. *J Biol Chem* 286: 4912–4921.
- Hung CC, Ichimura T, Stevens JL, Bonventre JV (2003) Protection of renal epithelial cells against oxidative injury by endoplasmic reticulum stress preconditioning is mediated by ERK1/2 activation. *J Biol Chem* 278: 29317–29326.
- Hayakawa K, Hiramatsu N, Okamura M, Yamazaki H, Nakajima S, et al. (2009) Acquisition of anergy to proinflammatory cytokines in nonimmune cells through endoplasmic reticulum stress response: a mechanism for subsidence of inflammation. *J Immunol* 182: 1182–1191.
- Anthoni C, Russell J, Wood KC, Stokes KY, Vowinkel T, et al. (2007) Tissue factor: a mediator of inflammatory cell recruitment, tissue injury, and thrombus formation in experimental colitis. *J Exp Med* 204: 1595–1601.
- Contrino J, Hair G, Kreutzer DL, Rickles FR (1996) In situ detection of tissue factor in vascular endothelial cells: correlation with the malignant phenotype of human breast disease. *Nat Med* 2: 209–215.
- Sabit R, Thomas P, Shale DJ, Collins P, Linnane SJ (2010) The effects of hypoxia on markers of coagulation and systemic inflammation in patients with COPD. *Chest* 138: 47–51.
- Undas A, Gissel M, Kwasny-Krochin B, Glusko P, Mann KG, et al. (2010) Thrombin generation in rheumatoid arthritis: dependence on plasma factor composition. *Thromb Haemost* 104: 224–230.
- Vaidyula VR, Criner GJ, Grabianowski C, Rao AK (2009) Circulating tissue factor procoagulant activity is elevated in stable moderate to severe chronic obstructive pulmonary disease. *Thromb Res* 124: 259–261.
- Paton AW, Beddoe T, Thorpe CM, Whisstock JC, Wilce MC, et al. (2006) AB5 subtilase cytotoxin inactivates the endoplasmic reticulum chaperone BiP. *Nature* 443: 548–552.
- Paton AW, Sriraman P, Talbot UM, Wang H, Paton JC (2004) A new family of potent AB(5) cytotoxins produced by Shiga toxicogenic Escherichia coli. *J Exp Med* 200: 35–46.
- Fazal F, Minhajuddin M, Bijli KM, McGrath JL, Rahman A (2007) Evidence for actin cytoskeleton-dependent and -independent pathways for RelA/p65 nuclear translocation in endothelial cells. *J Biol Chem* 282: 3940–3950.
- Bijli KM, Fazal F, Rahman A (2012) Regulation of RelA/p65 and endothelial cell inflammation by proline-rich tyrosine kinase 2. *Am J Respir Cell Mol Biol* 47: 660–668.
- Fazal F, Bijli KM, Minhajuddin M, Rein T, Finkelstein JN, et al. (2009) Essential role of cofilin-1 in regulating thrombin-induced RelA/p65 nuclear translocation and intercellular adhesion molecule 1 (ICAM-1) expression in endothelial cells. *J Biol Chem* 284: 21047–21056.
- Rahman A, Anwar KN, Uddin S, Xu N, Ye RD, et al. (2001) Protein kinase C-delta regulates thrombin-induced ICAM-1 gene expression in endothelial cells via activation of p38 mitogen-activated protein kinase. *Mol Cell Biol* 21: 5554–5565.
- Minhajuddin M, Fazal F, Bijli KM, Amin MR, Rahman A (2005) Inhibition of mammalian target of rapamycin potentiates thrombin-induced intercellular adhesion molecule-1 expression by accelerating and stabilizing NF-kappa B activation in endothelial cells. *J Immunol* 174: 5823–5829.
- Fazal F, Gu L, Ilnatovych I, Han Y, Hu W, et al. (2005) Inhibiting myosin light chain kinase induces apoptosis in vitro and in vivo. *Mol Cell Biol* 25: 6259–6266.
- Ledeber HC, Parks TP (1995) Transcriptional regulation of the intercellular adhesion molecule-1 gene by inflammatory cytokines in human endothelial cells. Essential roles of a variant NF-kappa B site and p65 homodimers. *J Biol Chem* 270: 933–943.
- Sen R, Baltimore D (1986) Inducibility of kappa immunoglobulin enhancer-binding protein NF-kappa B by a posttranslational mechanism. *Cell* 47: 921–928.
- Dudek SM, Garcia JG (2001) Cytoskeletal regulation of pulmonary vascular permeability. *J Appl Physiol* (1985) 91: 1487–1500.
- Rabiet MJ, Plantier JL, Rival Y, Genoux Y, Lampugnani MG, et al. (1996) Thrombin-induced increase in endothelial permeability is associated with changes in cell-to-cell junction organization. *Arterioscler Thromb Vasc Biol* 16: 488–496.
- Wolfson JJ, May KL, Thorpe CM, Jandhyala DM, Paton JC, et al. (2008) Subtilase cytotoxin activates PERK, IRE1 and ATF6 endoplasmic reticulum stress-signaling pathways. *Cell Microbiol* 10: 1775–1786.
- Dong D, Stapleton C, Luo B, Xiong S, Ye W, et al. (2011) A critical role for GRP78/BiP in the tumor microenvironment for neovascularization during tumor growth and metastasis. *Cancer Res* 71: 2848–2857.

Author Contributions

Conceived and designed the experiments: FF. Performed the experiments: AL ME. Analyzed the data: FF. Contributed reagents/materials/analysis tools: JP AP. Contributed to the writing of the manuscript: FF.

50. Yoo SA, You S, Yoon HJ, Kim DH, Kim HS, et al. (2012) A novel pathogenic role of the ER chaperone GRP78/BiP in rheumatoid arthritis. *J Exp Med* 209: 871–886.
51. Li J, Wang JJ, Yu Q, Wang M, Zhang SX (2009) Endoplasmic reticulum stress is implicated in retinal inflammation and diabetic retinopathy. *FEBS Lett* 583: 1521–1527.
52. Gando S, Nanzaki S, Morimoto Y, Kobayashi S, Kemmotsu O (1999) Systemic activation of tissue-factor dependent coagulation pathway in evolving acute respiratory distress syndrome in patients with trauma and sepsis. *J Trauma* 47: 719–723.
53. Gunther A, Mosavi P, Heinemann S, Ruppert C, Muth H, et al. (2000) Alveolar fibrin formation caused by enhanced procoagulant and depressed fibrinolytic capacities in severe pneumonia. Comparison with the acute respiratory distress syndrome. *Am J Respir Crit Care Med* 161: 454–462.
54. Aird WC (2007) Phenotypic heterogeneity of the endothelium: I. Structure, function, and mechanisms. *Circ Res* 100: 158–173.
55. Hu CC, Dougan SK, Winter SV, Paton AW, Paton JC, et al. (2009) Subtilase cytotoxin cleaves newly synthesized BiP and blocks antibody secretion in B lymphocytes. *J Exp Med* 206: 2429–2440.
56. Weiss WA, Taylor SS, Shokat KM (2007) Recognizing and exploiting differences between RNAi and small-molecule inhibitors. *Nat Chem Biol* 3: 739–744.
57. Harama D, Koyama K, Mukai M, Shimokawa N, Miyata M, et al. (2009) A subcytotoxic dose of subtilase cytotoxin prevents lipopolysaccharide-induced inflammatory responses, depending on its capacity to induce the unfolded protein response. *J Immunol* 183: 1368–1374.